

**510(k) SUBSTANTIAL EQUIVALENCE DETERMINATION
DECISION SUMMARY
ASSAY AND INSTRUMENT **COMBINATION** TEMPLATE**

A. 510(k) Number:

K162274

B. Purpose for Submission:

To obtain a substantial equivalence determination for the Solana[®] Strep Complete Assay from throat specimens.

C. Measurand:

Streptococcus pyogenes (Group A β -hemolytic Streptococcus) and *Streptococcus dysgalactiae* (pyogenic Group C and G β -hemolytic Streptococcus nucleic acids).

D. Type of Test:

The Solana Strep Complete Assay is a helicase-dependent amplification (HDA) *in vitro* diagnostic test for the rapid and qualitative detection and differentiation of *Streptococcus pyogenes* (Group A β -hemolytic Streptococcus) and *Streptococcus dysgalactiae* (pyogenic Group C and G β -hemolytic Streptococcus) nucleic acids isolated from throat swab specimens obtained from symptomatic patients.

E. Applicant:

Quidel Corporation

F. Proprietary and Established Names:

Solana Strep Complete Assay

G. Regulatory Information:

1. Regulation section:

21 CFR 866.2680 - *Streptococcus spp.* nucleic acid-based assay

2. Classification:

Class II

3. Product code:

PGX - Group C and G Beta-Hemolytic *Streptococcus* Nucleic Acid Amplification System

4. Panel:

83- Microbiology

H. Intended Use:

1. Intended use(s):

The Solana Strep Complete Assay is a rapid in vitro diagnostic test, using isothermal amplification technology (helicase-dependent amplification, HDA), for the qualitative detection and differentiation of *Streptococcus pyogenes* (Group A β -hemolytic Streptococcus) and *Streptococcus dysgalactiae* (pyogenic Group C and G β -hemolytic Streptococcus) nucleic acids isolated from throat swab specimens obtained from patients with signs and symptoms of pharyngitis, such as sore throat. The Solana Strep Complete Assay is intended for use only with the the Solana instrument.

2. Indication(s) for use:

Same as intended use.

3. Special conditions for use statement(s):

For *in vitro* diagnostic use only.

For prescription use only.

4. Special instrument requirements:

Solana™ Instrument

I. Device Description:

The Solana Strep Complete assay detects *Streptococcus pyogenes* (GAS; β -hemolytic Group A Streptococci) and *Streptococcus dysgalactiae* DNA isolated from throat swab specimens obtained from patients with the signs and symptoms of pharyngitis. The assay consists of three major steps: 1) sample processing 2) specimen preparation, and 3) amplification and detection of target sequences specific to GAS and *Streptococcus dysgalactiae*, respectively, using isothermal Helicase-Dependent Amplification (HDA) in the presence of target-specific fluorescence probe.

Patient specimen on a throat swab is transferred to a Lysis Tube and subjected to heat treatment at 95°C for 5 minutes. The heat-treated sample is added to a Dilution Tube, and

then transferred to a Reaction Tube. The Reaction Tube contains lyophilized HDA reagents, dNTPs, primers and probes. Once rehydrated with the diluted sample, the Reaction Tube is placed in the Solana for amplification and detection of GAS- and *Streptococcus dysgalactiae* - specific target sequences. Target sequences are amplified by analyte-specific primers and detected using fluorescence probes included in each Reaction Tube. Two competitive process controls (PRC) are included in the Lysis Tube to monitor sample processing, inhibitory substances in clinical samples, reagent failure or device failure. The PRC target is amplified by target-specific primers and detected by a PRC specific fluorescence probe.

Solana instrument platform measures and interprets the fluorescent signal, using on-board method-specific algorithms. Solana instrument will then report the test results to the user on the display screen. Results can also be printed out. The total time for assay completion from specimen processing to result is ≤ 25 minutes.

J. Substantial Equivalence Information:

1. Predicate device name(s):

Lyra Direct Strep Assay

2. Predicate 510(k) number(s):

k133883

3. Comparison with predicate:

Similarities		
Item	Device	Predicate
	Solana Strep Complete Assay (K162274)	Lyra Direct Strep Assay (K133883)
Intended Use	<p>The Solana Strep Complete Assay is a rapid in vitro diagnostic test, using isothermal amplification technology (helicase-dependent amplification, HDA), for the qualitative detection and differentiation of <i>Streptococcus pyogenes</i> (Group A β-hemolytic Streptococcus) and <i>Streptococcus dysgalactiae</i> (pyogenic Group C and G β-hemolytic Streptococcus) nucleic acids isolated from throat swab specimens obtained from patients with signs and symptoms of pharyngitis, such as sore throat. The Solana Strep Complete Assay is intended for use only with the the Solana instrument.</p>	<p>The Lyra Direct Strep Assay is a Real-Time PCR <i>in vitro</i> diagnostic test for the qualitative detection and differentiation of Group A β-hemolytic Streptococcus (<i>Streptococcus pyogenes</i>) and pyogenic Group C and G β-hemolytic Streptococcus nucleic acids isolated from throat swab specimens obtained from patients with signs and symptoms of pharyngitis, such as a sore throat. The assay does not differentiate between pyogenic Groups C and G β-hemolytic Streptococcus.</p> <p>All negative test results should be confirmed by bacterial culture, because negative results do not preclude Group A, C or G Strep infection and should not be used as the sole basis for treatment.</p> <p>The assay is intended for use in hospital, reference, or state laboratory settings. The device is not intended for point-of-care use.</p>
Specimen Type	Throat swab specimens	Same
Specimen Lysis	Manual, heat-based	Same
Detection Technique	Automatically detects fluorescence after dissociation of fluorophore from quencher during amplification	Same

Differences		
Item	Device	Predicate
	Solana Strep Complete Assay (K162274)	Lyra Direct Strep Assay (k133883)
DNA Amplification Technology	Isothermal Helicase-Dependent Amplification (HDA)	Real-Time Polymerase Chain Reaction (RT-PCR)
GAS Target Sequence Detected	78 base pair (bp) sequence <i>S. pyogenes</i> genome, resident in the DNase B (<i>sdaB</i>) gene 67 bp sequence Protein G gene in the <i>S. dysgalactiae</i> (pyogenic Group C and G β -hemolytic Streptococcus) genome	99 base pair (bp) sequence in the putative competence (<i>comX1.1</i>) gene in the <i>S. pyogenes</i> genome 188bp sequence in the tagatose-6-phosphate kinase (<i>lacC</i>) gene conserved in Group C and G streptococcal genomes
Testing Time	25 minutes	60-70 minutes
Instrument	Solana™	ABI 7500 Fast DX Thermocycler
Reagents/ Components	Dry heating blocks, Dilution Buffer, Lysis Buffer, Reaction Tubes	Lyra™ Direct Strep Master Mix, Process Buffer, and Rehydration Solution ABI 7500 Fast Dx 96-well PCR Plate, optical plate films and plate centrifuge Dry heating block
Performance Characteristics	GAS* Sensitivity: 98.9%[95% CI: 98.3% - 99.2%] GAS Specificity: 99.5%[95% CI: 99.1% - 99.7%] Pyo GCS/GGS* <i>S. dysgalactiae</i> Sensitivity: 100%[95% CI: 95.3% - 100%] Specificity: 99.5%[95% CI: 99.1% - 99.7%]	GAS* Sensitivity: 96.5%[95% CI: 91.3% - 98.6%] GAS Specificity: 98.0%[95% CI: 97.0% - 98.6%] Pyo GCS/GGS* Sensitivity: 95.7%[95% CI: 88.1% - 98.5%] Pyo GCS/GGS Specificity: 98.3%[95% CI: 97.4% - 98.9%]

*GAS = Group A Streptococcus; Pyo GCS/GGS = Pyogenic Group C/G Streptococcus

K. Standard/Guidance Document Referenced (if applicable):

Not applicable.

L. Test Principle:

Patient specimen on a throat swab is transferred to a Lysis Tube and subjected to heat treatment at 95°C for 5 minutes. The heat-treated sample is added to a Dilution Tube, and then transferred to two Reaction Tubes; one for GAS and one for *S. dysgalactiae*. Each Reaction Tube contains lyophilized helicase-dependent amplification (HDA) reagents, dNTPs, primers and probes for GAS or *S. dysgalactiae*. Once rehydrated with the diluted sample, the Reaction Tube is placed in Solana for amplification and detection of GAS- and *S. dysgalactiae*-specific target sequences. Target sequences are amplified by analyte-specific primers and detected using fluorescence probes included in each Reaction Tube. Two competitive process controls (PRC) are included in the Lysis Tube to monitor sample processing, inhibitory substances in clinical samples, reagent failure or device failure. The PRC target is amplified by target-specific primers and detected by a PRC specific fluorescence probe.

The target and PRC probes are labeled with a quencher on one end and a fluorophore on the other end (see **Table 1** for associated fluorophores).

Table 1: Solana Strep Complete Probe Labels	
Detectant	Fluorescence Dye/Channel
GAS or <i>S. dysgalactiae</i> Probe	FAM
Process Controls (PRC)	CY5
Reference (differentiates GAS from <i>S. dysgalactiae</i> reaction mix)	ROX

The target and PRC fluorescent probes have a RNA linker that is cleaved by RNaseH2 upon annealing to GAS, *S. dysgalactiae* or PRC amplicons, and the fluorescence signal increases due to separation of fluorophore from quencher. The Solana Instrument measures and interprets the fluorescent signal, using on-board method-specific algorithms. Solana instrument will then report the test results to the user on its display screen, and it can print out the results via a printer.

M. Performance Characteristics:

1. Analytical performance:

a. *Precision/Reproducibility:*

Reproducibility Study

The between-laboratory reproducibility of the Solana Strep Complete assay was evaluated at three sites (two external clinical sites and one in-house) using a panel of four contrived specimens:

- 1) Negative (N)
- 2) High Negative (HN), 0.3 x LoD
- 3) Low Positive (LP), 1 x LoD and

4) Moderate Positive (MP), 3.0 x LoD

Contrived specimens were made by diluting frozen bacterial stocks of GAS and *S. dysgalactiae* to the concentration noted above. The LoD values were based on the values obtained in the LoD study described in section M.1.d below. See section M.2.b below for details on the matrix equivalency and freeze-thaw study.

Three replicates of each panel member for each analyte and the controls were processed and tested on the Solana Strep Complete assay at each site by two operators for five non-consecutive days using three instruments (2 operators x 3 replicates x 5 days x 3 sites = 90 results per concentration for each analyte). The LoD values were based on the values obtained in the LoD study. The reproducibility study results are acceptable. The results are shown in the **Table 2**

Table 2: Solana Strep Complete Inter-Laboratory Reproducibility									
Analyte	Panel ID	Site 1		Site 2		Site 3		Combined	
		Detected Pos/Total	% Pos	Detected Pos/Total	% Pos	Detected Pos/Total	% Pos	Detected Pos/Total	% Pos
GAS (ATCC 19615)	HN	13/30	43.3%	10/30	33.3%	13/30	43.3%	36/90	40.0%
	LP	30/30	100.0%	30/30	100.0%	30/30	100.0%	90/90	100.0%
	MP	30/30	100.0%	30/30	100.0%	30/30	100.0%	90/90	100.0%
	N	0/30	0.0%	0/30	0.0%	0/30	0.0%	0/90	0.0%
<i>S. dysgalactiae</i> subsp <i>equisimilis</i> (ATCC 10009)	HN	10/30	33.3%	6/30	20.0%	5/30	16.7%	21/90	23.3%
	LP	30/30	100.0%	30/30	100.0%	30/30	100.0%	90/90	100.0%
	MP	30/30	100.0%	30/30	100.0%	30/30	100.0%	90/90	100.0%
	N	0/30	0.0%	0/30	0.0%	0/30	0.0%	0/90	0.0%

These results met the pre-defined acceptance criteria for the various panels and are described in labeling.

Precision Study

Within-laboratory, inter-instrument precision was evaluated for the Solana Strep Complete Assay using the same panel of contrived specimens as in the reproducibility study above.

The study was conducted over 12 days at one site (in-house) with two operators (2 operators x 12 days x 3 replicates = 72 results per panel member). One operator used one instrument and the other used two instruments. The results are shown in **Table 3**.

Table 3: Precision							
Analyte	Panel ID	Operator 1		Operator 2		Combined	
		Detected Pos/Total	% Pos	Detected Pos/Total	% Pos	Detected Pos/Total	% Pos
GAS (ATCC 19615)	HN	16/36	44.4%	15/36	41.7%	31/72	43.1%
	LP	36/36	100.0%	36/36	100.0%	72/72	100.0%
	MP	36/36	100.0%	36/36	100.0%	72/72	100.0%
	N	0/36	0.0%	0/36	0.0%	0/72	0.0%
<i>S. dysgalactiae</i> subsp <i>equisimilis</i> (ATCC 10009)	HN	6/36	16.7%	14/36	38.9%	20/72	27.8%
	LP	36/36	100.0%	36/36	100.0%	72/72	100.0%
	MP	36/36	100.0%	36/36	100.0%	72/72	100.0%
	N	0/36	0.0%	0/36	0.0%	0/72	0.0%

The precision study results met the pre-defined acceptance criteria for LP, MP, and PC specimens. The results of this study are acceptable and these results are described in labeling.

b. Linearity/assay reportable range:

Not applicable.

c. Traceability, Stability, Expected values (controls, calibrators, or methods):

Solana Strep Complete assay incorporates process controls (PRC) in the lysis buffer tube that is used to monitor sample processing and evaluate the presence of substances inhibitory to helicase dependent amplification (HDA) and to confirm the integrity of assay reagents. The Quidel Molecular A + G Streptococci Control Set #M111, which contains positive and negative controls, serves as an external processing and extraction controls for the Solana Strep Complete assay and were run each day of testing.

Studies were performed to determine the stability of specimens collected using the following routinely used swab systems: nylon, rayon and polyester swabs in Amies media, and rayon and polyester swabs in Stuart media, and rayon swabs in Amies gel. Freshly grown stocks of GAS or *S. dysgalactiae* of known titer were used to spike the swabs listed above. Triplicate testing for each analyte with each condition with each collection/transport system listed above demonstrated that specimens can be stored at 25°C ± 2°C for 2 days and then at 2 to 8 °C for up to 6 more days prior to being tested in the Solana Strep Complete assay. A separate study was performed where the spiked samples were stored at ≤-15°C or ≤-70°C for a minimum of 32 days before testing.

d. *Detection limit:*

The limit of detection (LoD) of the Solana Strep Complete assay was determined using contrived stocks of two GAS strains, one strain of Group C *S. dysgalactiae* subsp. *equisimilis* and one strain of Group G *S. dysgalactiae* subsp. *equisimilis*. For each strain, stocks were prepared by suspending and diluting freshly grown colonies on to 5% sheep blood tryptic soy agar. Colony counts from preliminary serial dilutions were associated with optical density to determine the LoD. The LoD was defined as the point at which at least 95% of all replicates tested positive (C_{95}). The GAS strains were serially diluted then spiked onto swabs. For each strain, the LoD was confirmed by testing 20 replicates for each of three dilutions at 0.3 x LoD, 1 x LoD and 3 x LoD. Each dilution for each strain was confirmed by plating on 5% sheep blood tryptic soy agar, incubating and counting colonies. The LoD study results are shown in **Table 4**.

Table 4: LoD for GAS and <i>Streptococcus dysgalactiae</i> Groups C and G		
Strain	Strain ID	CFU/ml
Group A Streptococcal strain 1 (<i>Streptococcus pyogenes</i>)	ATCC 19615	1.5×10^4
Group A Streptococcal strain 2 (<i>Streptococcus pyogenes</i>)	ATCC 12344	8.5×10^4
Pyogenic Group G Streptococcal (<i>Streptococcus dysgalactiae</i> subsp. <i>equisimilis</i>)	ATCC 12394	5.7×10^5
Pyogenic Group C Streptococcal (<i>Streptococcus dysgalactiae</i> subsp. <i>equisimilis</i>)	ATCC 10009	7.1×10^4

Based these studies, the highest observed LoD for *Streptococcus pyogenes* and *Streptococcus dysgalactiae* using the Solana Strep Complete Assay is 8.5×10^4 CFU/mL and 7.1×10^5 CFU/mL, respectively. These LoD values are described in the labeling and they were used in determining the spiking levels for all subsequent analytical studies.

e. *Analytical reactivity:*

Inclusivity studies were conducted with seven GAS strains, 14 Group C *S. dysgalactiae* strains and 11 Group G *S. dysgalactiae* strains (in addition to the four strains from the LoD studies above) against three different reagent lots with three instruments. Swabs were prepared as described in M.1.d above and run with the Solana Strep Complete assay at the concentrations described in **Table 5**. The inclusivity study results and the final organism concentrations tested are shown in **Table 5**.

Table 5: Group A β -hemolytic Streptococcus Inclusivity			
Strain #	ATCC/ CCUG** #	Lancfield Type*	Concentration Detected CFU/ml
1	ATCC BAA-595	GAS	8.5×10^4
2	ATCC 12384		
3	ATCC 49399		
4	ATCC 700294		
5	CCUG 33409		
6	CCUG 39158		
7	CCUG 53553		
1	ATCC 9542	GCS	7.1×10^4
2	ATCC 12388		
3	ATCC 35666		
4	CCUG 1483		
5	CCUG 6713		
6	CCUG 27479		
7	CCUG 27480		
8	CCUG 27658		
9	CCUG 27659		
10	CCUG 27664		
11	CCUG 28115		
12	CCUG 28116		
13	CCUG 28238		
14	CCUG 48477		
1	ATCC 6644	GGS	7.1×10^4
2	CCUG 502		
3	CCUG 27482		
4	CCUG 27483		
5	CCUG 33802		
6	CCUG 15679		
7	CCUG 15680		
8	CCUG 21557		
9	CCUG 24070		
10	CCUG 26147		
11	CCUG 27477		

*All GCS and GGS tested were *S. disgalactiae* subsp. *equisimilis* or *S. disgalactiae* subsp. *dysgalactiae*

**ATCC: American Type Culture Collection; CCUG: Culture Collection, University of Göteborg

Three results were obtained for each of the strains tested. Results showed that each strain was detected 100% of the time. These study results are acceptable and they are described in the labeling.

f. Analytical specificity:

i. Microbial cross-reactivity

An *in silico* BLAST analysis of primers used in the Solana Complete Strep assay against the NCBI database against 60 potential interfering virus/organisms did not show evidence of cross-reactivity.

The Solana Complete Strep assay was tested in the presence of 45 other microorganisms commonly found in throat specimens. Each potentially interfering microorganism was tested in the presence of 2 x LoD of each of the GAS and *S. dysgalactiae* organisms used in the LoD studies in M.1.d above, in the presence of clinically relevant levels of viruses (e.g., 10⁵pfu/ml) and bacteria/yeast (e.g., 10⁶cfu/mL). All strain combinations were spiked into contrived negative matrix. The strains included in the cross-reactivity study are shown in **Table 6**.

Table 6: Strains Included in Cross-Reactivity		
Strain		
<i>Acinetobacter lwoffii</i>	<i>Legionella jordanis</i>	<i>Streptococcus intermedius</i>
<i>Arcanobacterium haemolyticum</i>	<i>Legionella micdadei</i>	<i>Stenotrophomonas maltophilia</i>
<i>Bacillus cereus</i>	<i>Moraxella cartarrhalis</i>	<i>Streptococcus mitis</i>
<i>Bordetella pertussis</i>	<i>Neisseria gonorrhoeae</i>	<i>Streptococcus mutans</i>
<i>Burkholderia cepacia</i>	<i>Neisseria subflava</i>	<i>Streptococcus oralis</i>
<i>Candida albicans</i>	<i>Peptostreptococcus micros</i>	<i>Streptococcus pneumoniae</i>
<i>Corynebacterium diphtheria</i>	<i>Pseudomonas aeruginosa</i>	<i>Streptococcus salivarius</i>
<i>Enterococcus faecalis</i>	<i>Serratia marcescens</i>	<i>Streptococcus sanguinis</i>
<i>Escherichia coli</i>	<i>Staphylococcus aureus</i> MRSA	<i>Streptococcus suis</i>
<i>Fusobacterium necrophorum</i>	<i>Staphylococcus epidermidis</i>	Adenovirus Type 1
<i>Haemophilus influenzae</i> type A	<i>Streptococcus agalactiae</i>	Adenovirus Type 11 (Slobitski)
<i>Klebsiella pneumonia</i>	<i>Streptococcus anginosus</i>	Parainfluenza Type 4B (VR-1377)
<i>Lactobacillus acidophilus</i>	<i>Streptococcus bovis</i>	Influenza A
<i>Lactococcus lactis</i>	<i>Streptococcus canis</i>	Influenza B
<i>Legionella pneumophila</i>	<i>Streptococcus gordonii</i> (Virdans type)	Rhinovirus Type 15 (1734)

Of the 45 microorganisms tested that might be found in throat specimens, *Klebsiella pneumoniae*, *Serratia marcescens* and *Enterococcus faecalis* each cross-reacted once out of six times tested (triplicate testing was repeated for each cross-reactive strain) with the Sonala Strep Complete assay.

These study results are acceptable and they are described in the labeling.

ii. Microbial interference

Microbial interference studies were conducted in triplicate with each of the 45 bacteria, yeast and viruses listed above in **Table 6** in triplicate in the presence of clinically relevant levels of viruses (e.g., 10^5 PFU/ml) and bacteria/yeast (e.g., 10^6 CFU/ml) or higher. All of the strains used in the LoD studies in M.1.d above strains were prepared as described in the cross-reactivity studies in M.1.f.i above along with the spiked onto the swab at a 2 x LoD concentration. These studies were conducted using three instruments. None of the bacteria, yeast or viruses demonstrated interference with the detection of the Group A Streptococcal strains in any of the replicates tested.

These study results are acceptable and they are described in the labeling.

iii. Interfering substances

28 chemical and biological substances were evaluated for potential to interfere with the Solana Strep Complete assay, including blood (5% v/v) and human saliva (10% v/v). Each substance was tested in triplicate using the strains used in the LoD studies in M.1.d above at 2 x LoD at medically relevant concentrations. None of the substances tested were found to interfere with the Solana Strep Complete assay in any of the replicates tested.

These study results are acceptable and they are described in the labeling.

iv. Carryover/Cross-contamination

A study to assess carryover contamination was conducted by three operators using three different Solana Instruments with one GAS strain (ATCC 19615) and one *S. dysgalactiae* subsp *equisimilis* strain (ATCC 10009). Fifty high positive contrived specimens for each analyte (at ~2000 x LoD) and 50 negative contrived specimens were spiked onto swabs and tested with the Solana Strep Complete assay. Five positive and five negative swabs were tested in an alternating order. No carryover/cross contamination was observed in the course of this study.

These study results are acceptable and they are described in the labeling.

g. Assay cut-off:

Cut-off values are based on set time thresholds for amplification and were determined based on results from preliminary LoD studies and initial clinical specimen testing. The cut-offs were set based on the longest time observed to achieve a valid GAS and *S. dysgalactiae* results from these studies.

2. Comparison studies:

a. *Method comparison with predicate device:*

Not applicable.

b. *Matrix comparison:*

A comparison study was conducted between negative clinical matrix and a contrived negative matrix in order to validate the use of the contrived negative matrix in place of a clinical negative matrix for use in the analytical studies (described in section M1 above).

Contrived negative matrix consisted of liquid Amies transport medium, Porcine Gastric Mucin (PGM), Phosphate Buffered Saline (PBS), Bovine Serum Albumin and sodium azide. The matrix comparison study results are shown in **Table 7**.

Table 7: Matrix Comparison Study					
Panel ID		Contrived Negative Matrix		Pooled Negative Clinical Matrix	
		Detected	% Pos	Detected	% Pos
GAS (ATCC 19615)	1 x LoD	20/20	100%	20/20	100%
<i>Streptococcus dysgalactiae</i> subsp <i>equisimilis</i> (ATCC 12394)	1 x LoD	20/20	100%	20/20	100%

These studies demonstrate that the contrived negative matrix is equivalent to a clinical matrix. These study results are acceptable and support the use of contrived matrix to facilitate the conduct of analytical studies.

Fresh contrived specimens and frozen specimens were prepared as described in the LoD studies (**M.1.d**) and precision studies (**M.1.a**) respectively and compared. The results of the fresh versus frozen study are shown in **Table 8**.

Table 8: Fresh Versus Frozen Study					
Panel ID		Contrived Negative Matrix		Pooled Negative Clinical Matrix	
		Detected	% Pos	Detected	% Pos
GAS (ATCC 19615)	3 x LoD	20/20	100%	72/72	100%
	1 x LoD	20/20	100%	72/72	100%
	0.3 x LoD	12/20	60%	47/72	65%

These studies demonstrate that freezing does not appear to impact results for the Solana Strep Complete assay, in the context of the contrived analytical studies submitted in this submission. These study results are acceptable and support the use of frozen specimens in contrived matrix to facilitate the conduct of analytical studies.

3. Clinical studies:

a. *Clinical Sensitivity:*

The clinical performance of the Solana Strep Complete assay was demonstrated with 2688 prospectively collected fresh throat specimens at three sites across the United States. The assay was evaluated for the qualitative detection and differentiation of GAS and *S. dysgalactiae* using nucleic acids isolated from the patients' throat swabs. A single specimen was collected per patient. Samples were collected using Polyester or Rayon Swab and transported in liquid Amies or liquid Stuart non-nutritive medium.

All 2688 fresh throat swab specimens were processed for culture and identification of GAS, GCS and GGS at the testing sites using well-accepted microbiological methods. Cultured isolates were typed by latex agglutination. β -hemolytic isolates that were typed as Group C or G were subcultured and the species were determined using an FDA-cleared MALDI-TOF assay. Swab transport fluid was also tested using another FDA-cleared nucleic acid amplification test (NAAT) and cultured at a central reference laboratory.

A specimen was considered positive if culture from the swab, swab transport fluid or the results from another FDA cleared assay were positive for GAS and/or GCS/GGS.

Two results were determined to be invalid in these studies. The invalid rate observed during the clinical prospective study for this device was 0.07% (2/2688). The breakdown of performance by analyte based on the 2686 valid swab specimens is summarized in **Tables 9 and 10**.

Table 9: Clinical Performance Data for the Solana Strep Complete vs. Dual Composite Culture and NAAT for Group A β-hemolytic Streptococcus			
All Sites			
Solana Strep Complete	Composite Dual Culture and NAAT		
	GAS	Negative	Total
GAS	475	25	500
Negative	6	2180	2186
Total	481	2205	2686
Sensitivity: 98.8% (475/481) 95% CI (97.3%-99.4%) Specificity: 98.9% (2180/2205) 95% CI (98.3%-99.2%)			
Site 1			
Solana Strep Complete	Composite Dual Culture and NAAT		
	GAS	Negative	Total
GAS	90	4	94
Negative	2	679	681
Total	92	683	775
Sensitivity: 97.8% (90/92) 95% CI (92.4%-99.4%) Specificity: 99.4% (679/683) 95% CI (98.5%-99.8%)			
Site 2			
Solana Strep Complete	Composite Dual Culture and NAAT		
	GAS	Negative	Total
GAS	84	6	90
Negative	1	510	511
Total	85	516	601
Sensitivity: 98.8%% (84/85) 95% CI (93.6%-99.8%) Specificity: 98.8%% (510/516) 95% CI (97.5%-99.5%)			
Site 3			
Solana Strep Complete	Composite Dual Culture and NAAT		
	GAS	Negative	Total
GAS	100	3	103
Negative	3	492	495
Total	103	495	598
Sensitivity: 97.1% (100/103) 95% CI (91.8%-99.0%) Specificity: 99.4% (492/495) 95% CI (98.2%-99.8%)			

Site 4			
Solana Strep Complete	Composite Dual Culture and NAAT		
	GAS	Negative	Total
GAS	83	12	95
Negative	0	254	254
Total	83	266	349
Sensitivity: 100% (37/39) 95% CI (95.6%-100%) Specificity: 95.5% (254/266) 95% CI (92.3%-97.4%)			
Site 5			
Solana Strep Complete	Composite Dual Culture and NAAT		
	GAS	Negative	Total
GAS	118	0	118
Negative	0	245	245
Total	118	245	363
Sensitivity: 100% (118/118) 95% CI (96.8%-100%) Specificity: 100% (245/245) 95% CI (98.5%-100%)			

These study results are acceptable and they are described in the labeling.

Table 10: Clinical Performance Data for the Solana Strep Complete vs. Dual Composite Culture and NAAT for Pyogenic Group C and G β -hemolytic <i>Streptococcus dysgalactiae</i>			
All Sites			
Solana Strep Complete	Composite Dual Culture and NAAT		
	GCS/GGS	Negative	Total
<i>S. dysgalactiae</i>	78	14	92
Negative	0	2594	2594
Total	78	2608	2686
Sensitivity: 100% (78/78) 95% CI (95.3%-100%) Specificity: 99.5% (2594/2608) 95% CI (99.1%-99.7%)			

Site 1			
Solana Strep Complete	Composite Dual Culture and NAAT		
	GCS/GGS	Negative	Total
<i>S. dysgalactiae</i>	32	4	36
Negative	0	739	739
Total	32	743	775
Sensitivity: 100.0% (32/32) 95% CI (89.3%-100.0%) Specificity: 96.6% (739/743) 95% CI (98.6%-99.8%)			
Site 2			
Solana Strep Complete	Composite Dual Culture and NAAT		
	GCS/GGS	Negative	Total
<i>S. dysgalactiae</i>	16	5	21
Negative	0	580	580
Total	16	585	601
Sensitivity: 100% (16/16) 95% CI (80.6%-100%) Specificity: 99.1% (593/605) 95% CI (98.0%-99.6%)			
Site 3			
Solana Strep Complete	Composite Dual Culture and NAAT		
	GCS/GGS	Negative	Total
<i>S. dysgalactiae</i>	26	4	30
Negative	0	568	568
Total	26	572	598
Sensitivity: 100% (26/26) 95% CI (87.1%-100%) Specificity: 99.3% (568/572) 95% CI (98.2%-99.7%)			
Site 4			
Solana Strep Complete	Composite Dual Culture and NAAT		
	GCS/GGS	Negative	Total
<i>S. dysgalactiae</i>	2	0	2
Negative	0	347	347
Total	2	347	349
Sensitivity: 100% (2/2) 95% CI (34.2%-100%) Specificity: 100% (347/347) 95% CI (98.9%-100.0%)			

Site 5			
Solana Strep Complete	Composite Dual Culture and NAAT		
	GCS/GGS	Negative	Total
<i>S. dysgalactiae</i>	2	1	3
Negative	0	360	360
Total	2	361	363
Sensitivity: 100% (2/2) 95% CI (34.2%-100%)			
Specificity: 99.7% (361/360) 95% CI (98.4%-100.0%)			

All pyogenic Group C and Group G streptococcal species found during this assay belong to the species *S. dysgalactiae*. These study results are acceptable and they are described in the labeling.

The external quality control isolates used in these studies were from the Quidel Molecular A+G Control Set #M111 consisting of *Streptococcus pyogenes* Z018 (for the GAS primer set) and *Streptococcus dysgalactiae* Z068 (for the GCS/GGS primer set), which serve as processing and extraction controls. The control isolates were tested with acceptable results.

All GAS positive external controls were detected accurately (100%, 180/180) by the Solana Strep Complete assay. All *S. dysgalactiae* positive controls were detected accurately (100%, 180/180). All negative controls were not detected (0%, 0/180). There were no invalid control results.

b. Clinical specificity:

See **Tables 9 and 10** above.

c. Other clinical supportive data (when a. and b. are not applicable):

Not applicable.

4. Clinical cut-off:

Not applicable.

5. Expected values/Reference range:

The overall prevalence of GAS or GCS/GGS in patients tested during this study based on culture results alone was 16.0% (431/2686) for GAS and 2.4%(65/2686) for GCS/GGS. The overall prevalence of GAS or GCS/GGS in patients tested during this study based on a combination of culture results and another FDA-cleared NAAT assay was 17.9% (481/2686) for GAS and 2.9% (78/2686) for GCS/GGS. All clinical specimens collected during this study were collected between February, 2016 and July 2016.

N. Instrument Name:

Solana™ Instrument

O. System Descriptions:

1. Modes of Operation:

Does the applicant's device contain the ability to transmit data to a computer, webserver, or mobile device?

Yes _____ or No X

Does the applicant's device transmit data to a computer, webserver, or mobile device using wireless transmission?

Yes _____ or No X

2. Software:

FDA has reviewed applicant's Hazard Analysis and software development processes for this line of product types:

Yes X or No _____

3. Specimen Identification:

Specimens are identified by scanning a barcode or by manual entry.

4. Specimen Sampling and Handling:

Swab specimens are expressed in transport medium. 50µl of the expressed specimen are transferred to a lysis tube. After heat lysis, 50µl of lysed specimen is transferred to a reaction tube for automated amplification and detection. See section I above for more information.

5. Calibration:

The end user is not required to calibrate the instrument. Automated calibration happens by comparing between the measured magnitude of the optical signal of and an integrated calibration standard and the expected magnitude of the optical signal.

6. Quality Control:

See section M.1.c for information on internal and external controls.

See section M.3.a for information on external control performance during clinical trials.

P. Other Supportive Instrument Performance Characteristics Data Not Covered In The “Performance Characteristics” Section above:

Not applicable.

Q. Proposed Labeling:

The labeling is sufficient and it satisfies the requirements of 21 CFR Part 809.10.

R. Conclusion

The submitted information in this premarket notification is complete and supports a substantial equivalence decision.