



May 15, 2026

Roche Molecular Systems, Inc.
Deborah Leu
Regulatory Affairs Project Manager
4300 Hacienda Dr.
Pleasanton, California 94588

Re: K253756

Trade/Device Name: cobas liat CT/NG nucleic acid test

Regulation Number: 21 CFR 866.3393

Regulation Name: Device to detect nucleic acids from non-viral microorganism(s) causing sexually transmitted infections and associated resistance marker(s)

Regulatory Class: Class II

Product Code: QEP

Dated: November 25, 2025

Received: November 25, 2025

Dear Deborah Leu:

We have reviewed your section 510(k) premarket notification of intent to market the device referenced above and have determined the device is substantially equivalent (for the indications for use stated in the enclosure) to legally marketed predicate devices marketed in interstate commerce prior to May 28, 1976, the enactment date of the Medical Device Amendments, or to devices that have been reclassified in accordance with the provisions of the Federal Food, Drug, and Cosmetic Act (the Act) that do not require approval of a premarket approval application (PMA). You may, therefore, market the device, subject to the general controls provisions of the Act. Although this letter refers to your product as a device, please be aware that some cleared products may instead be combination products. The 510(k) Premarket Notification Database available at <https://www.accessdata.fda.gov/scripts/cdrh/cfdocs/cfpmn/pmn.cfm> identifies combination product submissions. The general controls provisions of the Act include requirements for annual registration, listing of devices, good manufacturing practice, labeling, and prohibitions against misbranding and adulteration. Please note: CDRH does not evaluate information related to contract liability warranties. We remind you, however, that device labeling must be truthful and not misleading.

If your device is classified (see above) into either class II (Special Controls) or class III (PMA), it may be subject to additional controls. Existing major regulations affecting your device can be found in the Code of Federal Regulations, Title 21, Parts 800 to 898. In addition, FDA may publish further announcements concerning your device in the Federal Register.

Additional information about changes that may require a new premarket notification are provided in the FDA guidance documents entitled "Deciding When to Submit a 510(k) for a Change to an Existing Device" (<https://www.fda.gov/media/99812/download>) and "Deciding When to Submit a 510(k) for a Software Change to an Existing Device" (<https://www.fda.gov/media/99785/download>).

Your device is also subject to, among other requirements, the Quality Management System Regulation (QMSR) (21 CFR Part 820), which includes, but is not limited to, ISO 13485 clause 7.3 (Design controls), ISO 13485 clause 8.3 (Nonconforming product), ISO 13485 clause 8.5.2 (Corrective action), and ISO 13485 clause 8.5.3 (Preventative action). Please note that regardless of whether a change requires premarket review, the QMSR requires device manufacturers to review and approve changes to device design and production (ISO 13485 clause 7.3 and ISO 13485 clause 7.5) and document changes and approvals in the Medical Device File (ISO 13485 clause 4.2.3).

Please be advised that FDA's issuance of a substantial equivalence determination does not mean that FDA has made a determination that your device complies with other requirements of the Act or any Federal statutes and regulations administered by other Federal agencies. You must comply with all the Act's requirements, including, but not limited to: registration and listing (21 CFR Part 807); labeling (21 CFR Part 801 and Part 809); medical device reporting (reporting of medical device-related adverse events) (21 CFR Part 803) for devices or postmarketing safety reporting (21 CFR Part 4, Subpart B) for combination products (see <https://www.fda.gov/combination-products/guidance-regulatory-information/postmarketing-safety-reporting-combination-products>); good manufacturing practice requirements as set forth in the Quality Management System Regulation (QMSR) (21 CFR Part 820) for devices or current good manufacturing practices (21 CFR Part 4, Subpart A) for combination products; and, if applicable, the electronic product radiation control provisions (Sections 531-542 of the Act); 21 CFR Parts 1000-1050.

All medical devices, including Class I and unclassified devices and combination product device constituent parts are required to be in compliance with the final Unique Device Identification System rule ("UDI Rule"). The UDI Rule requires, among other things, that a device bear a unique device identifier (UDI) on its label and package (21 CFR 801.20(a)) unless an exception or alternative applies (21 CFR 801.20(b)) and that the dates on the device label be formatted in accordance with 21 CFR 801.18. The UDI Rule (21 CFR 830.300(a) and 830.320(b)) also requires that certain information be submitted to the Global Unique Device Identification Database (GUDID) (21 CFR Part 830 Subpart E). For additional information on these requirements, please see the UDI System webpage at <https://www.fda.gov/medical-devices/device-advice-comprehensive-regulatory-assistance/unique-device-identification-system-udi-system>.

Also, please note the regulation entitled, "Misbranding by reference to premarket notification" (21 CFR 807.97). For questions regarding the reporting of adverse events under the MDR regulation (21 CFR Part 803), please go to <https://www.fda.gov/medical-devices/medical-device-safety/medical-device-reporting-mdr-how-report-medical-device-problems>.

For comprehensive regulatory information about medical devices and radiation-emitting products, including information about labeling regulations, please see Device Advice (<https://www.fda.gov/medical-devices/device-advice-comprehensive-regulatory-assistance>) and CDRH Learn (<https://www.fda.gov/training-and-continuing-education/cdrh-learn>). Additionally, you may contact the Division of Industry and Consumer Education (DICE) to ask a question about a specific regulatory topic. See the DICE website (<https://www.fda.gov/medical-devices/device-advice-comprehensive-regulatory->

[assistance/contact-us-division-industry-and-consumer-education-dice](#)) for more information or contact DICE by email (DICE@fda.hhs.gov) or phone (1-800-638-2041 or 301-796-7100).

Sincerely,

HIMANI BISHT -S

Himani Bisht
Assistant Director
Viral Respiratory and HPV Branch
Division of Microbiology Devices
OHT7: Office of In Vitro Diagnostics
Office of Product Evaluation and Quality
Center for Devices and Radiological Health

Enclosure

Indications for Use

510(k) Number (if known)

K253756

Device Name

cobas liat CT/NG nucleic acid test

Indications for Use (Describe)

The cobas liat CT/NG nucleic acid test is an automated, qualitative in vitro nucleic acid diagnostic test that utilizes real-time polymerase chain reaction (PCR) for the direct detection of *Chlamydia trachomatis* (CT) and *Neisseria gonorrhoeae* (NG) nucleic acid in male/female urine and vaginal swabs, all in cobas PCR Media (Roche Molecular Systems, Inc.). This test is intended as an aid in the diagnosis of urogenital infections in both symptomatic and asymptomatic individuals.

Type of Use (Select one or both, as applicable)

☒ Prescription Use (Part 21 CFR 801 Subpart D)

☐ Over-The-Counter Use (21 CFR 801 Subpart C)

CONTINUE ON A SEPARATE PAGE IF NEEDED.

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cobas[®] liat CT/NG nucleic acid test
510(k) Summary

This summary of 510(k) safety and effectiveness information is being submitted in accordance with the requirements of 21 CFR 807.92.

Submitter Name	Roche Molecular Systems, Inc.
Address	4300 Hacienda Drive, Pleasanton, CA 94588-2722
Contact	Deborah Leu Phone: 925-523-8362 Email: deborahleu@roche.com
Date Prepared	May 11, 2026
Proprietary Name	cobas[®] liat CT/NG nucleic acid test
Common Name	cobas[®] liat CT/NG
Classification Name	866.3393 Nucleic Acid Detection System For Non-Viral Microorganism(S) Causing Sexually Transmitted Infections
Product Codes	QEP MKZ LSL
Predicate Devices	cobas[®] 6800/8800 CT/NG
Establishment Registration	Roche Molecular Systems, Inc. (2243471)

1. DEVICE DESCRIPTION

The test is performed on the **cobas® liat** analyzer which automates and integrates sample purification, nucleic acid amplification, and detection of the target sequence in biological samples using real-time PCR assays. The assay targets both the Cryptic plasmid and 23S rRNA of *Chlamydia trachomatis* and the pivNG and NGR9 of *Neisseria gonorrhoeae*. An Internal Control (IC) is also included. The IC is present to control for adequate processing of the target bacteria through steps of sample purification, nucleic acid amplification, and to monitor the presence of inhibitors in the PCR processes.

2. INDICATIONS FOR USE

The **cobas® liat** CT/NG nucleic acid test is an automated, qualitative in vitro nucleic acid diagnostic test that utilizes real-time polymerase chain reaction (PCR) for the direct detection of *Chlamydia trachomatis* (CT) and *Neisseria gonorrhoeae* (NG) nucleic acid in male/female urine and vaginal swabs, all in **cobas®** PCR Media (Roche Molecular Systems, Inc.).

This test is intended as an aid in the diagnosis of urogenital infections in both symptomatic and asymptomatic individuals.

3. TECHNOLOGICAL CHARACTERISTICS

The primary technological characteristics and intended use of the RMS **cobas® liat** CT/NG nucleic acid test are substantially equivalent to other legally marketed nucleic acid amplification tests intended for the qualitative detection of CT and NG.

As indicated in Table 1, the RMS **cobas® liat** CT/NG nucleic acid test is substantially equivalent to significant characteristics of the identified predicate device, the currently cleared **cobas®** 6800/8800 CT/NG (K202408) for use on **cobas®** 6800/8800 Systems.

Table 1: Comparison of the cobas® liat CT/NG nucleic acid test and the Predicate Device

	Submitted Device: cobas® liat CT/NG nucleic acid test	Predicate Device: cobas® 6800/8800 CT/NG for use on cobas® 6800/8800 Systems.
Regulation Name	866.3393	866.3393
Product Code	QEP MKZ LSL	QEP MKZ LSL

	Submitted Device: cobas® liat CT/NG nucleic acid test	Predicate Device: cobas® 6800/8800 CT/NG for use on cobas® 6800/8800 Systems.
Intended Use	The cobas® liat CT/NG nucleic acid test is an automated, qualitative in vitro nucleic acid diagnostic test that utilizes real-time polymerase chain reaction (PCR) for the direct detection of Chlamydia trachomatis (CT) and Neisseria gonorrhoeae (NG) nucleic acid in male/female urine and vaginal swabs, all in cobas® PCR Media (Roche Molecular Systems, Inc.). This test is intended as an aid in the diagnosis of urogenital infections in both symptomatic and asymptomatic individuals.	The cobas CT/NG for use on the cobas 6800/8800 system is an automated, qualitative in vitro nucleic acid diagnostic test, that utilizes real-time polymerase chain reaction (PCR), for the direct detection of Chlamydia trachomatis (CT) and/or Neisseria gonorrhoeae (NG) DNA in male and female urine, clinician-instructed self-collected vaginal swab specimens (collected in a clinical setting), and clinician-collected vaginal swab specimens, endocervical swab specimens, oropharyngeal (throat) swab specimens and anorectal swab specimens all collected in cobas PCR Media (Roche Molecular Systems, Inc.), and cervical specimens collected in PreservCyt solution. This test is intended as an aid in the diagnosis of chlamydial and gonococcal disease in both symptomatic and asymptomatic individuals.
Sample Type	Male and female urine, vaginal swabs	Male and female urine, Self-collected/clinician-collected vaginal swab specimens in cobas® PCR Media, Endocervical swab specimens in cobas® PCR Media, Cervical specimens in PreservCyt® solution. Oropharyngeal (throat) swab Anorectal swab specimens
Analyte Targets	Chlamydia trachomatis (CT) Neisseria gonorrhoeae (NG)	Chlamydia trachomatis (CT), Neisseria gonorrhoeae (NG)
Ancillary Collection Kits	cobas® PCR Urine Sample Kit cobas® PCR Media Uni Swab Sample Kit	cobas® PCR Media Dual Swab Sample Kit cobas® PCR Media Uni Swab Sample Kit cobas® PCR Urine Sample Kit
Sample Preparation	Automated	Same
Amplification Technology	Real-time PCR	Same
Detection Chemistry	Assay using different reporter dyes for target and control	Paired reporter and quencher fluorescence labeled probes (TaqMan Technology) using fluorescence resonance energy transfer (FRET)
Controls Used	Sample processing control (IC) Positive and negative control	Same
Results Analysis	PCR Cycle threshold analysis	Same

4. NON-CLINICAL PERFORMANCE EVALUATION

4.1. Analytical sensitivity (Limit of Detection)

Analytical sensitivity was determined by analyzing a dilution series of two representative strains/serovars of *Chlamydia trachomatis* (CT, Serovar D and I) and *Neisseria gonorrhoeae* (NG, Strains 2948 and 891). The CT and NG cultures were diluted in pooled negative urine (UR) or pooled negative vaginal swab (VS) clinical specimens to 7 concentration levels. All levels were tested with at least 20 replicates per concentration tested across 3 unique lots of reagents. LoD for each specimen type is shown in [Table 2](#) and [Table 3](#) for CT and NG respectively as the target concentration which can be detected in $\geq 95\%$ of the replicates for all lots.

Table 2: CT concentration levels with at least 95% observed hit rate for all lots tested

Specimen Types	CT Serovar D LoD (EB/mL)	CT Serovar D Mean Ct Value	CT Serovar I LoD (EB/mL)	CT Serovar I Mean Ct Value
Urine in cobas ® PCR Media	0.085	36.2	0.784	36.0
Vaginal Swab in cobas ® PCR Media	0.170	35.3	0.784	35.7

EB = Elementary Bodies

Table 3: NG concentration levels with at least 95% observed hit rate for all lots tested

Specimen Types	NG Strain 2948 LoD (CFU/mL)	NG Strain 2948 Mean Ct Value	NG Strain 891 LoD (CFU/mL)	NG Strain 891 Mean Ct Value
Urine in cobas ® PCR Media	0.250	34.7	0.200	34.5
Vaginal Swab in cobas ® PCR Media	0.500	34.2	0.200	34.5

CFU = Colony Forming Units

4.2. Inclusivity

Inclusivity was performed for an additional 15 CT serovars and 43 NG strains using one lot of reagents. Testing was performed using CT and NG cultures that were spiked into pools of negative clinical specimens. Three replicates per dilution level were tested for each subtype per specimen type. The lowest level at which all three replicates tested as positive are reported in [Table 4](#) and [Table 5](#) for CT and NG respectively.

Table 4: Inclusivity testing for CT serovars

Serovar Type or Variant	Urine Specimens (EB/mL)	Vaginal Swab Specimens (EB/mL)
A	0.1	0.2
B	0.4	0.2
Ba	0.4	1
C	0.7	0.7
E	2	36
F	0.4	0.04
G	0.4	0.4
H	0.4	3
J	0.1	0.2
K	0.1	0.04
LGV Type 1	0.1	0.04
LGV Type 2	1600	200
LGV Type 3	0.1	0.7
nvCT	0.1	0.7
Finnish-nvCT	1:100 of Patient Sample	1:100 of Patient Sample

Table 5: Inclusivity testing for NG strains

Strain ID	Urine Specimens (CFU/mL)	Vaginal Swab Specimens (CFU/mL)
ATCC 27633	0.2	0.5
ATCC 49226	1	0.006
ATCC 700825	0.01	0.001
Clinical Isolate SS169	0.06	0.02
NBL 1606	0.3	0.08
NBL 1952	0.2	0.1
NBL 2012	0.2	0.3
NRL 1977	0.02	0.02
NRL 8042 - Belgium	0.02	0.02
NRL 13477	0.09	0.1
NRL 13819	0.006	0.004
NRL 33155 - Atlanta	0.09	0.001
NRL 33641	0.01	0.07
NRL 35495	0.01	0.07
NRL DAN 09612	0.02	0.03
NRL DN 7896 - DENMARK	0.9	0.3

Strain ID	Urine Specimens (CFU/mL)	Vaginal Swab Specimens (CFU/mL)
NRL DN 7901 - DENMARK	0.02	0.02
NRL DOM 362 - Dominican Republic	0.09	0.09
NRL DOM 1271 - Dominican Republic	0.4	0.1
NRL KPO 1148 - KENYA (KPO)	0.2	0.07
NRL KPO 1161 - KENYA (KPO)	0.02	0.02
NRL Peru 33	0.07	0.07
NRL Peru 83	0.02	0.02
NRL PITT 94-4833 - PITTSBURGH (PITT)	0.02	0.02
NRL PITT 94-8561 - PITTSBURGH (PITT)	0.09	0.1
NRL PP 132 - PHILLIPINES	0.09	0.1
NRL SEN 97 P-292 - SENEGAL (SEN)	0.006	0.02
NRL SEN 97 P-301 - SENEGAL (SEN)	0.006	0.07
Roche Diagnostics K.K.,Japan RDN001-00193	0.02	0.03
Roche Diagnostics, Australia 04D125: Darwin Northern Territory, Australia	0.09	0.1
Roche Diagnostics, Australia 04D127: Darwin Northern Territory, Australia	0.09	0.1
Roche Diagnostics, Australia 04D129: Darwin Northern Territory, Australia	0.09	0.1
Roche Diagnostics, Australia 04D130: Darwin Northern Territory, Australia	0.4	0.1
Roche Diagnostics, Australia 04D132: Darwin Northern Territory, Australia	0.09	0.09
Roche Diagnostics, Australia 05D003: Darwin Northern Territory, Australia	0.02	0.03
Roche Diagnostics, Australia 05D004: Darwin Northern Territory, Australia	0.006	0.004
Roche Diagnostics, Australia 4551 - Western Australia	0.02	0.02
Statens Serum Institut 223/06	0.006	0.006
Statens Serum Institut 1498/46	0.02	0.02
Statens Serum Institut 2170/46	0.02	0.02
Statens Serum Institut 2222/46	0.4	0.09
Statens Serum Institut 6973/45	0.09	0.09
UCSF58	0.06	0.07

4.3. Analytical specificity/cross reactivity

A panel of 181 strains of bacteria, fungi and viruses, including those commonly found in patient specimens, as well as 52 representative strains of non-*gonorrhoeae* *Neisseria* species and other phylogenetically unrelated organisms, were tested to assess analytical specificity. The organisms listed in [Table 12](#) were spiked at concentrations of $\geq 1 \times 10^6$ units/mL* for bacteria or fungi and $\geq 1 \times 10^5$ units/mL for viruses into pools of negative vaginal swab specimens collected in **cobas**® PCR Media and negative urine specimens stabilized in **cobas**® PCR Media. Testing was performed with each potential interfering organism in the absence of, as well as mixed with, CT and NG cultures at $\sim 3x$ LoD. Results indicated that 180 of the

non-target organisms tested did not generate any false positive or false negative results due to cross-reactivity or interference.

*Four bacteria could only be tested at a concentration below 1×10^6 units/mL and above 7×10^4 units/mL due to low stock titers.

Table 6: Microorganisms tested for analytical specificity/cross reactivity

<i>Acholeplasma laidlawii</i>	<i>Eikenella corrodens</i>	<i>Mobiluncus curtisii</i>	<i>Peptostreptococcus anaerobius</i>
<i>Acholeplasma oculi</i> ^{1,3}	<i>Enterobacter aerogenes</i> (<i>Klebsiella aerogenes</i>)	<i>Moraxella catarrhalis</i>	<i>Plesiomonas shigelloides</i>
<i>Acinetobacter calcoaceticus</i>	<i>Enterobacter cloacae</i>	<i>Moraxella lacunata</i>	<i>Prevotella bivia</i>
<i>Acinetobacter lwoffii</i>	<i>Enterococcus avium</i>	<i>Moraxella osloensis</i>	<i>Cutibacterium acnes</i>
<i>Actinomyces israelii</i> ^{1,3}	<i>Enterococcus faecalis</i> (2 strains)	<i>Morganella morganii</i>	<i>Proteus mirabilis</i>
<i>Actinomyces pyogenes</i> (<i>Trueperella pyogenes</i>)	<i>Enterococcus faecium</i> (2 strains)	<i>Mycobacterium smegmatis</i>	<i>Proteus vulgaris</i>
<i>Aerococcus viridans</i>	<i>Erwinia herbicola</i> (<i>Pantoea agglomerans</i>)	<i>Mycoplasma faucium</i> ^{1,3}	<i>Providencia stuartii</i>
<i>Aeromonas hydrophila</i>	<i>Erysipelothrix rhusiopathiae</i>	<i>Mycoplasma fermentans</i>	<i>Pseudomonas aeruginosa</i>
<i>Alcaligenes faecalis</i>	<i>Escherichia coli</i>	<i>Mycoplasma hominis</i>	<i>Pseudomonas fluorescens</i>
<i>Atopobium vaginae</i> (<i>Fannyhessea vaginae</i>)	<i>Flavobacterium meningosepticum</i> (<i>Elizabethkingia meningoseptica</i>)	<i>Mycoplasma orale</i>	<i>Pseudomonas putida</i>
<i>Bacillus subtilis</i>	<i>Fusobacterium nucleatum</i>	<i>Mycoplasma penetrans</i>	<i>Rahnella aquatilis</i>
<i>Bacteroides fragilis</i>	<i>Gardnerella vaginalis</i>	<i>Mycoplasma pirum</i>	<i>Rhizobium radiobacter</i> (<i>Agrobacterium tumefaciens</i>)
<i>Bacteroides ureolyticus</i> (<i>Campylobacter ureolyticus</i>)	<i>Gemella haemolysans</i>	<i>Mycoplasma pneumoniae</i>	<i>Rhodospirillum rubrum</i>
<i>Bifidobacterium adolescentis</i>	<i>Giardia Intestinalis</i>	<i>Mycoplasma primatum</i>	<i>Saccharomyces cerevisiae</i>
<i>Bifidobacterium breve</i>	<i>Haemophilus ducreyi</i>	<i>Mycoplasma salivarium</i>	<i>Salmonella minnesota</i>
<i>Blautia producta</i>	<i>Haemophilus influenzae</i>	<i>Mycoplasma spermatophilum</i>	<i>Salmonella typhimurium</i>
<i>Brevibacterium linens</i>	Herpes simplex virus I	<i>Neisseria cinerea</i> (4 strains)	<i>Serratia marcescens</i>
<i>Campylobacter jejuni</i>	Herpes simplex virus II	<i>Neisseria denitrificans</i> (<i>Bergeriella denitrificans</i>)	<i>Staphylococcus aureus</i>
<i>Candida albicans</i> (2 strains)	HIV-1	<i>Neisseria elongata</i> (3 strains)	<i>Staphylococcus epidermidis</i>
<i>Candida glabrata</i> (<i>Nakaseomyces glabratus</i>)	Human papilloma virus 16 (CaSki cells)	<i>Neisseria flava</i>	<i>Staphylococcus saprophyticus</i>
<i>Candida parapsilosis</i>	<i>Kingella denitrificans</i>	<i>Neisseria flavescens</i> (2 strains)	<i>Streptococcus agalactiae</i>

<i>Candida tropicalis</i>	<i>Kingella kingae</i>	<i>Neisseria lactamica</i> (9 strains) ²	<i>Streptococcus bovis</i>
<i>Chlamydia pneumoniae</i>	<i>Klebsiella oxytoca</i>	<i>Neisseria macacae</i>	<i>Streptococcus mitis</i>
<i>Chlamydia psittaci</i>	<i>Klebsiella pneumoniae</i>	<i>Neisseria meningitidis</i> Serogroup A	<i>Streptococcus mutans</i>
<i>Chromobacterium violaceum</i>	<i>Lactobacillus acidophilus</i>	<i>Neisseria meningitidis</i> Serogroup B	<i>Streptococcus pneumoniae</i>
<i>Citrobacter braakii</i>	<i>Lactobacillus brevis</i> (<i>Levilactobacillus brevis</i>)	<i>Neisseria meningitidis</i> Serogroup C (4 strains)	<i>Streptococcus pyogenes</i>
<i>Citrobacter freundii</i>	<i>Lactobacillus crispatus</i>	<i>Neisseria meningitidis</i> Serogroup D	<i>Streptococcus salivarius</i>
<i>Clostridium difficile</i> (<i>Clostridioides difficile</i>)	<i>Lactobacillus jensenii</i>	<i>Neisseria meningitidis</i> Serogroup W135	<i>Streptococcus sanguinis</i>
<i>Clostridium perfringens</i>	<i>Lactobacillus lactis</i>	<i>Neisseria meningitidis</i> Serogroup Y	<i>Streptomyces griseinus</i>
<i>Corynebacterium genitalium</i>	<i>Lactobacillus vaginalis</i> (<i>Limosilactobacillus vaginalis</i>)	<i>Neisseria mucosa</i> (3 strains)	<i>Trichomonas tenax</i>
<i>Corynebacterium xerosis</i>	<i>Legionella pneumophila</i> (2 strains)	<i>Neisseria perflava</i>	<i>Ureaplasma parvum</i>
<i>Cryptococcus neoformans</i>	<i>Leptotrichia buccalis</i>	<i>Neisseria polysaccharea</i>	<i>Ureaplasma urealyticum</i> ^{1,3}
<i>Cytomegalovirus</i>	<i>Leuconostoc mesenteroides</i>	<i>Neisseria sicca</i> (3 strains)	<i>Veillonella parvula</i>
<i>Deinococcus radiodurans</i>	<i>Leuconostoc paramesenteroides</i> (<i>Weissella paramesenteroides</i>)	<i>Neisseria subflava</i> (14 strains)	<i>Vibrio parahaemolyticus</i>
<i>Derxia gummosa</i>	<i>Listeria monocytogenes</i>	<i>Paracoccus denitrificans</i>	<i>Yersinia enterocolitica</i>
<i>Dientamoeba fragilis</i>	<i>Micrococcus luteus</i>	<i>Pentatrichomonas hominis</i>	-

¹Organism was tested at a concentration of < 1.0e+6 units/mL and > 7.0e+4 units/mL.

²One strain of organism was tested at a concentration of < 1.0e+6 units/mL and > 1.0e+4 units/mL.

³Tested at highest concentration possible per stock concentration.

4.4. Interference

The effects of over-the-counter or prescription products that may be present in urine or vaginal swab clinical specimens were evaluated at the concentration listed in Table 7. Testing was executed using pooled clinical specimens spiked with potential interferents at levels expected from normal patient usage. Interferents were tested in CT/NG negative specimen pools as well as in positive specimen pools spiked with CT/NG at ~3x LoD for each specimen type using one lot of reagents. Five replicates each of CT/NG negative sample and CT/NG positive sample (for each of two culture subtypes per microorganism) were tested with each exogenous substance in each specimen type, except for Azo Urinary Pain Relief, which was tested in urine only.

Of the products tested, no interference was observed in 15 substances when tested at concentrations of 1.5 mg/mL. Azo Urinary Pain Relief and carbomer-containing Replens™ Long-Lasting Vaginal Moisturizer resulted in false negative results in at least one replicate

when tested at higher concentrations. Azo Urinary Pain Relief and Replens™ Long-Lasting Vaginal Moisturizer at concentrations greater than 0.5 mg/mL and 1.0 mg/mL, respectively, may interfere with the assay performance. Levels of substances tolerated by the assay for all specimen types are shown in [Table 7](#).

Table 7: List of products tested for interference

Product Name	Urine (mg/mL)	Vaginal Swabs (mg/mL)
Azo Urinary Pain Relief (urine only)	0.5*	-
Clindamycin Phosphate Vaginal Cream	1.5	1.5
Equate tioconazole 1 Day	1.5	1.5
Equate Vagicaïne Anti-Itch Cream	1.5	1.5
Estradiol Vaginal Cream	1.5	1.5
7 Day vaginal cream	1.5	1.5
K-Y® UltraGel	1.5	1.5
Metronidazole Vaginal Gel	1.5	1.5
Monistat Miconazole Nitrate Vaginal Cream (2%)	1.5	1.5
Monistat® Instant Itch Relief Cream	1.5	1.5
Norforms Deodorant Suppositories	1.5	1.5
Premarin Vaginal Cream	1.5	1.5
Replens™ Long-Lasting Vaginal Moisturizer	1.0*	1.5
Summer's Eve Ultra Freshening Spray	1.5	1.5
VCF - Vaginal Contraceptive Gel	1.5	1.5
Yeast Gard Gel Treatment	1.5	1.5
RepHresh™ Vaginal Gel	1.5	1.5

*Note: Concentrations above this level may cause interference in clinical samples.

Endogenous substances that may be present in urine or vaginal swab clinical specimens were evaluated at the concentration listed in [Table 8](#). Testing was executed using pooled clinical specimens spiked with potential endogenous interferents at levels expected in a typical clinical sample. Endogenous substances were tested in CT/NG negative specimen pools as well as in positive specimen pools spiked with CT/NG at ~3x LoD for each relevant specimen type using one lot of reagents. Five replicates each of CT/NG negative sample and CT/NG positive sample (for each of two culture subtypes per microorganism) were tested with each endogenous substance in each relevant sample type.

For all endogenous substances tested, no interference was observed. Levels of endogenous substances tolerated by the assay for each specimen types are shown in [Table 8](#).

Table 8: Summary of endogenous substance concentrations that do not show interference

Endogenous Substance	Urine	Vaginal Swab
Human cells (PBMCs) cells/mL	1.0E+06	1.0E+06
Mucus	1 swab dipped into mucus	1 swab dipped into mucus
Whole blood (v/v)	10%	10%
Semen (v/v) (vaginal swab only)	-	1.5%
Albumin (w/v) (urine only)	5%	-
Bilirubin (w/v) (urine only)	1% (w/v)	-
Glucose (w/v) (urine only)	1% (w/v)	-
Acidic pH (urine only)	pH 4	-
Alkaline pH (urine only)	pH 9	-

4.5. Competitive inhibition

To assess competitive inhibition between CT and NG, a total of six different combinations of low concentration of target (~2x LoD) were mixed with high concentrations of the other targets in both urine and vaginal swab clinical specimen matrices. Each combination was tested in replicates of 10 using one lot of reagents.

Testing results indicated that when one or two target microorganisms were present at high concentrations, no interference was observed for microorganisms that were present at low concentrations (~2x LoD), when tested in both urine and vaginal swab clinical specimen matrices.

4.6. Reproducibility studies

A reproducibility study was performed across different sites, lots, days, operators, instruments for panels prepared from vaginal swabs and urine in **cobas®** PCR Media. Testing was performed at three external sites with a minimum of 3 **cobas® liat** analyzers per site. Operators at the CLIA-waived sites that met the definition of intended use operators were considered for this study. Selected operators were provided with the assay's IFU, Quick Reference Instructions, and the **cobas® liat** system User Guide. Operators were asked to read the materials before beginning any study testing. No assay or instrument training was provided to the operators.

Two operators at each site each tested 1 panel per specimen type per day (1 complete panel consists of 3 panel members each tested in triplicate) for a total of 15 days. All replicates for each panel member were always tested on the same analyzer. Each panel, per specimen type, consisted of a negative panel member (negative for all 3 analytes), a low positive panel member, and a moderate positive panel member with each positive panel member being co-formulated with all 3 analytes. For each panel member, approximately 270 results were produced.

The Reproducibility Study was executed with a total of 1618 tests consisting of 811 tests for the vaginal specimen type and 807 tests for the urine specimen type.

Table 9 and Table 10 show the site-to-site Reproducibility Study results for cobas® liat CT/NG by sample type and panel member concentration, respectively for CT and NG.

Table 9: Summary CT of site-to-site reproducibility results with cobas® liat CT/NG

Specimen Type	Panel Member Concentration	Site 1*	Site 2*	Site 3*	Overall*
Vaginal	1-2x LoD	100% (90/90) (95.9% - 100.0%)	100% (89/89) (95.9% - 100.0%)	100% (90/90) (95.9% - 100.0%)	100% (269/269) (98.6% - 100.0%)
Vaginal	3-5x LoD	100% (90/90) (95.9% - 100.0%)	100% (90/90) (95.9% - 100.0%)	100% (90/90) (95.9% - 100.0%)	100% (270/270) (98.6% - 100.0%)
Vaginal	Negative	100% (90/90) (95.9% - 100.0%)	100% (83/83) (95.6 - 100.0%)	100% (90/90) (95.9% - 100.0%)	100% (263/263) (98.6% - 100.0%)
Urine	1-2x LoD	87.8% (79/90) (79.4% - 93.0%)	93.3% (83/89) (86.1% - 96.9%)	91.1% (82/90) (83.4% - 95.4%)	90.7% (244/269) (86.6% - 93.6%)
Urine	3-5x LoD	95.6% (86/90) (89.1% - 98.3%)	98.9% (88/89) (93.9% - 99.8%)	94.4% (85/90) (87.6% - 97.6%)	96.3% (259/269) (93.3% - 98.0%)
Urine	Negative	100% (90/90) (95.9% - 100.0%)	100% (80/80) (95.6 - 100.0%)	100% (90/90) (95.9% - 100.0%)	100% 260/260 (98.5% - 100.0%)

Note: LoD: limit of detection

*Percent Agreement with Expected Results (n/N) (95% Confidence Interval)

Table 10: Summary NG of site-to-site reproducibility results with cobas® liat CT/NG

Specimen Type	Panel Member Concentration	Site 1*	Site 2*	Site 3*	Overall*
Vaginal	1-2x LoD	100% (90/90) (95.9% - 100.0%)	100% (89/89) (95.9% - 100.0%)	100% (90/90) (95.9% - 100.0%)	100% (269/269) (98.6% - 100.0%)
Vaginal	3-5x LoD	100% (90/90) (95.9% - 100.0%)	100% (90/90) (95.9% - 100.0%)	100% (90/90) (95.9% - 100.0%)	100% (270/270) (98.6% - 100.0%)
Vaginal	Negative	100% (90/90) (95.9% - 100.0%)	100% (83/83) (95.6 - 100.0%)	100% (90/90) (95.9% - 100.0%)	100% (263/263) (98.6% - 100.0%)
Urine	1-2x LoD	100% (90/90) (95.9% - 100.0%)	98.9% (88/89) (93.9% - 99.8%)	100% (90/90) (95.9% - 100.0%)	99.6% (268/269) (97.9% - 99.9%)

Specimen Type	Panel Member Concentration	Site 1*	Site 2*	Site 3*	Overall*
Urine	3-5x LoD	100% (90/90) (95.9% - 100.0%)	100% (89/89) (95.9% - 100.0%)	100% (90/90) (95.9% - 100.0%)	100% (269/269) (98.6% - 100.0%)
Urine	Negative	100% (90/90) (95.9% - 100.0%)	100% (80/80) (95.6% - 100.0%)	100% (90/90) (95.9% - 100.0%)	100% 260/260 (98.5% - 100.0%)

Note: LoD : limit of detection

*Percent Agreement with Expected Results (n/N) (95% Confidence Interval)

Table 11 and Table 12 present the total SD, and total percent CV (%) for Cycle Threshold Values from the Reproducibility Study for each specimen panel type run in **cobas® liat** CT/NG, respectively for CT and NG.

Table 11: CT - Overall mean estimate, standard deviations, and coefficients of variation (%) for cycle threshold values by sample type and expected concentration for cobas® liat CT/NG by sample type and positive panel member concentration

				Bet- wee n Site	Bet- wee n Site	Bet- wee n Lot	Bet- wee n Lot	Bet- wee n Day	Bet- wee n Day	Bet- wee n Operato r/ Run	Bet- wee n Operato r/ Run	Withi n- Run	Withi n- Run	Tot al	Tota l
Sampl e Type	Panel Memb er Concen - tration	n/N ^a	Mea n Ct	SD	CV%	SD	CV%	SD	CV%	SD	CV%	SD	CV%	SD	CV%
Vagin al	1x-2x LoD	269/269	33.4	0.00	0.00	0.53	1.60	0.22	0.67	0.00	0.00	0.84	2.52	1.02	3.06
Vagin al	3x-5x LoD	270/270	32.1	0.21	0.64	0.58	1.82	0.30	0.93	0.00	0.00	1.00	3.13	1.22	3.79
Urine	1x-2x LoD	244/269	34.8	0.15	0.44	0.84	2.41	0.31	0.88	0.00	0.00	0.91	2.61	1.28	3.69
Urine	3x-5x LoD	259/269	34.0	0.15	0.45	0.70	2.07	0.23	0.68	0.00	0.00	0.98	2.89	1.24	3.65

Ct: cycle threshold; CV%: percent coefficient of variation; LoD: Limit of Detection; SD: standard deviation.

^an is the number of tests in agreement with expected results. N is the total number of valid tests for the panel member.

Table 12: NG - Overall mean estimate, standard deviations, and coefficients of variation (%) for cycle threshold values by sample type and expected concentration for cobas® liat CT/NG by sample type and positive panel member concentration

				Bet- wee n Site	Bet- wee n Site	Bet- wee n Lot	Bet- wee n Lot	Bet- wee n Day	Bet- wee n Day	Bet- wee n Operator/Run	Bet- wee n Operator/Run	Withi n- Run	Withi n- Run	Tota l	Tota l
Sampl e Type	Panel Memb er Concen - tration	n/N ^a	Mea n Ct	SD	CV%	SD	CV%	SD	CV%	SD	CV%	SD	CV%	SD	CV%
Vagin al	1x-2x LoD	269/269	32.2	0.11	0.34	0.59	1.83	0.29	0.89	0.14	0.42	0.59	1.83	0.90	2.79
Vagin al	3x-5x LoD	270/270	30.9	0.10	0.33	0.15	0.50	0.18	0.57	0.00	0.00	0.41	1.33	0.48	1.56
Urine	1x-2x LoD	268/269	32.9	0.16	0.47	0.70	2.12	0.26	0.78	0.46	1.41	0.74	2.25	1.16	3.51
Urine	3x-5x LoD	269/269	31.4	0.07	0.23	0.25	0.80	0.16	0.51	0.00	0.00	0.56	1.79	0.64	2.04

Ct: cycle threshold; CV%: percent coefficient of variation; LoD: Limit of Detection; SD: standard deviation.

^an is the number of tests in agreement with expected results. N is the total number of valid tests for the panel member.

In the Reproducibility Study, the PPA for CT in urine panel members was less than the expected 95%. Therefore, a supplemental Precision Study was performed at one site across different lots, days, operators and instruments for **cobas® liat** CT/NG for the detection of CT in urine from urine panels prepared at negative, 1x-2x and 3x-5xLoD concentration levels. There were six total untrained operators and the level of instructional material were the same for this supplemental Precision study. Each operator tested 1 panel per day for 5 non-consecutive days for each lot (1 complete panel consisted of 3 panel members). This supplemental Precision Study was executed with a total of 810 evaluable tests on urine panel members.

[Table 13](#) shows the supplemental between operator Precision Study for **cobas® liat** CT/NG by panel member concentration for CT in urine.

Table 13: Summary of CT Precision/Repeatability study results

Panel Member Concentration	Operator	n/N ^a	Agreement with Expected Results (%)
1-2x LoD	1	44/45	97.8%
1-2x LoD	2	44/44	100.0%
1-2x LoD	3	45/45	100.0%
1-2x LoD	4	44/44	100.0 %

Panel Member Concentration	Operator	n/N ^a	Agreement with Expected Results (%)
1-2x LoD	5	45/45	100.0%
1-2x LoD	6	45/45	100.0%
3-5x LoD	1	45/45	100.0%
3-5x LoD	2	45/45	100.0%
3-5x LoD	3	45/45	100.0%
3-5x LoD	4	45/45	100.0%
3-5x LoD	5	45/45	100.0%
3-5x LoD	6	44/44	100.0%
Negative	1	43/44	97.7%
Negative	2	45/45	100.0%
Negative	3	45/45	100.0%
Negative	4	45/45	100.0%
Negative	5	45/45	100.0%
Negative	6	44/44	100.0%

^an is the number of tests with expected results. N is the total number of valid tests.

Table 14 shows the supplemental Reproducibility Study for **cobas® liat** CT/NG standard deviation (SD) and coefficient of variation (CV) of Cycle Threshold Values for each factor as well as the total SD and total CV (%) for each positive panel member.

Table 14: CT - Overall mean estimate, standard deviations, and coefficients of variation (%) for cycle threshold values and expected concentration for cobas® liat CT/NG by positive panel member concentration in urine

			Between n Instrum ent	Between n Instrum ent	Bet - we en Lot	Bet - we en Lot	Bet - we en Day	Bet - we en Day	Bet-ween Operator/ Run	Bet-ween Operator/ Run	Withi n- Run	Withi n- Run	Tot al	Tot al
Panel Member Concentra tion	n/N ^a	Me an Ct	SD	CV%	SD	CV%	SD	CV%	SD	CV%	SD	CV%	SD	CV %
1x-2xLOD	267/268	35.3	0.00	0.00	0.03	0.08	0.00	0.00	0.00	0.00	0.85	2.41	0.85	2.41
3x-5xLOD	269/269	33.7	0.00	0.00	0.00	0.00	0.00	0.00	0.46	1.35	1.07	3.19	1.17	3.47

Note: Ct = cycle threshold, CT=Chlamydia trachomatis, CV(%) = percent coefficient of variation, LoD = Limit of Detection, NG=Neisseria gonorrhoeae, SD =standard deviation.

^an is the number of tests in agreement with expected results. N is the total number of valid tests for the panel member.

5. CLINICAL PERFORMANCE EVALUATION

5.1. Clinical study

The clinical utility and performance of **cobas® liat** CT/NG was established in a multi-site, prospective study by comparing the results to a Patient Infected Status (PIS) or a Composite Comparator Algorithm (CCA) derived from a combination of FDA-cleared NAATs for the 2 analytes. A result for PIS (for male urine urine) or a CCA (for vaginal swabs and female urine) was generated for CT and NG. Male urine, vaginal swabs, and female urine were collected and tested at 13 geographically diverse intended use clinical sites across the US. There were 48 operators that took part in **cobas® liat** CT/NG testing, of which, 43 represented CLIA-waived operators. Five of the 48 operators represented experienced laboratorians in a moderate complexity laboratory.

A total of 4852 subjects (2512 females and 2340 males) were enrolled in the study and provided specimens for collection. Note, two subjects, declared male at birth, provided vaginal swab specimens. Of these subjects, 72 were non-evaluable due to protocol deviations and incidents (18), invalid **cobas® liat** CT/NG and/or final comparator result (45), or sample collection incidents (9). Of the evaluable subjects, 2304 male subjects provided 2302 male urine specimens (2 subjects provided vaginal swab specimens) and 2476 females provided 1240 clinician-collected vaginal swabs and 1236 self-collected vaginal swabs for evaluation in the clinical study.

Prospectively enrolled female subjects provided 4 vaginal swab specimens, three for comparator tests and one for the **cobas® liat** CT/NG nucleic acid test. Vaginal swab specimen for the **cobas® liat** CT/NG nucleic acid test was either collected by clinician or self-collected. Female subjects also provided a urine specimen that was aliquoted into the respective manufacturers' collection devices and **cobas®** PCR Media.

Prospectively enrolled male subjects provided a urine specimen that was aliquoted into the respective manufacturers' collection devices and **cobas®** PCR Media.

Specimens were tested for CT and NG with the investigational and the reference comparator NAATs. All tests were run according to the respective IFUs.

The clinical performance of **cobas® liat** CT/NG was evaluated by comparing the results from collected specimen types to a pre-specified PIS/CCA result. The PIS/CCA result for each analyte was derived from a combination of 3 reference NAATs (NAAT1, NAAT2, and NAAT3). If NAAT1 and NAAT2 are concordant, then the final PIS/CCA result for the respective analyte is the concordant result obtained from NAAT1 and NAAT2. If NAAT1 and NAAT2 are discordant, then NAAT3 is performed to be the tiebreaker between the first 2 discordant results. [Table 15](#) below shows the PIS and CCA algorithm for each analyte.

Table 15: Determination of the PIS/CCA result for CT and NG, respectively

NAAT 1	NAAT 2	NAAT 3	Patient Infected Status ^a	Composite Comparator Algorithm
+	+	N/A	Infected	Positive
+	-	+	Infected	Positive
-	+	+	Infected	Positive
-	-	N/A	Not Infected	Negative
+	-	-	Not Infected	Negative
-	+	-	Not Infected	Negative
-	Invalid	+	Indeterminate	Indeterminate
-	Invalid	-	Not Infected	Negative
Invalid	-	+	Indeterminate	Indeterminate
Invalid	-	-	Not Infected	Negative
+	Invalid	-	Indeterminate	Indeterminate
Invalid	+	-	Indeterminate	Indeterminate
+	Invalid	+	Infected	Positive
Invalid	+	+	Infected	Positive
Invalid	Invalid	N/A	Indeterminate	Indeterminate

N/A: not applicable; NAAT: nucleic acid amplification test.

^aThe “Infected” or “Not Infected” patient infected status was derived from the total combination of results obtained from the reference NAATs.

The sample types of male urine and vaginal swab were used to create the PIS and CCA results, respectively, for men and women. The **cobas® liat** CT/NG results of each analyte from each sample type (male urine and vaginal swab) were compared to the PIS/CCA result to determine the clinical performance of the assay. Sensitivity (SENS), specificity (SPEC), positive percent agreement (PPA), and negative percent agreement (NPA), of **cobas® liat** CT/NG were calculated separately for CT and NG.

Supplementation with archived specimens was included in this study due to the expected low NG prevalence for male urine and vaginal swabs. The archived specimens were prospectively collected samples from a prior clinical trial study (K173887).

A supplemental study was conducted to evaluate the performance of **cobas® liat** CT/NG for the assessment of detecting CT in female urine. There were a total of 785 prospective samples from subjects who met the study eligibility criteria. Of these 785 samples, there were a total of 751 samples with evaluable results and included in the final data set. In this study, comparator testing and final evaluation was determined by comparing results from **cobas® liat** CT/NG to a CCA as described above and shown in [Table 15](#). This supplemental set of data for CT was combined with the female urine data from the aforementioned study. The performance data tables below ([Table 17](#)) will be labeled to indicate that the performance was generated as part of the supplemental study.

Additionally, the **cobas® liat** CT/NG performance for the detection NG, also from female urine is presented below ([Table 19](#)). Similarly, the CCA was determined from 3 comparator NAATs as shown in [Table 15](#). This set of data was part of the original study described above.

A vaginal swab (self-collected or clinician-collected) is the preferred specimen type for CT and NG testing in females due to a higher sensitivity when compared to urine. If a female urine is used and CT and/or NG testing is negative, further testing with a vaginal swab may be indicated if *C. trachomatis* and/or *N. gonorrhoeae* infection is strongly suspected.

Analysis of both studies show that the **cobas® liat** CT/NG test detected 8.4% fewer CT infections and 6.9% fewer NG infections in female urine when compared to a vaginal swab CCA versus a female urine CCA.

5.2. Performance results

Sensitivity/PPA and specificity/NPA of **cobas® liat** CT/NG as defined by the PIS/CCA results are presented by gender, sample type, and symptom status in [Table 16](#) and [Table 17](#) for CT, and [Table 18](#) and [Table 19](#) for NG.

Upon initial testing, the **cobas® liat** CT/NG invalid rate was 0.6% and after retesting the final invalid rate was 0.2%.

Table 16: CT - Clinical performance of cobas® liat CT/NG compared with PIS/CCA by specimen type and symptom status - prospectively collected specimens

Specimen Type	Symptom Status	N	Sensitivity Estimate (95% CI)	Sensitivity n/N	Specificity Estimate (95% CI)	Specificity n/N
Male Urine	Symptomatic	808	98.2% (90.6%, 99.7%)	55/56	99.9% (99.3%, 100.0%)	751/752
Male Urine	Asymptomatic	1488	96.4% (87.7%, 99.0%)	53/55	99.9% (99.6%, 100.0%)	1432/1433
Male Urine	Total	2296	97.3% (92.4%, 99.1%)	108/111	99.9% (99.7%, 100.0%)	2183/2185
Specimen Type	Symptom Status	N	Positive Percent Agreement Estimate (95% CI)	Positive Percent Agreement n/N	Negative Percent Agreement Estimate (95% CI)	Negative Percent Agreement n/N
Vaginal Swabs	Symptomatic	1116	98.4% (91.3%, 99.7%)	60/61	99.7% (99.2%, 99.9%)	1052/1055
Vaginal Swabs	Asymptomatic	1357	97.9% (89.1%, 99.6%)	47/48	99.8% (99.4%, 100.0%)	1307/1309
Vaginal Swabs	Total	2473	98.2% (93.6%, 99.5%)	107/109	99.8% (99.5%, 99.9%)	2359/2364

CI: confidence interval

Table 17: CT – Clinical Performance of cobas® liat® CT/NG compared with CCA by symptom status - Female Urine

Study	Symptom Status	N	Positive Percent Agreement Estimate (95% CI)	Positive Percent Agreement n/N	Negative Percent Agreement Estimate (95% CI)	Negative Percent Agreement n/N
Supplemental Study	Symptomatic	312	100.0% (81.6%, 100.0%)	17/17	99.3% (97.6%, 99.8%)	293/295
Supplemental Study	Asymptomatic	439	93.8% (71.7%, 98.9%)	15/16	99.3% (97.9%, 99.8%)	420/423
Supplemental Study	Overall	751	97.0% (84.7%, 99.5%)	32/33	99.3% (98.4%, 99.7%)	713/718
Original LIA-497 Study FU Subset	Symptomatic	1113	93.0% (83.3%, 97.2%)	53/57	99.8% (99.3%, 99.9%)	1054/1056
Original LIA-497 Study FU Subset	Asymptomatic	1346	97.6% (87.4%, 99.6%)	40/41	99.8% (99.3%, 99.9%)	1302/1305
Original LIA-497 Study FU Subset	Overall	2459	94.9% (88.6%, 97.8%)	93/98	99.8% (99.5%, 99.9%)	2356/2361
Combined	Symptomatic	1425	94.6% (86.9%, 97.9%)	70/74	99.7% (99.2%, 99.9%)	1347/1351
Combined	Asymptomatic	1785	96.5% (88.1%, 99.0%)	55/57	99.7% (99.2%, 99.8%)	1722/1728
Combined	Overall	3210	95.4%* (90.4%, 97.9%)	125/131	99.7% (99.4%, 99.8%)	3069/3079

CI: confidence interval

Table 18: NG - Clinical performance of cobas® liat CT/NG compared with PIS/CCA by specimen type and symptom status

Specimen Type	Symptom Status	N	Sensitivity Estimate (95% CI)	Sensitivity n/N	Specificity Estimate (95% CI)	Specificity n/N
Male Urine	Symptomatic	813	100.0% (94.7%, 100.0%)	68/68	100.0% (99.5%, 100.0%)	745/745
Male Urine	Asymptomatic	1488	100.0% (74.1%, 100.0%)	11/11	99.8% (99.4%, 99.9%)	1474/1477
Male Urine	Total	2301	100.0% (95.4%, 100.0%)	79/79	99.9% (99.6%, 100.0%)	2219/2222

Archived Male Urine	Symptomatic	125	100.0% (95.2%, 100.0%)	77/77	100.0% (92.6%, 100.0%)	48/48
Archived Male Urine	Asymptomatic	38	100.0% (56.6%, 100.0%)	5/5	100.0% (89.6%, 100.0%)	33/33
Archived Male Urine	Total	163	100.0% (95.5%, 100.0%)	82/82	100.0% (95.5%, 100.0%)	81/81
Overall Male Urine	Symptomatic	938	100.0% (97.4%, 100.0%)	145/145	100.0% (99.5%, 100.0%)	793/793
Overall Male Urine	Asymptomatic	1526	100.0% (80.6%, 100.0%)	16/16	99.8% (99.4%, 99.9%)	1507/1510
Overall Male Urine	Total	2464	100.0% (97.7%, 100.0%)	161/161	99.9% (99.6%, 100.0%)	2300/2303
Specimen Type	Symptom Status	N	Positive Percent Agreement Estimate (95% CI)	Positive Percent Agreement n/N	Negative Percent Agreement Estimate (95% CI)	Negative Percent Agreement n/N
Vaginal Swabs	Symptomatic	1115	91.7% (74.2%, 97.7%)	22/24	99.8% (99.3%, 99.9%)	1089/1091
Vaginal Swabs	Asymptomatic	1357	100.0% (82.4%, 100.0%)	18/18	99.9% (99.5%, 100.0%)	1337/1339
Vaginal Swabs	Total	2472	95.2% (84.2%, 98.7%)	40/42	99.8% (99.6%, 99.9%)	2426/2430
Archived Vaginal Swabs	Symptomatic	42	100.0% (83.9%, 100.0%)	20/20	100.0% (85.1%, 100.0%)	22/22
Archived Vaginal Swabs	Asymptomatic	48	100.0% (86.7%, 100.0%)	25/25	100.0% (85.7%, 100.0%)	23/23
Archived Vaginal Swabs	Total	90	100.0% (92.1%, 100.0)	45/45	100.0% (92.1%, 100.0%)	45/45
Overall Vaginal Swabs	Symptomatic	1157	95.5% (84.9%, 98.7%)	42/44	99.8% (99.3%, 100.0%)	1111/1113
Overall Vaginal Swabs	Asymptomatic	1405	100.0% (91.8%, 100.0%)	43/43	99.9% (99.5%, 100.0%)	1360/1362
Overall Vaginal Swabs	Total	2562	97.7% (92.0%, 99.4%)	85/87	99.8% (99.6%, 99.9%)	2471/2475

CI: confidence interval

Table 19: NG - Clinical performance of cobas® liat® CT/NG compared with CCA by symptom status- Female Urine

Symptom Status	N	Positive Percent Agreement Estimate (95% CI)	Positive Percent Agreement n/N	Negative Percent Agreement Estimate (95% CI)	Negative Percent Agreement n/N
Symptomatic	1111	100.0% (83.9%, 100.0%)	20/20	99.9% (99.5%, 100.0%)	1090/1091
Asymptomatic	1347	100.0% (81.6%, 100.0%)	17/17	99.9% (99.6%, 100.0%)	1329/1330
Overall	2458	100.0%* (90.6%, 100.0%)	37/37	99.9% (99.7%, 100.0%)	2419/2421

CI: confidence interval

5.3. Expected values for urogenital specimens

The positivity rate of the **cobas® liat** CT/NG nucleic acid assay test for CT, and NG observed during the study is shown for each specimen type, by collection site in [Table 20](#) below and from the Supplemental Female Urine CT study in [Table 21](#)..

Table 20: Positivity of CT/NG as Determined by the cobas® liat CT/NG nucleic acid test by Specimen Type and Clinical Site

Collection Site	CT	CT	CT	NG	NG	NG
	Male Urine	Vaginal Swab	Female Urine	Male Urine	Vaginal Swab	Female Urine
1	8.7% (30/343)	9.2% (14/152)	6.6% (10/151)	11.0% (38/346)	3.3% (5/152)	2.0% (3/151)
2	2.6% (9/346)	6.2% (15/241)	5.4% (13/240)	1.7% (6/346)	3.7% (9/241)	3.8% (9/240)
3	11.9% (18/151)	8.2% (30/366)	7.4% (27/365)	6.6% (10/151)	0.55% (2/364)	0.6% (2/362)
4	11.2% (12/107)	0.63% (1/160)	0.0% (0/161)	9.3% (10/107)	1.25% (2/160)	1.2% (2/161)
5	0.0% (0/4)	NC	NC	0.0% (0/4)	NC	NC
6	0.9% (1/117)	1.2% (1/85)	2.4% (2/83)	0.0% (0/118)	1.2% (1/85)	0.0% (0/83)
7	5.3% (3/57)	5.7% (2/35)	7.5% (3/40)	1.8% (1/57)	2.9% (1/35)	2.5% (1/40)
8	0.0% (0/80)	0.0% (0/19)	0.0% (0/17)	2.5% (2/80)	0.0% (0/19)	0.0% (0/17)
9	1.3% (6/468)	1.9% (10/527)	1.9% (10/524)	0.4% (2/469)	2.3% (12/528)	1.5% (8/526)
10	0.5% (1/198)	1.4% (5/347)	0.9% (3/348)	1.0% (2/198)	0.86% (3/347)	0.9% (3/348)
11	17.1% (18/105)	5.6% (17/305)	4.6% (14/304)	4.8% (5/105)	2.0% (6/305)	2.3% (7/304)
12	3.5% (10/289)	8.8% (12/136)	9.6% (13/136)	1.7% (5/289)	1.5% (2/136)	1.5% (2/136)
13	6.5% (2/31)	5.0% (5/100)	4.0% (4/101)	3.2% (1/31)	1.0% (1/100)	2.0% (2/101)

Note: two subjects were born male, but were evaluated using self-collected vaginal swabs.

NC: non calculable as no female subjects were enrolled at this site

Table 21: Positivity of CT as Determined by the cobas® liat CT/NG nucleic acid test from Supplement Female Urine Study

-	CT
Collection Site	Female Urine
14	4.9% (37/751)

5.4. Summary of Positive Predictive Value, Negative Predictive Value for Hypothetical Prevalence

The hypothetical PPVs and NPVs of **cobas® liat** CT/NG derived from disease prevalences of 1% to 50% are shown in [Table 22](#) and [Table 23](#) for CT, and [Table 24](#) and [Table 25](#) for NG.

Table 22: CT - Positive predictive value and negative predictive value for hypothetical CT prevalence

Specimen type for CNMA testing	Hypothetical Prevalence (%)	PPV (%)	NPV (%)
Male Urine	1	91.5	100.0
Male Urine	3	97.0	99.9
Male Urine	5	98.2	99.9
Male Urine	10	99.2	99.7
Male Urine	15	99.5	99.5
Male Urine	20	99.6	99.3
Male Urine	30	99.8	98.9
Male Urine	50	99.9	97.4
Vaginal Swabs	1	82.4	100.0
Vaginal Swabs	3	93.5	99.9
Vaginal Swabs	5	96.1	99.9
Vaginal Swabs	10	98.1	99.8
Vaginal Swabs	15	98.8	99.7
Vaginal Swabs	20	99.1	99.5
Vaginal Swabs	30	99.5	99.2
Vaginal Swabs	50	99.8	98.2

Note: NPV: negative predictive value; PPV: positive predictive value; PPA: positive percent agreement; NPA: negative percent agreement;

The PPV and NPV were calculated using the sensitivity/PPA and specificity/NPA of **cobas® liat** CT/NG from the prospectively collected population.

Table 23: CT - Positive predictive value and negative predictive value for hypothetical CT prevalence (original and supplemental studies) – Female Urine

Hypothetical Prevalence (%)	PPV (%)	NPV (%)
1	74.8	100.0
3	90.1	99.9
5	93.9	99.8
10	97.0	99.5
15	98.1	99.2

Hypothetical Prevalence (%)	PPV (%)	NPV (%)
20	98.7	98.9
30	99.2	98.1
50	99.7	95.6

Note: NPV: negative predictive value; PPV: positive predictive value; PPA: positive percent agreement; NPA: negative percent agreement

The PPV and NPV were calculated using the PPA and NPA of **cobas® liat** CT/NG from the prospectively collected population.

Table 24: NG - Positive predictive value and negative predictive value for hypothetical NG prevalence

Specimen type for CNMA testing	Hypothetical Prevalence (%)	PPV (%)	NPV (%)
Male Urine	1	88.2	100.0
Male Urine	3	95.8	100.0
Male Urine	5	97.5	100.0
Male Urine	10	98.8	100.0
Male Urine	15	99.2	100.0
Male Urine	20	99.5	100.0
Male Urine	30	99.7	100.0
Male Urine	50	99.9	100.0
Vaginal Swabs	1	85.4	100.0
Vaginal Swabs	3	94.7	99.9
Vaginal Swabs	5	96.8	99.7
Vaginal Swabs	10	98.5	99.5
Vaginal Swabs	15	99.0	99.2
Vaginal Swabs	20	99.3	98.8
Vaginal Swabs	30	99.6	98.0
Vaginal Swabs	50	99.8	95.4

Note: NPV: negative predictive value; PPV: positive predictive value; PPA: positive percent agreement; NPA: negative percent agreement;

The PPV and NPV were calculated using the sensitivity/PPA and specificity/NPA of **cobas® liat** CT/NG from the prospectively collected population.

Table 25: NG - Positive predictive value and negative predictive value for hypothetical NG prevalence – Female Urine

Hypothetical Prevalence (%)	PPV (%)	NPV (%)
1	92.4	100.0
3	97.4	100.0
5	98.5	100.0
10	99.3	100.0

Hypothetical Prevalence (%)	PPV (%)	NPV (%)
15	99.5	100.0
20	99.7	100.0
30	99.8	100.0
50	99.9	100.0

Note: NPV: negative predictive value; PPV: positive predictive value;

The PPV and NPV were calculated using the sensitivity and specificity of **cobas® liat** CT/NG from the prospectively collected population.

6. CONCLUSIONS

A comparison of the intended use, technological characteristics, and the results of non-clinical analytical and clinical performance studies demonstrate that **cobas® liat** CT/NG nucleic acid test is **substantially equivalent** to the predicate devices.