Serology Test Evaluation Report for “Platelia SARS-CoV-2 Total Ab” from Bio-Rad

June 20, 2021

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1 Introduction

The Platelia SARS-CoV-2 Total Ab from Bio-Rad was tested on 2021-05-04 at the Frederick National Laboratory for Cancer Research (FNLCR), a Federally Funded Research and Development Center (FFRDC) sponsored by the National Cancer Institute (NCI). Tests were from lot number 0F0015. The Platelia SARS-CoV-2 Total Ab is intended to qualitatively detect Pan Ig.

1.1 Panel composition

![Graph A: IgM+ Titers in Panel 3](image)

![Graph B: IgG+ Titers in Panel 3](image)

Figure 1: Titer levels for (A) IgM+ and (B) IgG+ samples according to the CDC SARS-CoV-2 Spike antigen assay

The test was evaluated against “Panel 3,” which includes frozen SARS-CoV-2 antibody-positive serum samples \(n = 30\) and frozen antibody-negative Anticoagulant Citrate Dextrose Solution Formula A (ACD-A) plasma samples \(n = 80\). While ACD-A plasma may not be commonly used in clinical practice for serological testing, ACD-A plasma samples were used here because these pre-pandemic samples were most easily acquired from blood banks. The panel size and composition were chosen to enable a laboratory-based evaluation and to provide reasonable estimates and confidence intervals for test performance in the context of limited sample availability. The sample size is comparable to that of a typical sample size used to support Emergency Use Authorization (EUA) by FDA for tests of this type.
1.1.1 Positive samples

Positive samples used in Panel 3 were from patients previously confirmed to have SARS-CoV-2 infection with a nucleic acid amplification test (NAAT). Time between symptom onset, NAAT testing, and sample collection is not known for all samples. Both SARS-CoV-2 IgM and IgG antibodies are present in all Panel 3 positive samples. The Centers for Disease Control and Prevention (CDC) detected the presence of IgG and IgM antibodies at their laboratory using their SARS-CoV-2 spike enzyme-linked immunosorbent assay (ELISA) tests.\(^1\) The presence of antibodies was confirmed at FNLCR using CDC's developed ELISAs (Pan-Ig, IgG, and IgM) as well as an IgG Receptor Binding Domain (RBD) ELISA developed by the Krammer Laboratory at the Icahn School of Medicine at Mount Sinai.\(^2\) The positive samples selected may not reflect the distribution of antibody levels in patient populations that would be evaluated by such a test. Because all samples are positive for both IgM and IgG, this evaluation cannot verify that tests intended to detect IgM and IgG antibodies separately detect these antibodies independently.

Positive samples were assessed at dilutions of 1:100, 1:400, 1:1600, and 1:6400 by CDC on their Pan-Ig assay, their IgM assay, and their IgG assay. Some samples were run at additional dilutions. Any samples that were positive at a dilution greater than 1:6400 were assigned a titer of 6400 because 1:6400 was the highest dilution at which all positive samples used in these evaluations were assessed.

1.1.2 Negative samples

All Panel 3 negative samples were collected prior to 2020, before the SARS-CoV-2 virus is known to have circulated in the United States. Panel 3 groups include:

- “Negatives” \((n = 70)\): selected without regard for clinical status. This group includes a sample, C0063, that showed reactivity in the Pan-Ig CDC spike ELISA at FNLCR.

- “HIV+” \((n = 10)\): selected from banked plasma from HIV+ patients.\(^3\) This group includes 3 samples, C0018, C0155, and C0182, that showed reactivity in the IgG RBD ELISA at FNLCR.

All Panel 3 negative samples were assessed at dilutions of 1:100 and 1:400 by CDC on their Pan-Ig assay. A subset of samples was assessed in parallel at additional dilutions and on the CDC IgM assay.

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\(^1\)See [https://www.cdc.gov/coronavirus/2019-ncov/lab/serology-testing.html](https://www.cdc.gov/coronavirus/2019-ncov/lab/serology-testing.html), which notes “CDC's serologic test has been designed and validated for surveillance and research purposes. It is designed to estimate the percentage of the U.S. population previously infected with the virus – information needed to guide the response to the pandemic and protect the public's health. The CDC test is not currently designed to test individuals who want to know if they have been previously infected with SARS-CoV-2. Commercial tests are available to provide test results to individuals.”

\(^2\)An implementation of this test, the COVID-19 ELISA IgG Antibody Test, has been granted an EUA authorization by FDA for use at the Mount Sinai Laboratory (MSL), Center for Clinical Laboratories, a division of the Department of Pathology, Molecular, and Cell-Based Medicine, New York, NY. See [https://www.fda.gov/media/137029/download](https://www.fda.gov/media/137029/download).

\(^3\)HIV+ samples were deemed appropriate for inclusion in the panel: (1) to increase the sample size and reduce the confidence interval; and (2) to identify any possibility of cross-reactivity with HIV+ samples. It is anticipated that other types of samples, as they become available, may also be evaluated in any future analyses.
and IgG assays. All Panel 3 negative samples were negative at a dilution of 1:100 on the CDC Pan-Ig assay. These samples were assigned an undetectable titer (represented as zero (0) in the line data) for the Pan-Ig assay, the IgM assay, and the IgG assay.

1.2 Analysis

Samples used in this evaluation were not randomly selected, and sensitivity (PPA) and specificity (NPA) estimates in this report may not be indicative of the real-world performance of the Bio-Rad Platelia SARS-CoV-2 Total Ab. Sensitivity and specificity were calculated for each antibody (e.g., IgM, IgG, IgA, and Pan-Ig, as applicable) separately. For sensitivity and specificity calculations, equivocal results on positive samples were counted as false negative results, and equivocal results on negative samples were counted as false positive results. In addition, sensitivity and specificity were estimated in a combined manner, where a positive result for any antibody the Bio-Rad Platelia SARS-CoV-2 Total Ab is intended to detect was considered as a positive test result and a negative result meant that a sample tested negative for all antibodies the Bio-Rad Platelia SARS-CoV-2 Total Ab is intended to detect. Positive and negative predictive values were calculated for combined sensitivity and specificity assuming a prevalence of 5%. Cross-reactivity with HIV+ was evaluated, and results are presented separately. If cross-reactivity was detected, the samples with HIV+ were not included in calculations of specificity.

Confidence intervals for sensitivity and specificity were calculated per a score method described in CLSI EP12-A2 (2008). Confidence intervals for PPV and NPV were calculated using the values from the 95% confidence intervals for sensitivity and specificity. For evaluation of cross-reactivity with HIV+, it was evaluated whether an increased false positive rate among antibody negative samples with HIV was statistically higher than the false positive rate among antibody negative samples without HIV (for this, a confidence interval for the difference in false positive rates was calculated per a score method described by Altman).

1.3 Important caveats

Sensitivity and specificity estimates in this report may not be indicative of the real world performance of the Bio-Rad Platelia SARS-CoV-2 Total Ab.

These results are based on serum and plasma samples only and may not be indicative of performance with other sample types, such as whole blood, including finger stick blood.

The number of samples in the panel is a minimally viable sample size that still provides reasonable estimates and confidence intervals for test performance, and the samples used may not be repre-

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4In this report, device outputs indicating equivocal results, including outputs such as “borderline” or similar, are referred to as “equivocal.”


sentative of the antibody profile observed in patient populations.

1.4 Notes about the evaluation procedure

- The Bio-Rad Platelia SARS-CoV-2 Total Ab was used per the manufacturer's package insert.
- Devices were tested within any expiration dates provided.
- Devices were not obviously defective / compromised.
- Devices were stored at the FNLCR within their labeled conditions.
- A single operator conducted the test.
- The personnel who performed the testing were blinded to the identity / code of the sample and the expected results.
- The testing was performed in a non-clinical laboratory environment.
- Negative and positive samples were ordered randomly and then tested serially.
- The Bio-Rad Platelia SARS-CoV-2 Total Ab was run with positive and negative controls.

1.5 Additional notes, anomalies, and clarifications

The FNLCR provided the following additional information:

While the package insert specifies the use of human plasma (EDTA), and serum, this evaluation used antibody-positive serum samples and antibody-negative ACD-A plasma samples.
2 Results

Table 1: Summary Results

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<tr>
<th>Comparator Method</th>
<th>Collected pre-2020</th>
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<td></td>
<td>Antibody Positive</td>
</tr>
<tr>
<td></td>
<td>Antibody Negative</td>
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<td>Platelia SARS-CoV-2 Total Ab</td>
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<td>Total</td>
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Table 2: Summary Statistics

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<td>(70.3%; 94.7%)</td>
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<td>Pan Ig Specificity</td>
<td>100% (80/80)</td>
<td>(95.4%; 100%)</td>
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<tr>
<td>Combined Sensitivity</td>
<td>86.7% (26/30)</td>
<td>(70.3%; 94.7%)</td>
</tr>
<tr>
<td>Combined Specificity</td>
<td>100% (80/80)</td>
<td>(95.4%; 100%)</td>
</tr>
<tr>
<td>Combined PPV for prevalence = 5.0%</td>
<td>100%</td>
<td>(44.7%; 100%)</td>
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<td>Combined NPV for prevalence = 5.0%</td>
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<td>Cross-reactivity with HIV+</td>
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### 3 Line Data

In the table below, “Days” refers to “Days from symptom onset to blood collection.”

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