510(k) SUBSTANTIAL EQUIVALENCE DETERMINATION DECISION SUMMARY

A. 510(k)

K163536

B. Purpose for Submission:

To obtain a substantial equivalent determination for the MALDI Biotyper CA System.

C. Measurand:

See Intended Use.

D. Type of Test:

The MALDI Biotyper CA System is a qualitative *in vitro* diagnostic device intended for the identification of Gram-negative bacterial colonies cultured from human specimens. The device is comprised of an ionization source, a mass analyzer and a spectral database. The device is indicated for use in conjunction with other clinical and laboratory findings to aid in the diagnosis of Gram-negative bacterial infections.

E. Applicant:

Bruker Daltonik GmbH

F. Proprietary and Established Names:

Trade Name: MALDI Biotyper CA (MBT-CA) System, MBT smart CA System

Common Names: MBT-CA, System, mass spectrometry

G. Regulatory Information:

- 1. <u>Regulation section:</u> 21 CFR 866.3361 Instrumentation for clinical multiplex test systems
- 2. <u>Classification:</u> Class II (special controls)
- 3. <u>Product code:</u> PEX
- 4. <u>Panel:</u> Microbiology (83)

H. Intended Use:

1. Intended use(s):

The MALDI Biotyper CA System is a mass spectrometer system using matrix-assisted laser desorption/ionization - time of flight (MALDI-TOF) for the identification of microorganisms cultured from human specimens.

The MALDI Biotyper CA System is a qualitative in vitro diagnostic device indicated for use in conjunction with other clinical and laboratory findings to aid in the diagnosis of bacterial and yeast infections.

The following organisms are claimed:

Bacteria:			
Abiotrophia defectiva	Achromobacter xylosoxidans		
Acinetobacter baumannii / nosocomialis group	Acinetobacter calcoaceticus		
Acinetobacter haemolyticus	Acinetobacter johnsonii		
Acinetobacter junii	Acinetobacter lwoffii		
Acinetobacter pittii	Acinetobacter radioresistens		
Acinetobacter ursingii	Actinomyces europaeus		
Actinomyces funkei	Actinomyces graevenitzii		
Actinomyces hyovaginalis	Actinomyces meyeri		
Actinomyces neuii	Actinomyces odontolyticus		
Actinomyces oris	Actinomyces radingae		
Actinomyces turicensis	Actinomyces urogenitalis		
Actinotignum schaalii group	Aerococcus sanguinicola		
Aerococcus urinae	Aerococcus viridans		
Aeromonas salmonicida	Aeromonas sp[7]		
Aggregatibacter actinomycetemcomitans	Aggregatibacter aphrophilus		
Aggregatibacter segnis	Alcaligenes faecalis		
Alloiococcus otitis	Alloscardovia omnicolens		
Anaerococcus murdochii	Anaerococcus vaginalis		
Arthrobacter cumminsii	Bacteroides caccae		
Bacteroides fragilis	Bacteroides nordii		
Bacteroides ovatus group	Bacteroides pyogenes		
Bacteroides salyersiae	Bacteroides stercoris group		
Bacteroides thetaiotaomicron group	Bacteroides uniformis		
Bacteroides vulgatus group	Bifidobacterium breve		
Bordetella group [3]	Bordetella hinzii		
Brevibacterium casei	Brevundimonas diminuta group		
Burkholderia cepacia complex [13]	Burkholderia gladioli		
Burkholderia multivorans	Campylobacter coli		
Campylobacter jejuni	Campylobacter ureolyticus		
Capnocytophaga ochracea	Capnocytophaga sputigena		
Chryseobacterium gleum	Chryseobacterium indologenes		
Citrobacter amalonaticus complex	Citrobacter freundii complex		
Citrobacter koseri	Clostridium beijerinckii		
Clostridium bifermentans	Clostridium butyricum		
Clostridium clostridioforme group	Clostridium difficile		
Clostridium innocuum	Clostridium paraputrificum		

Bacteria:			
Clostridium perfringens	Clostridium ramosum		
Clostridium septicum	Clostridium sordellii		
Clostridium sporogenes /	Clostridium tertium		
Clostridium botulinum (group I)			
Corynebacterium accolens	Corynebacterium afermentans group		
Corynebacterium amycolatum	Corynebacterium aurimucosum group		
Corynebacterium bovis	Corynebacterium coyleae		
Corynebacterium diphtheriae	Corynebacterium freneyi		
Corynebacterium glucuronolyticum	Corynebacterium glutamicum		
Corynebacterium jeikeium	Corynebacterium kroppenstedtii		
Corynebacterium macginleyi	Corynebacterium minutissimum		
Corynebacterium mucifaciens / ureicelerivorans group	Corynebacterium propinquum		
Corynebacterium pseudodiphtheriticum	Corynebacterium pseudotuberculosis		
Corynebacterium resistens	Corynebacterium riegelii		
Corynebacterium striatum group	Corynebacterium tuberculostearicum		
Corynebacterium ulcerans	Corynebacterium urealyticum		
Corynebacterium xerosis	Cronobacter sakazakii group		
Cupriavidus pauculus group	Delftia acidovorans group		
Dermabacter hominis	Dermacoccus nishinomiyaensis		
Edwardsiella tarda	Eikenella corrodens		
Elizabethkingia meningoseptica group	Enterobacter aerogenes		
Enterobacter amnigenus	Enterobacter cloacae complex		
Enterococcus avium	Enterococcus casseliflavus		
Enterococcus durans	Enterococcus faecalis		
Enterococcus faecium	Enterococcus gallinarum		
Enterococcus hirae	Enterococcus mundtii		
Enterococcus raffinosus	Escherichia coli		
Escherichia hermannii	Escherichia vulneris		
Ewingella americana	Facklamia hominis		
Finegoldia magna	Fluoribacter bozemanae		
Fusobacterium canifelinum	Fusobacterium necrophorum		
Fusobacterium nucleatum	Gardnerella vaginalis		
Gemella haemolysans	Gemella morbillorum		
Gemella sanguinis	Granulicatella adiacens		
Haemophilus haemolyticus	Haemophilus influenzae		

Bacteria:		
Haemophilus parahaemolyticus group	Haemophilus parainfluenzae	
Hafnia alvei	Helcococcus kunzii	
Kingella denitrificans	Kingella kingae	
Klebsiella oxytoca / Raoultella ornithinolytica	Klebsiella pneumoniae	
Klebsiella variicola	Kocuria kristinae	
Kytococcus sedentarius	Lactobacillus gasseri	
Lactobacillus jensenii	Lactobacillus rhamnosus	
Lactococcus garvieae	Lactococcus lactis	
Leclercia adecarboxylata	Legionella longbeachae	
Legionella pneumophila	Leuconostoc citreum	
Leuconostoc mesenteroides	Leuconostoc pseudomesenteroides	
Listeria monocytogenes	Macrococcus caseolyticus	
Mannheimia haemolytica group	Micrococcus luteus	
Micrococcus lylae	Mobiluncus curtisii	
Moraxella sg Branhamella catarrhalis*	Moraxella sg Moraxella nonliquefaciens*	
Moraxella sg Moraxella osloensis*	Morganella morganii	
Myroides odoratimimus	Myroides odoratus	
Neisseria bacilliformis	Neisseria cinerea	
Neisseria elongata	Neisseria flavescens / subflava group	
Neisseria gonorrhoeae	Neisseria lactamica	
Neisseria meningitidis	Neisseria sicca group	
Neisseria weaveri	Nocardia brasiliensis	
Nocardia cyriacigeorgica	Nocardia farcinica group	
Nocardia nova	Nocardia otitidiscaviarum	
Ochrobactrum anthropi	Oligella ureolytica	
Oligella urethralis	Pantoea agglomerans	
Parabacteroides distasonis	Parabacteroides goldsteinii	
Parabacteroides johnsonii / merdae group	Parvimonas micra	
Pasteurella multocida	Pediococcus acidilactici	
Pediococcus pentosaceus	Peptoniphilus harei group	
Peptostreptococcus anaerobius	Plesiomonas shigelloides	
Pluralibacter gergoviae	Porphyromonas gingivalis	
Porphyromonas somerae	Prevotella bivia	
Prevotella buccae	Prevotella denticola	
Prevotella intermedia	Prevotella melaninogenica	

Bacteria:		
Propionibacterium acnes	Proteus mirabilis	
Proteus vulgaris group	Providencia rettgeri	
Providencia stuartii	Pseudomonas aeruginosa	
Pseudomonas fluorescens group	Pseudomonas oryzihabitans	
Pseudomonas putida group	Pseudomonas stutzeri	
Ralstonia pickettii	Rhizobium radiobacter	
Rothia aeria	Rothia dentocariosa	
Rothia mucilaginosa	Salmonella sp**	
Serratia fonticola	Serratia liquefaciens	
Serratia marcescens	Serratia odorifera	
Serratia plymuthica	Serratia rubidaea	
Sphingobacterium multivorum	Sphingobacterium spiritivorum	
Sphingomonas paucimobilis group	Staphylococcus aureus	
Staphylococcus auricularis	Staphylococcus capitis	
Staphylococcus caprae	Staphylococcus carnosus	
Staphylococcus cohnii	Staphylococcus delphini	
Staphylococcus epidermidis	Staphylococcus equorum	
Staphylococcus felis	Staphylococcus haemolyticus	
Staphylococcus hominis	Staphylococcus intermedius	
Staphylococcus lentus	Staphylococcus lugdunensis	
Staphylococcus pasteuri	Staphylococcus pettenkoferi	
Staphylococcus pseudintermedius	Staphylococcus saccharolyticus	
Staphylococcus saprophyticus	Staphylococcus schleiferi	
Staphylococcus sciuri	Staphylococcus simulans	
Staphylococcus vitulinus	Staphylococcus warneri	
Staphylococcus xylosus	Stenotrophomonas maltophilia	
Streptococcus agalactiae	Streptococcus anginosus	
Streptococcus canis	Streptococcus constellatus	
Streptococcus dysgalactiae	Streptococcus equi	
Streptococcus gallolyticus	Streptococcus gordonii	
Streptococcus intermedius	Streptococcus lutetiensis	
Streptococcus mitis / oralis group	Streptococcus mutans	
Streptococcus parasanguinis	Streptococcus pneumoniae	
Streptococcus pyogenes	<i>Streptococcus salivarius / vestibularis</i> group	
Streptococcus sanguinis	Streptococcus sobrinus	

Bacteria:	
Streptococcus thermophilus	Sutterella wadsworthensis
Trueperella bernardiae	Turicella otitidis
Vagococcus fluvialis	Veillonella parvula group
Vibrio parahaemolyticus	Vibrio vulnificus
Weeksella virosa	Yersinia enterocolitica
Yersinia frederiksenii	Yersinia intermedia
Yersinia kristensenii	Yersinia pseudotuberculosis
* = subgenus	
$sp^{**} = species$	

Yeasts:	
Candida albicans	Candida boidinii
Candida dubliniensis	Candida duobushaemulonii
Candida famata	Candida glabrata
Candida guilliermondii	Candida haemulonis
Candida inconspicua	Candida intermedia
Candida kefyr	Candida krusei
Candida lambica	Candida lipolytica
Candida lusitaniae	Candida metapsilosis
Candida norvegensis	Candida orthopsilosis
Candida parapsilosis	Candida pararugosa
Candida pelliculosa	Candida tropicalis
Candida valida	Candida zeylanoides
Cryptococcus gattii	Cryptococcus neoformans var grubii*
Cryptococcus neoformans var neoformans*	Cyberlindnera jadinii
Geotrichum candidum	Geotrichum capitatum
Kloeckera apiculata	Malassezia furfur
Malassezia pachydermatis	Pichia ohmeri
Rhodotorula mucilaginosa	Saccharomyces cerevisiae
Trichosporon asahii	Trichosporon inkin
Trichosporon mucoides group	
* = variety	

- 2. <u>Indication(s) for use</u>: Same as intended use.
- 3. <u>Special conditions for use statement(s):</u>

The MALDI Biotyper CA System is for prescription use only in accordance with 21 CFR

801.109.

4. Special instrument requirements:

Mass Spectrometer: microflex LT/SH mass spectrometer

Target Plates: US IVD 48 Spot Target

Reagents:

- US IVD Bacterial Test Standard (BTS)
- US IVD HCCA portioned (α-Cyano-4-hydroxycinnamic acid)

Database: MALDI Biotyper for Clinical Applications (MBT-CA)

Software:

• MBT-CA System Software Package:

MBT-CA System client software displaying the user interface

MBT-CA System Server

MBT-CA System DB Server

• flexControl Software Package:

GTPS firmware

flexControl acquisition software

I. Device Description:

The MBT-CA System consists of the microflex LT/SH mass spectrometer, reference library, kit reagents (US IVD HCCA, US IVD Bacterial Test Standard), US IVD 48 Spot Target or MBT Biotarget 96 US IVD plate, and software. The MALDI Biotyper CA System with closed safety covers is a Class 1 Laser product. With the safety cover opened it becomes a Class 4 Laser product. The laser is a 337 nm fixed focus, nitrogen laser. The MALDI Biotyper CA System is also referred to as the MBT-CA System.

The reference library, MALDI Biotyper for Clinical Applications, includes type strains, clinical strains and culture collection strains. The MALDI Biotyper CA System reference library was established by analyzing the type strain from each claimed species combined with 5 to 10 additional strains from the same species provided by clinical laboratories or different commercial strain collections for a total of 528 strains. Library mass spectra used for matching contain up to 70 peaks. The MALDI Biotyper for Clinical Applications is also referred to as the MBT-CA.

US IVD HCCA portioned (α -Cyano-4-hydroxycinnamic acid) is a solution that is used when processing test organisms for identification on the MALDI Biotyper CA System. US IVD HCCA is reconstituted in accordance with instructions provided using recommended solvent. 1.0 μ L of the matrix is added to the spot with the sample and allowed to dry.

US IVD Bacterial Test Standard (BTS) is an in-vitro-diagnostic product used for quality control and validation of the microflex LT/SH mass spectrometers. US IVD BTS contains a manufactured extract of *Escherichia coli* DH5 alpha that demonstrates a characteristic peptide and protein profile mass spectrum, when tested on the MALDI Biotyper CA System. US IVD BTS is spiked with two additional proteins that extend the upper boundary of the mass range of the US IVD BTS. The overall mass range covered by US IVD BTS is 3.6 to 17 kDa.

US IVD 48 Spot Target plates are reusable steel plates which have been developed for the

preparation and identification of test organisms using the MALDI Biotyper CA System. The target allows for the identification of 48 test organisms. The target has five cross-joint positions which should be used for US IVD BTS control. Target plate cleaning is performed after each run.

MALDI Biotyper CA System client software displays a user-interface which guides the user through the MALDI Biotyper CA System workflow. The MALDI Biotyper CA System client also interfaces to the flexControl software for automated acquisition of mass spectra on the microflex LT/SH instrument.

The MALDI Biotyper CA System server communicates with the MALDI Biotyper CA System client and the MBT-DB server. It performs preprocessing on acquired spectra, and matches peaks lists against the Main Spectrum (reference pattern, (MSP)) for matching and calculates the score value (log (score)).

The MBT-DB server stores all information for the MALDI Biotyper CA System. The MBT-DB maintains spectra data (creation information and mass/intensity lists), project data (results of defined and executed runs), method data (parameter lists for spectra preprocessing and identification), user management data, reference patterns and other peak lists plus additional maintenance data.

GTPS firmware communicates with the flexControl PC software, controls and monitors the vacuum, moves the sample carrier and performs the docking of the target plate, controls and monitors high voltages in the ion source, generates trigger signals, and monitors instrument status.

The flexControl acquisition software communicates with the MALDI Biotyper CA System client, loads automatic run jobs, communicates with the GTPS firmware, communicates with the laser in the microflex LT/SH instrument, sets the acquisition parameters in the digitizer and reads the acquired data from the digitizer, performs automated data acquisition, evaluates acquired spectra, adjusts the laser power during automatic data acquisition, performs a recalibration of the time-of-flight to mass transformation, stored acquired spectra on disk and performs source cleaning. The flexControl software does not display a user interface.

The optional Honeywell (Hyperion 1300g) Barcode Reader USB cable is connected to the MALDI Biotyper CA System computer. The barcode reader scans the unique ten-digit target ID which appears in the Target ID box on the target plate. After the target ID has been entered, the a new Run page opens and the ten-digit target ID appears as the Plate Id and is appended to the Run name. Sample identifications are entered into the computer corresponding to the target plate position for that run.

J. Substantial Equivalence Information:

1. Predicate device name(s):

MALDI Biotyper CA System

2. Predicate 510(k) number(s):

K142677

3. Comparison with predicate:

SIMILARITIES					
Characteristic	NEW DEVICE (K163536)	PREDICATE DEVICE (K142677)			
	MALDI Biotyper CA System	MALDI Biotyper CA System			
Product Code	PEX	PEX			
Intended use	The MALDI Biotyper CA System is a mass spectrometer system using matrix-assisted laser desorption/ionization - time of flight (MALDI-TOF) for the identification of microorganisms cultured from human specimens.	The MALDI Biotyper CA System is a mass spectrometer system using matrix-assisted laser desorption/ionization - time of flight (MALDI-TOF) for the identification of microorganisms cultured from human specimens.			
	The MALDI Biotyper CA System is a qualitative <i>in vitro</i> diagnostic device indicated for use in conjunction with other clinical and laboratory findings to aid in the diagnosis of bacterial and yeast infections.	The MALDI Biotyper CA System is a qualitative in vitro diagnostic device indicated for use in conjunction with other clinical and laboratory findings to aid in the diagnosis of bacterial and yeast infections.			

SIMILARITIES					
Characteristic	NEW DEVICE (K163536)	PREDICATE DEVICE (K142677)			
	MALDI Biotyper CA System	MALDI Biotyper CA System			
	Isolated colony from any patient sample	Isolated colony from any patient sample			
	source.	source.			
Sample type	Acceptable media:	Acceptable media:			
	Columbia blood agar with 5% sheep	Columbia blood agar with 5% sheep			
	blood (Gram-negative bacteria)	blood (Gram-negative bacteria)			
	Trypticase soy agar with 5% sheep	Trypticase soy agar with 5% sheep			
	blood (Gram-negative bacteria, Gram-	blood (Gram-negative bacteria, Gram-			
	positive bacteria, yeasts)	positive bacteria, yeasts)			
	Chocolate agar (Gram-negative	Chocolate agar (Gram-negative			
	bacteria, Gram-positive bacteria)	bacteria, Gram-positive bacteria)			
	MacConkey Agar (Gram-negative	MacConkey Agar (Gram-negative			
	bacteria)	bacteria)			
	Columbia CNA agar with 5% sheep	Columbia CNA agar with 5% sheep			
	blood (Gram-positive bacteria)	blood (Gram-positive bacteria)			
	Brucella Agar with 5% horse blood	Brucella Agar with 5% horse blood			
	(Gram-negative anaerobic bacteria, Gram-	(Gram-negative anaerobic bacteria, Gram-			
	(DC) anaerobic bacteria)	CDC anagradia Assurable 50(share			
	CDC anaerobe Agar with 5% sneep	CDC anaerobe Agar with 5% sheep			
	Crom positive apparabie besterie)	Grom positive anaerobic bacteria,			
	Gram-positive anaerobic bacteria)	Gram-positive anaerobic bacteria)			
	CDC anaerobe 5% sneep blood Agar	CDC anaerobe 5% sneep blood Agar			
anaerobic bacteria. Gram-positive anaerobic		with phenylethyl alcohol (Gran-negative			
bacteria)		anaerobic bacteria, Gram-positive anaerobic			
bacteria)		CDC anagraba lakad shaan blood			
CDC anaerobe laked sheep blood		CDC anaerobe laked sheep blood			
Agar with kanamycin and vancomycin		Agar with Kanamychi and Vancomychi (Gram pogetive speersbie begilli)			
	(Grann-negative anaerobic bachin)	(Gram-negative anaerobic bachin)			
	Bacteroides one esculin Agar with	Bacteroides blie esculin Agar with			
	Clostridium difficile A cor with 7%	Clostridium difficile A cor with 7%			
	closuridium difficile)	closuridium difficile Agar with 7%			
	Sabouroud Devtrose Ager (Veeste)	Subcuraud Dextrace Ager (Vesets)			
	Sabouraud-Dexirose Agar (Teasis)	Drain Heart Infusion Ager (Yeaste)			
	General chapter A consult 5	General charter A consult 5			
	Campylobacter Agar with 5	Campylobacter Agar with 5			
	Anumicrobics and 10% Sneep Blood	(Compute heater species)			
	(Campylobacter species)	(Campylobacter species)			
Bordet Gengou Agar with 15% sheep		Bordet Gengou Agar with 15% sheep			
	blood (Boraetella species)	blood (Bordetella species)			
Type of Test	Automated Mass Spectrometry System	Automated Mass Spectrometry System			
Matrix	α–Cyano-4-hydroxycinnamic acid	α-Cyano-4-hydroxycinnamic acid			

SIMILARITIES				
Characteristic	NEW DEVICE (K163536) MALDI Biotyper CA System	PREDICATE DEVICE (K142677) MALDI Biotyper CA System		
Method of Testing	Bacteria & Yeast: Direct testing If after initial analysis the log(score) is reported at <2.00, organisms may be processed using the Extraction (Ext) procedure or extended Direct Transfer (eDT, 70% aqueous formic acid) procedure. If eDT procedure still yields log (score) <2.00, organisms may be processed via Ext procedure.	Bacteria & Yeast: Direct testing If after initial analysis the log(score) is reported at <2.00, organisms may be processed using the Extraction (Ext) procedure or extended Direct Transfer (eDT, 70% aqueous formic acid) procedure. If eDT procedure still yields log (score) <2.00, organisms may be processed via Ext procedure.		
Result Reporting	Organism identification is reported with high confidence if the log(score) is ≥ 2.00 An organism identification is reported with low confidence if the log (score) is between 1.70 and <2.00.	Organism identification is reported with high confidence if the log(score) is ≥ 2.00 An organism identification is reported with low confidence if the log (score) is between 1.70 and <2.00.		
Matching AlgorithmCalculates matches by comparing a new spectrum against each single reference entry of a reference database.		Calculates matches by comparing a new spectrum against each single reference entry of a reference database.		
Mass range	2,000 - 20,000 m/z	2,000 - 20,000 m/z		
Calibration	Bruker US IVD Bacterial Test Standard	Bruker US IVD Bacterial Test Standard		
Database	MALDI Biotyper Reference Library for Clinical Applications (MBT-CA)-update	MALDI Biotyper Reference Library for Clinical Applications (MBT-CA)		

DIFFERENCES					
Characteristic	NEW DEVICE (K163536) MALDI Biotyper CA System	PREDICATE DEVICE (K142677) MALDI Biotyper CA System			
System Update	System claim additional organisms but no additional changes	N/A			
Additional Media validated:• Buffered Charcoal Yeast Extract Agar (Legionella species)• Buffered Charcoal Yeast Extract Selective Agar with polymyxin, anisomycin and vancomycin (Nocardia species)• Modified Thayer-Martin Agar (Neisseria species)		See Similarities			
MALDI-TOFBruker microflex LT/SH (benchtop)MS instrumentsBruker microflex LT/SH smart (benchtop)		Bruker microflex LT/SH (benchtop)			
MALDI Target Plates	US IVD 48 Spot Target (48 positions reusable steel targets) MBT Biotarget 96 US IVD (96 positions disposable targets)	US IVD 48 Spot Target (48 positions reusable steel targets)			

The differences do not affect substantial equivalence of the predicate and the new divice. Both systems are mass spectrometers using matrix-assisted laser desorption/ionization-time to flight (MALDI-TOF) for the identification of microorganisms cultured from human specimens. The differences noted above do not impact the intended use and do not raise new questions related to the safety and effectiveness of the test (new) device.

	Standards No.	Recognition No.(FDA)	Standards Title	Date
1	CLSI MM-18A	7-192	Criteria for Identification of Bacteria and Fungi by DNA Target Sequencing; Approved Guideline, 1 st Edition	4/28/2008
2	CLSI EP09-A2- IR	7-92	Method Comparison and Bias Estimation Using Patient Samples; Approved Guideline- -Second Edition (Interim Revision),	7/30/2010

K. Standard/Guidance Document Referenced (if applicable):

L. Test Principle:

Biochemical methods are currently the most commonly used approach for the identification of microorganisms. Organisms are tested against a range of reagents and organism identification is based on a microorganism's reaction to these reagents.

The MBT-CA System uses a different methodology for organism identification based on unique protein patterns of the microorganisms obtained from mass spectrometry. The test organism's spectrum (a pattern of mass peaks) is compared with a reference spectra library (database). Using biostatistical analysis, a probability ranking of the organism identification is generated. The probability ranking is represented as a log (score) between 0.00 and 3.00. Organism identification is reported with high confidence if the log (score) is \geq 2.00. An organism identification is reported with low confidence if the log (score) is between 1.70 and <2.00.

Organisms to be identified with the MBT-CA System should be isolated for purity on appropriate isolation media.

Direct Transfer procedure (DT): An individual colony from a subculture plate is transferred to a selected position on a US IVD 48 Spot Target plate or a MBT Biotarget 96 US IVD Target plate (targets) and overlaid with US IVD HCCA portioned (matrix). The standard solvent (50% acetonitrile / 47.5% H2O / 2.5% trifluoroacetic acid) in the matrix solution extracts proteins (mainly ribosomal proteins, which are present in high concentration) from the microorganisms. When dried matrix crystallizes, the inoculated target is ready to be analyzed on the MBT-CA System. If after initial analysis the log (score) is reported as <2.00, organisms can be processed using the extended Direct Transfer (eDT) procedure or the Extraction procedure and analysis repeated. If eDT is employed and log (score) is reported as <2.00, reanalysis via the Extraction procedure may be used.

extended Direct Transfer procedure (eDT): If DT analysis yields a (log(score) <2.00) result, an individual colony from a subculture plate may be transferred to a selected position on a target and overlaid with 70% aqueous formic acid solution. The target is air-dried and then matrix is overlaid. When dried matrix crystallizes, the inoculated target is ready to be analyzed on the MBT-CA System. If a high confidence result is not achieved (log (score) is reported at <2.00), organisms can be processed using the Extraction procedure and analysis repeated.

Extraction procedure (Ext): If after initial analysis and eDT procedure the log (score) is reported at <2.00, organisms are processed using the Extraction procedure and analysis repeated. For this purpose, isolated colonies from the subculture plate are extracted in accordance with instructions of the user manual (Ext sample preparation procedure). An aliquot of extracted material is transferred to a selected position on a target, air-dried and then overlaid with matrix material. When dried matrix crystallizes, the inoculated target is ready to be analyzed on the MBT-CA System.

MALDI-TOF Analysis: Samples are analyzed using MALDI (matrix-assisted laser desorption/ionization) TOF (time of- flight) mass spectrometry. The matrix transfers protons onto the extracted proteins and absorbs UV light. A laser in the MALDI- TOF mass spectrometer irradiates the matrix sample composite, causing evaporation and release of positively charged intact proteins and peptides ("soft" ionization technique). These ions are electrostatically accelerated over a short distance and arrive in the flight tube at a mass-dependent speed. As different proteins/peptides have different masses, ions arrive at the detector at different times (time of flight). The system measures the time (in the nanosecond range) between pulsed acceleration and the corresponding detector signal, the speed is converted into an exact molecular mass. The mass-to-charge ratio of an ion is proportional to the square of its drift time.

Highly abundant microbial proteins (mainly ribosomal proteins) result in a mass spectrum with characteristic mass and intensity distribution. It is species-specific for many bacteria and is interpreted as a molecular fingerprint to identify the test organism.

Data acquisition is controlled with MBT-CA System software. The spectrum of the unknown organism is first transformed into a peak list. Using a biostatistical algorithm, this peak list is compared to the reference peak lists of organisms in the reference library (database) and a log (score) is generated. A higher log (score) indicates a higher degree of similarity to the organism in the reference library. Organism identification is reported with high confidence if the log (score) is \geq 2.00. An organism identification is reported with low confidence if the log (score) is between 1.70 and <2.00.

The log (score) ranges, defined in the MBT-CA System, are indicative of the probability of organism identification. Results should be reviewed by a trained microbiologist and final organism identification should be based on all relevant information available. This information includes but is not limited to: Gram staining, colony morphology, growth characteristics, sample matrix, etc.

M. Performance Characteristics (if/when applicable):

- 1. Analytical performance:
 - a. Precision/Reproducibility:

Reproducibility

The reproducibility study for Gram-negative and Gram-positive aerobic bacteria, Gramnegative and Gram-positive microaerophilic bacteria, Gram-negative and Gram-positive anaerobic bacteria and yeasts was carried out to confirm day-to-day reproducibility and precision of the MALDI Biotyper CA System at different clinical study sites. The study was conducted for five days with two runs and two operators each day per clinical site.

The sources of variability tested were:

- Two operators/each clinical study site
- Three clinical study sites

- At least two target plates/each clinical study site
- Four microflex LT/SH instruments

Ten well-characterized organisms were chosen for this study and tested in duplicate via Direct Transfer procedure in accordance with product instructions. When the DT log (score) was <2.00, per product instructions, the test organism was tested following the extended Direct Transfer and Extraction procedure. The results are shown in the table below.

Blinded Test Organism Reproducibility Panel		Site A: MBT-CA ID (DT+eDT+Ext)	Site B: MBT-CA ID (DT+eDT+Ext)	Site C: MBT-CAID (DT+eDT+Ext)
Dermabacter hominis	REPRO-01	20/20 (100%)	20/20 (100%)	20/20 (100%)
Listeria monocytogenes	REPRO-02*	N/A	N/A	N/A
Nocardia farcinica group	REPRO-03	19/20 (95%)	19/20 (95%)	19/20 (95%)
Legionella pneumophila	REPRO-04	20/20 (100%)	20/20 (100%)	20/20 (100%)
Clostridium tertium	REPRO-05	20/20 (100%)	20/20 (100%)	20/20 (100%)
Facklamia hominis	REPRO-06	20/20 (100%)	20/20 (100%)	20/20 (100%)
Bacteroides caccae	REPRO-07	20/20 (100%)	20/20 (100%)	20/20 (100%)
Trueperella bernardiae	REPRO-08	20/20 (100%)	20/20 (100%)	20/20 (100%)
Neisseria meningitidis	REPRO-09	20/20 (100%)	20/20 (100%)	20/20 (100%)
Rhodotorula mucilaginosa	REPRO-10	20/20 (100%)	20/20 (100%)	20/20 (100%)
TOTAL		179/180 (99%)	179/180 (99%)	179/180 (99%)

Table: Reproducibility Study Summary:

* REPRO-02: Organism skipped as MBT-CA ID applying DT procedure was not confirmed via eDT and Ext procedure.

The data showed that 99% of all test organisms were correctly identified with a log (score) \geq 2.00 at each test site after final extraction testing confirming reproducibility of the MALDI Biotyper CA System.

The reproducibility of the MALDI Biotyper smart CA System was evaluated at one laboratory site (in-house). Reproducibility was assessed with ten well-characterized organisms that were chosen for this study and tested in duplicate via Direct Transfer procedure in accordance with MALDI Biotyper CA System instruction for use. When the DT log (score) was <2.00, per product instructions, the test organism was tested following the extended Direct Transfer and Extraction procedure. The study was conducted for five days with two runs and two operators each day per clinical site. Results are summarized below:

Blinded Test Organism	Reproducibility Panel	Site C: MBT-smart CA ID (DT+eDT+Ext)
Dermabacter hominis	REPRO-01	20/20 (100%)
Listeria monocytogenes	REPRO-02*	N/A
Nocardia farcinica group	REPRO-03	20/20 (100%)

Table: Reproducibility Study Summary:

Legionella pneumophila	REPRO-04	20/20 (100%)
Clostridium tertium	REPRO-05	20/20 (100%)
Facklamia hominis	REPRO-06	20/20 (100%)
Bacteroides caccae	REPRO-07	20/20 (100%)
Trueperella bernardiae	REPRO-08	20/20 (100%)
Neisseria meningitidis	REPRO-09	20/20 (100%)
Rhodotorula mucilaginosa	REPRO-10	20/20 (100%)
TOTAL		180/180 (100%)

* REPRO-02: Organism skipped as MBT-CA ID applying DT procedure was not confirmed via eDT and Ext procedure.

The data show that 100% of all test organisms were correctly identified with a log (score) >2.00 at each test site after final extraction testing confirming reproducibility of the MALDI Biotyper smart CA System.

b. Linearity/assay reportable range:

Not applicable, qualitative assay.

c. Traceability, Stability, Expected values (controls, calibrators, or methods):

Calibrator:

US IVD BTS is used for mass spectrum calibration and optimization as well as a performance control for the identification of microorganisms with the MALDI Biotyper CA System. US IVD BTS contains a manufactured extract of *Escherichia coli* DH5 alpha that demonstrates a characteristic peptide and protein profile mass spectrum, when tested on the MALDI Biotyper CA System. US IVD BTS is spiked with two additional proteins that extend the upper boundary of the mass range of the US IVD BTS. The overall mass range covered by US IVD BTS is 3.6 to 17 kDa. Two US IVD BTS control positions on a US IVD 48 Spot Target are selected and inoculated with US IVD BTS solution. The US IVD BTS solution is allowed to dry at room temperature and then overlaid with reconstituted US IVD HCCA portioned solution. If US IVD BTS does not meet all required performance specifications, the test run will be invalid. If US IVD BTS is not inoculated onto a target prior to processing, the test run will be invalid.

Controls:

Klebsiella pneumoniae, Haemophilus influenzae, Proteus vulgaris, Pseudomonas aeruginosa, and *Esherichia coli*, were used as controls. Of the 193 Quality Control runs conducted during the course of the method comparison study, there were five instances where a Quality Control organism failed to yield an expected result. As a result, all isolates included in that plate run were repeated using a fresh QC organism transfer. The overall plate repeat rate was 2.6% (5/193).

d. Assay cut-off:

The assay cut-off was established and reported in 510(k) K142677.

e. Detection limit:

The Limit of Detection/Dynamic Range study for Gram-negative bacteria was previously performed and reported in 510(k) K130831 (pp. 46-50). The Limit of Detection/Dynamic Range study for Gram-positive bacteria and yeasts was performed and reported in 510(k) K142677.

f. Analytical specificity:

The Interference & Specificity Study was previously established and reported in 510(k) K130831.

g. Sample stability studies:

Sample Stability after Matrix Overlay:

The sample stability study on target plates for Gram-negative bacteria was previously validated and reported in 510(k) K130831. The sample stability on target plates for Grampositive bacteria and yeasts was validated and reported in 510(k) K142677.

US IVD Bacterial Test Standard (BTS)

BTS Stability was established and described in 510(k) K130831.

HCCA portioned (Matrix) Stability

HCCA portioned (Matrix) Stability was established and described in 510(k) K130831.

Target plates stability:

Target plate stability was established and described in 510(k) K142677.

h. Carry-Over and Cross Contamination:

The carry-over, cross-contamination and target cleaning study was previously performed and reported in 510(k) K130831.

i. Influence of Agar Media

The validation of sample preparation of test organism to demonstrate that culture media inoculated onto US IVD 48 Spot targets with or without an organism present does not interfere with system performance was previously performed and reported in 510(k) K130831.

j. Organism Stability

Media and Colony Stability

In accordance with device instructions for use, primary or secondary isolation plates of recommended media [Trypticase Soy Agar with 5% sheep blood (TSA), Buffered Charcoal Yeast Extract Agar (BCYE), Buffered Charcoal Yeast Extract Selective Agar with polymyxin, anisomycin and vancomycin (BCYE/PAV) and Modified Thayer-Martin Agar (MTM))] may be held for up to 12 hours at room temperature prior to testing on the MALDI Biotyper CA System.

In the previous submission (K142677) TSA medium was only validated for Gram-positive /Gram-negative aerobic bacteria and yeast. This submission was intended to establish the suitability of this medium for Gram-positive /-negative microaerophilic and anaerobic bacteria.

Testing was conducted using one Gram-negative bacterium, one microaerophilic

Gram-negative bacterium, one Gram-positive bacterium, one microaerophilic Grampositive bacterium, two Legionella species, three Nocardia species and three Neisseria species, at varying incubation time points in replicates of eight. Additionally, thirteen clinical isolates derived from Neisseria species were tested in duplicate at varying incubation time points. Results are summarized below:

 Table: TSA Summary: Anaerobic Gram-negative/Gram-positive Bacteria (applying MBT-CA System workflow)

Testing Condition	≥2.00 ID (DT)	false ID (DT)	≥2.00 ID (DT+eDT)	false ID (DT+eDT)	≥2.00 ID (DT+eDT+Ex)	false ID (DT+eDT+Ex)
18h/37°C, 0h/25°C	16/16	0/16	NA	NA	NA	NA
18h/37°C, 12h/25°C	16/16	0/16	NA	NA	NA	NA
24h/37°C, 0h/25°C	16/16	0/16	NA	NA	NA	NA
24h/37°C, 12h/25°C	16/16	0/16	NA	NA	NA	NA
48h/37°C, 0h/25°C	16/16	0/16	NA	NA	NA	NA
48h/37°C, 12h/25°C	16/16	0/16	NA	NA	NA	NA

 Table: TSA Summary: Anaerobic Gram-negative/Gram-positive Bacteria (applying eDT and Ext procedure)

Testing Condition	>2.00 ID (eDT)	false ID (eDT)	>2.00 ID (Ext)	false ID (Ext)
18h/37°C, 0h/25°C	4/4	0/4	4/4	0/4
18h/37°C, 12h/25°C	4/4	0/4	4/4	0/4
24h/37°C, 0h/25°C	4/4	0/4	4/4	0/4
24h/37°C, 12h/25°C	4/4	0/4	4/4	0/4
48h/37°C, 0h/25°C	4/4	0/4	4/4	0/4
48h/37°C, 12h/25°C	4/4	0/4	4/4	0/4

 Table: TSA Summary: Microaerophilic Gram-negative/Gram-positive Bacteria (applying MBT-CA System workflow)

Testing Condition	≥2.00 ID (DT)	false ID (DT)	≥2.00 ID (DT+eDT)	false ID (DT+eDT)	≥2.00 ID (DT+eDT+Ext)	false ID (DT+eDT+Ext)
18h/37°C, 0h/25°C	16/16	0/16	NA	NA	NA	NA
18h/37°C, 12h/25°C	16/16	0/16	NA	NA	NA	NA
24h/37°C, 0h/25°C	16/16	0/16	NA	NA	NA	NA
24h/37°C, 12h/25°C	16/16	0/16	NA	NA	NA	NA
48h/37°C, 0h/25°C	16/16	0/16	NA	NA	NA	NA
48h/37°C, 12h/25°C	16/16	0/16	NA	NA	NA	NA

 Table: TSA Summary: Microaerophilic Gram-negative/Gram-positive Bacteria (applying eDT and Ext procedure)

Testing Condition	>2.00 ID (eDT)	false ID (eDT)	>2.00 ID (Ext)	false ID (Ext)
18h/37°C, 0h/25°C	4/4	0/4	4/4	0/4
18h/37°C, 12h/25°C	4/4	0/4	4/4	0/4
24h/37°C, 0h/25°C	4/4	0/4	4/4	0/4
24h/37°C, 12h/25°C	4/4	0/4	4/4	0/4

48h/37°C, 0h/25°C	4/4	0/4	4/4	0/4
48h/37°C, 12h/25°C	4/4	0/4	4/4	0/4

Testing Condition	≥2.00 ID (DT)	false ID (DT)	≥2.00 ID (DT+eDT)	false ID (DT+eDT)	≥2.00 ID (DT+eDT+Ext)	false ID (DT+eDT+Ext)
18h/37°C, 0h/25°C	16/16	0/16	NA	NA	NA	NA
18h/37°C, 12h/25°C	16/16	0/16	NA	NA	NA	NA
24h/37°C, 0h/25°C	16/16	0/16	NA	NA	NA	NA
24h/37°C, 12h/25°C	16/16	0/16	NA	NA	NA	NA
48h/37°C, 0h/25°C	16/16	0/16	NA	NA	NA	NA
48h/37°C, 12h/25°C	16/16	0/16	NA	NA	NA	NA

 Table: BCYE Summary: Legionella organisms (applying MBT-CA System workflow)

 Table: BCYE Summary: Legionella organisms (applying eDT and Ext procedure)

Testing Condition	>2.00 ID (eDT)	false ID (eDT)	>2.00 ID (Ext)	false ID (Ext)
18h/37°C, 0h/25°C	4/4	0/4	4/4	0/4
18h/37°C, 12h/25°C	4/4	0/4	4/4	0/4
24h/37°C, 0h/25°C	4/4	0/4	4/4	0/4
24h/37°C, 12h/25°C	4/4	0/4	4/4	0/4
48h/37°C, 0h/25°C	4/4	0/4	4/4	0/4
48h/37°C, 12h/25°C	4/4	0/4	4/4	0/4

 Table: BCYE/PAV Summary: Nocardia organisms (applying MBT-CA System workflow)

Testing Condition	≥2.00 ID (DT)	false ID (DT)	≥2.00 ID (DT+eDT)	false ID (DT+eDT)	≥2.00 ID (DT+eDT+Ex)	false ID (DT+eDT+Ext)
18h/37°C, 0h/25°C	24/24	0/24	NA	NA	NA	NA
18h/37°C, 12h/25°C	24/24	0/24	NA	NA	NA	NA
24h/37°C, 0h/25°C	24/24	0/24	NA	NA	NA	NA
24h/37°C, 12h/25°C	24/24	0/24	NA	NA	NA	NA
48h/37°C, 0h/25°C	19/24	0/24	23/24	0/24	24/24	0/24
48h/37°C, 12h/25°C	20/24	0/24	24/24	0/24	NA	NA

Table: BCYE/PAV Summary: Nocardia organisms (applying eDT and Ext procedure)

Testing Condition	>2.00 ID (eDT)	false ID (eDT)	>2.00 ID (Ext)	false ID (Ext)
18h/37°C, 0h/25°C	6/6	0/6	6/6	0/6
18h/37°C, 12h/25°C	6/6	0/6	6/6	0/6
24h/37°C, 0h/25°C	6/6	0/6	6/6	0/6
24h/37°C, 12h/25°C	5/6	0/6	6/6	0/6
48h/37°C, 0h/25°C	6/6	0/6	6/6	0/6
48h/37°C, 12h/25°C	5/6	0/6	6/6	0/6

Table: MTM Summary: Neisseria	organisms (applying	MBT-CA System workflow)
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Testing Condition	≥2.00 ID (DT)	false ID (DT)	≥2.00 ID (DT+eDT)	false ID (DT+eDT)	≥2.00 ID (DT+eDT+Ext)	false ID (DT+eDT+Ext)
18h/37°C, 0h/25°C	24/24	0/24	NA	NA	NA	NA
18h/37°C, 12h/25°C	24/24	0/24	NA	NA	NA	NA
24h/37°C, 0h/25°C	24/24	0/24	NA	NA	NA	NA
24h/37°C, 12h/25°C	24/24	0/24	NA	NA	NA	NA
48h/37°C, 0h/25°C	17/24	0/24	19/24	0/24	23/24	0/24
48h/37°C, 12h/25°C	20/24	0/24	20/24	0/24	24/24	0/24

 Table: MTM Summary: Neisseria organisms (applying eDT and Ext procedure)

Testing Condition	>2.00 ID (eDT)	false ID (eDT)	>2.00 ID (Ext)	false ID (Ext)
18h/37°C, 0h/25°C	6/6	0/6	6/6	0/6
18h/37°C, 12h/25°C	6/6	0/6	6/6	0/6
24h/37°C, 0h/25°C	6/6	0/6	6/6	0/6
24h/37°C, 12h/25°C	4/6	0/6	6/6	0/6
48h/37°C, 0h/25°C	6/6	0/6	5/6	0/6
48h/37°C, 12h/25°C	4/6	0/6	5/6	0/6

The study results confirm that the following culture media can be used on the MALDI Biotyper CA System (18 – 48 hours incubation at 37 (\pm 2) °C):

Anaerobic Gram-negative/-positive bacteria and microaerophilic Gram-negative/Gram-positive bacteria:

• Trypticase Soy Agar with 5% sheep blood (TSA)

Legionella organisms:

• Buffered Charcoal Yeast Extract Agar (BCYE)

Nocardia organisms:

• Buffered Charcoal Yeast Extract Selective Agar with polymyxin, anisomycin and vancomycin (BCYE/PAV)

The cultivation of Neisseria using Modified Thayer-Martin Agar (MTM) should not be longer than 24 hours.

Organism Stability prior to MALDI Biotyper CA System Analysis

The organism stability study for Gram-negative bacteria prior to MALDI Biotyper CA analysis was previously validated and reported in 510(k) K130831. Organism stability for Gram-positive bacteria and yeasts was previously established and reported in 510(k) K142677.

Other supportive Instrument Performance Characteristics

Mixed Culture:

The validation of mixed cultures derived from a target organism and varying amounts of non-target organisms was previously performed and reported in 510(k) K130831.

Viability Study

Viability studies with gram negative rods mixed with matrix on the target plate were not performed. The user is advised to consider all samples, microbial cultures and inoculated products as infectious. Aseptic techniques and usual precautions for handling organisms should be observed throughout the MALDI Biotyper CA System workflow according to "CLSI M29-A, *Protection of Laboratory Workers from Occupationally Acquired Infections*; Approved Guideline - Current revision". For additional handling precautions, refer to "*Biosafety in Microbiological and Biomedical Laboratories* - CDC/NIH -Latest edition".

Resolution of the Acinetobacter baumannii complex:

A study was performed to evaluate the resolution of the *Acinetobacter baumannii* complex to the species level after final extraction procedure. Sixty-six (66) clinical isolates of *Acinetobacter baumannii* complex (members: *A. baumannii*, *A. calcoaceticus*, *A. nosocomialis*, *A. pittii*) collected during initial Clinical Method Comparison protocol (see K130831) and identified by 16S rRNA and/or protein gene sequencing were re-tested using Direct Transfer, extended Direct Transfer and Extraction procedure in parallel. The results are summarized below:

Test sequence	High confidence ID (DT)	false ID (DT)	high confidence ID (eDT)	false ID (eDT)	high confidence ID (Ext)	false ID (Ext)
1 st Day	65/66	0/66	66/66	0/66	66/66	0/66
2 nd Day	66/66	0/66	66/66	0/66	66/66	0/66

Table: MBT-CA ID of clinical isolates from Acinteobacter baumannii complex

The study results demonstrated that the *Acinetobacter baumannii* complex cannot be completely resolved applying MBT-CA System workflow. The reference library/software was updated to report complex *Acinetobacter baumannii* and *Acinetobacter nosocomialis* under the *Acinetobacter baumannii/nosocomialis* group. After formation of an *Acinetobacter baumannii/nosocomialis* group unambiguous MBT-CA identification with high confidence (log(score) \geq 2.00) of *Acinetobacter baumannii/nosocomialis* group, *Acinetobacter calcoaceticus* and *Acinetobacter pittii* is possible if the final extraction procedure has been applied. A matching hint will be included in the package insert which contains the following alert: The displayed species should be considered a member of the *Acinetobacter baumannii* complex. For organisms identified by the MBT-CA System as Acinetobacter calcoaceticus, Acinetobacter pittii or Acinetobacter baumannii/nosocomialis group the full Extraction procedure (Ext) is mandatory for secure species differentiation.

Cultivation of yeast organisms at 37 °C:

A study was performed showing the general applicability of the MBT-CA System workflow for identification of yeasts cultivated at 37 $(\pm 2)^{\circ}$ C. Testing was conducted using three yeast species at varying incubation time points in replicates of eight. After initial testing, isolates were further tested at room temperature after twelve hours post-incubation. The results are summarized below:

Evaluation of cultivation time for yeast of	organisms on TSA me	edia at 37 °C (applying	g MBT-CA
System workflow)			

Testing Condition	≥2.00 ID (DT)	false ID (DT)	≥2.00 ID (DT+eDT)	false ID (DT+eDT)	≥2.00 ID (DT+eDT+Ext)	false ID (DT+eDT+Ext)
18h/37°C, 0h/25°C	4/24	0/24	7/24	0/24	24/24	0/24
18h/37°C, 12h/25°C	6/24	0/24	24/24	0/24	NA	NA
24h/37°C, 0h/25°C	10/24	0/24	14/24	0/24	24/24	0/24
24h/37°C, 12h/25°C	12/24	0/24	23/24	0/24	24/24	0/24
48h/37°C, 0h/25°C	9/24	0/24	13/24	0/24	24/24	0/24
48h/37°C, 12h/25°C	9/24	0/24	20/24	0/24	24/24	0/24

The study results confirmed the acceptability of $37 (\pm 2)^{\circ}$ C cultivation of yeasts and sample colony stability of up to 12 hours.

Biological / Technical Equivalency Study - MBT-CA smart System:

A study was performed to demonstrate equivalence of the MBT-CA output when using MALDI-TOF mass spectrometers equipped with smartbeam laser technology and nitrogen laser technology.

A panel of thirty four species which are part of the MBT-CA library were measured, with the 34 species representing Gram negative, Gram positive, and yeast. All three sample preparation techniques (DT, eDT, Ext) were used in parallel (eDT and Ext were always additionally performed independent from the DT result). Each sample preparation technique (DT, eDT, Ext) was spotted eight times onto the MALDI target. The measurement was performed on two nitrogen laser instruments and three smart laser instruments.

Overall 4080 spectra were collected in this study. The performance of all spectra for each single species, each single sample prep (DT, eDT and Ext), each MBT-CA result (no ID "red", low confidence ID "yellow", high confidence ID "green") for each instrument type is summarized in the table below.

Table: Biological / Technical Equivalency Study Results; Overall performance of 4080 measured samples. "High confidence ID" (green), "low confidence ID" (yellow) and "no ID" (red) are shown.

	Nitr	ogen	(544)	Sm	nart (8	316)		Nitrogen (544)			Smart (816)		
DT	187	72	285	300	117	399	DT	34.4%	13.2%	52.4%	36.8%	14.3%	48.9%
eDT	37	61	446	63	120	633	eDT	6.8%	11.2%	82.0%	7.7%	14.7%	77.6%
Ext	2	3	539	0	13	803	Ext	0.4%	0.6%	99.1%	0.0%	1.6%	98.4%
Σ	226	136	1270	363	250	1835	Σ	13.8%	8.3%	77.8%	14.8%	10.2%	75.0%

The study results demonstrated equivalence between the Nitrogen Laser System and the Smartbeam Laser System.

Equivalence Study MBT Biotarget 96 US IVD (Nitrogen Laser):

Studies were performed to verify and validate the use of the MBT Biotarget 96 US IVD in conjunction with the MBT-CA System (Nitrogen Laser). The study goal was to show the equivalency between the cleared US IVD MSP 48 Target Polished Steel and the new MBT Biotarget 96 US IVD. The functionality and performance of the MBT-CA System using the MBT Biotarget 96 US IVD under varying conditions was shown. Test runs were performed always in parallel using US IVD MSP 48 Target Polished Steel plates. The following studies were performed:

Repeatability and Precision:

The repeatability and precision study of the MBT Biotarget 96 US IVD was evaluated using three different testing-operators, three unique Biotarget 96 lots, three unique HCCA lots, three unique BTS lots and two unique MBT-CA Systems. The tests were performed on six working-days with two assay runs per day (microbial samples prepared as triplicate). The data showed 100% of test organisms were correctly MBT-CA identified at high confidence level [log (score) ≥ 2.0] applying the MBT-CA System workflow (combination of DT, eDT and Ext procedure). The rate of MBT-CA System false identification was 0% for all test samples after final sample preparation procedure.

The study results demonstrate that both target types performed similarly.

Tolerance Range (Dynamic Range):

The Limit of Detection study was designed to establish the estimated dynamic range of sample size of Gram-negative and Gram-positive bacteria and yeast enabling automated mass spectra acquisition and species-identification using the Biotarget 96. Ten different species were analyzed representing Gram positive, Gram negative bacteria and yeast. All sample preparation techniques (DT, eDT and Ext) were conducted. Up to five dilutions steps were performed for each scenario. Cell density and cell concentration were estimated by measuring the optical density of the suspension at a wavelength of 600 nm.

The rate of MBT-CA System false identifications at low confidence level [log(score) \geq 1.70] was 0%. The estimated dynamic range (limit of detection) of bacteria and yeast onto the MBT Biotarget 96 US IVD / US IVD MSP 48 Target Polished Steel required for MBT-CA System identification did not show significant differences.

Study results demonstrated that the estimated dynamic range for the direct (or cell equivalents using extraction) technique is 5×10^5 cells/uL.

Sample Stability prior to Matrix application:

This study was conducted to assess sample stability of microbial material (bacteria and yeasts isolates) on Biotarget 96 prior to addition of HCCA solution. Five species were

analyzed on two instruments. Measurement was applied directly after matrix application and after 15, 30, 60 and 120 minutes and was carried out at ambient temperature (23 ± 2) °C to simulate working conditions. All sample preparation techniques (DT, eDT and Ext) were conducted. Each sample was spotted and measured eight times. For comparison, evaluation of sample stability was performed in parallel using Steel Targets. The rate of MBT-CA System false identifications was 0% for all tests.

The study results demonstrate that both target types performed similarly.

Sample Stability post Matrix application:

This study was conducted to access the stability of the transferred microbial samples after addition of HCCA solution on a Biotarget 96 / Steel Target the MBT-CA workflow (DT, eDT, Ext) was tested in parallel. Two temperature conditions were analyzed in this study (21°C and 25°C). All measurements were conducted using five different species on two MALDI instruments. All sample preparation techniques (DT, eDT and Ext) were performed and each sample was spotted eight times on both target types. The ready prepared sample stability was analyzed immediately after matrix application (standard condition) and after 4h, 8h and 24h of aging.

No MBT-CA false identifications occurred. The study results demonstrate that both target types performed similarly.

Validation Study - Validation of 50 Representative Claimed Species:

This study was conducted to validate the Biotarget 96 performance using 50 different organisms 22 Gram-negative aerobic/anaerobic bacteria, 22 Gram-positive aerobic/anaerobic bacteria and six which are content of the FDA cleared MBT-CA System software and reference library. Single sample preparations for each of the test organisms were carried out using MBT-CA System preparation procedures (DT, eDT and Ext) in parallel. For comparison purposes, this validation study was performed in parallel using six steel Targets. In this study 50 claimed species representing Gram negative aerobic / anaerobic, Gram positive aerobic / anaerobic bacteria and yeast were analyzed. All sample preparation techniques (DT, eDT and Ext) were conducted. The rate of MBT-CA false identifications was 0% for this test. The overall identification rate of test samples at high confidence level (log (score) \geq 2.0) was 100%.

The study results demonstrate that both target types performed similarly.

Technical Study - Mass Accuracy/Target Edge Effects:

This study was designed for validation of Biotarget 96 in terms of "mass accuracy" using BTS, MBTCA System software and reference library and at least two microflex LT/SH mass spectrometer systems. Targets were prepared with dissolved BTS, dried and then overlaid with HCCA solution.

Two parameters were evaluated during this study: Target Flatness (the mass accuracy was used as an indicator for target flatness) and edge effects (comparison of mass accuracy from edge positions vs inner target positions). For evaluation, one prominent reference protein (e.g. 6255.4 m/z) within the mass range of the BTS was analyzed concerning mass reproducibility. Additionally, the log (scores) of each BTS spectrum were calculated as an indicator of identification performance.

Equivalence Study MBT Biotarget 96 US IVD (Smartbeam Laser):

The study design of this study was identical to the previous section (Equivalence Study MBT Biotarget 96 US IVD (Nitrogen Laser)).

The following six studies were performed:

- Repeatability / Precision
- Tolerance range (dynamic range) limit of detection (LOD)
- Sample stability prior to matrix application
- Sample stability post matrix application
- Mass Accuracy / Target Edge Effects
- Identification of 50 FDA cleared organisms according to MBT-CA System workflow.

Generally it can be stated that the technical performance between the MBT Biotarget 96 US IVD compared to the US IVD MSP 48 Target Polished Steel was comparable.

Nocardia Study

This study was designed to verify and validate the measurement and identification of different strains and different species of the genus *Nocardia* by using the MBT-CA System as well as the MBT smart CA System in parallel. In addition the study was performed using the US IVD MSP 48 Target Polished Steel and the MBT Biotarget 96 US IVD in parallel. The results of this study can be evaluated as analytical study for identification performance of the MBT-CA System for the genus *Nocardia* and additionally as demonstration of equivalence between the "MBT-CA and MBT smart CA Systems" as well as equivalence of the "MBT Biotarget 96 US IVD and US IVD MSP 48 Target Polished Steel".

Analytical Studies:

- Measurement of 30 strains covering 6 Nocardia species.
- DT, eDT and Ext was used for sample preparation.
- Each sample preparation technique was spotted 8 times on the targets.
- Each spot was measured twice.
- MALDI targets "US IVD MSP 48 Target Polished Steel" and "MBT Biotarget 96 US IVD" was used.
- Measurement on MBT smart CA System and MBT-CA System.
- Nocardia were measured after 24h, 48h and after 5 days of cultivation.

The identification performance of *Nocardia* using the standard identification MBT-CA workflow is shown in the diagram below. The general log(score) distribution was similar and the equivalence for all examined components (MBT-CA System, MBT smart CA System, MBT Biotarget 96 US IVD, and US IVD MSP 48 Target Polished Steel) was shown.



Table: An example all 5760 log (scores) after 48h cultivation

From 5760 spectra derived log(scores) after 48h cultivation overlaid. Each "log(score) lane" represents 480 single log(scores).

Data showed that the MBT smart System and the MBT System, as well as the MBT Biotarget 96 US IVD and US IVD MSP 48 Target Polished Steel had equivalent performance

Comparison studies:

Method comparison with predicate device:

Not applicable. Refer to the Clinical Studies section of this document.

Matrix comparison:

Not applicable

Clinical studies:

Clinical Sensitivity:

Challenge Panel

To demonstrate intra-laboratory performance, a challenge panel of 46 organisms (12 Gram-positive aerobic bacteria, eight Gram-negative aerobic bacteria, four Gram-positive microaerophilic bacteria, five Gram-negative microaerophilic bacteria, 10 Gram-positive anaerobic bacteria, four Gram-negative anaerobic bacteria, three yeasts) was tested at three study sites. All of the 46 organisms included in the panel were selected from stored organisms tested during the clinical study. A reference laboratory prepared the panel. Organism identifications were blinded to test sites. Each site tested the challenge panel member via Direct Transfer, extended Direct Transfer and Extraction procedure in parallel. Results are summarized below:

Table: Challenge Panel Study Summary

Test presedure	Site A*	Site B**	Site C***		
Test procedure	MALDI Biotyper CA System				
DT method	43/45 (96%)	38/44 (86%)	36/41 (88%)		
eDT method	44/45 (98%)	36/44 (82%)	38/41 (93%)		
Ext method	43/45 (96%)	36/44 (82%)	39/41 (95%)		
MBT-CA workflow	45/45 (100%)	40/44 (91%)	40/41 (98%)		

* One (1) sample was not identified due to isolate failure to grow.

** Two (2) samples were not identified due to isolate failure to grow.

*** Five (5) samples were not identified due to isolate failure to grow.

Method Comparison:

To demonstrate performance of the MALDI Biotyper CA (MBT-CA) System and the MALDI Biotyper smart CA (MBT-smart CA), a method comparison study was performed at four US clinical test sites and in-house laboratory. Fresh and stored organisms were tested on the MALDI Biotyper CA System in accordance to manufacturer's instructions for use. All organisms included in the study were sub-cultured for purity. Testing on the MBT-CA System was done from a freshly isolated colony.

All organisms included in the study and tested by US study sites were also sub-cultured onto an agar slant or appropriate media for isolation and shipped to the study reference laboratory. The reference laboratory stored all organisms included in the study and when requested, sent organisms to the sequencing reference laboratory for 16S rRNA or ITS sequencing and protein gene sequencing when requested. In-house laboratory and some testing sites processed their own isolates to the sequencing reference laboratory. The results are summarized below:

Table: Overall Performance

Overall Performance								
	REF	ERENCE ALGORIT	НM					
MBT-CA RESULT	high resolution low resolution species species / genus Negative		Total					
Organism ID ≥ 2.0 (High Confidence)	1906	131	18 ^{1) + 2)}	2055				
Organism ID (≥1.7; <2.0) (Low Confidence)	23	4	5 ^{3) + 4)}	32				
- INCORRECT MBT-CA ID (≥1.7) - NO ID (<1.7)	3 ⁵⁾	1 ⁵⁾	n/a	4				
Total	1932	136	23	2091				

Posi	tive	
high	high & low	Negative
confidence ID	confidence ID	
98.5%	99.8%	n/a

Discordant results:

1) MBT-CA Organism ID >2.0; Correct Genus ID - Incorrect Species ID (17 isolates); Reference Method reported a different organism.

2) MBT-CA Organism ID >2.0; Incorrect Genus ID (4 isolates); Reference Method reported a different organism.

3) MBT-CA Organism ID (≥1.7; <2.0); Correct Genus ID - Incorrect Species ID (3 isolates); Reference Method reported a different organism.

4) MBT-CA Organism ID (\geq 1.7; <2.0); Incorrect Genus ID (1 isolate); Reference Method reported a different organism.

5) MBT-CA Organism ID <1.7; no ID (4 isolates); Reference Method reported a species from the included in the MBT-CA Reference Library.

Con Gr	Correct: rrect: Sp roup / C	Genus I Decies ID Complex	D) or ID	C Inco Gi	Correct: Genus ID Incorrect: Genus ID no Incorrect: Species ID or Incorrect: Genus ID no Group / Complex ID Incorrect: Genus ID Incorrect: Genus ID			Incorrect: Genus ID			ID		
MBT ≥2	г-са 2.0	MB⊺ ≥1.7 t	Г-СА о <2.0	MB1 ≥2	г-са 2.0	MB⊺ ≥1.7 t	MBT-CA MBT-CA MBT-CA ≥1.7 to <2.0		MB1 <1	Г-СА L.7			
				RES	OLUTIO	N REFE	RENCE A	LGORITH	HM				
high	low	high	low	high	low	high	low	high	low	high	low	high	low
1906	131	23	4	5	9	2	2	0	4	0	1	3	1
91.2%	6.3%	1.1%	0.2%	0.2%	0.4%	0.1%	0.1%	0.0%	0.2%	0.0%	0.0%	0.1%	0.0%
20	37	2	7	1	4	4 4 1		4	1				
97.	4%	1.2	9%	0.6	7%	0.1	9%	0.1	9%	0.0	5%	0.1	9%

Correct: Genus ID Correct: Species ID or Group / Complex ID	Correct: Genus ID Incorrect: Species ID or Group / Complex ID	Incorrect: Genus ID	no ID
2064	18	5	4
98.7%	0.86%	0.24%	0.19%

Performance BACTERIA							
MBT-CA RESULT	REF	ERENCE ALGORIT	НМ				
	high resolution low resolution species species / genus Negative						
Organism ID ≥ 2.0 (High Confidence)	1827	130	21	1978			
Organism ID (≥1.7; <2.0) (Low Confidence)	21	5	4	30			
- INCORRECT MBT-CA ID (≥1.7) - NO ID (<1.7)	2	1	n/a	3			
Total	1850	136	25	2011			

					Positive								
							n	high esolution		high & resolu	tion	Neg	ative
							g	98.54% 99.85%		5%	n/a		
Correct: Genus ID Correct: Species ID or Group / Complex ID			D) or ID	Correct: Genus ID Incorrect: Species ID or Group / Complex ID			Incorrect: Genus ID				no ID		
MB1 ≥2	Г-СА 2.0	MB⊺ ≥1.7 t	r-CA o <2.0	MB1 ≥2	Г-СА 2.0	MBT ≥1.7 to	-CA o <2.0	MBT-CA MBT-CA ≥2.0 ≥1.7 to <2.0		MBT-CA <1.7			
RESOLUTION REFERENCE ALGORITHM													
high	low	high	low	high	low	high	low	high low		high	low	high	low
1829	131	21	4	5	9	2	2	0	4	0	1	2	1
90.9%	6.5%	1.0%	0.2%	0.2%	0.4%	0.1%	0.1%	0.0%	0.2%	0.0%	0.0%	0.1%	0.0%
19	60	2	5	14		4		4		1		3	
97.	5%	1.2	4%	0.7	0%	0.2	0%	0.20%		0.0	5%	0.15%	
Correct: Genus ID Correct: Species ID or Group / Complex ID		Correct: Genus ID Incorrect: Species ID or Group / Complex ID		D D or ID	Incorrect: Genus ID			ID	no	ID			
1985			18				5				3		
	98.	7%			0.9	0%			0.2	25%		0.15%	

Table: Overall Performance – Gram Negative Bacteria:

Performance Gram negative bacteria								
	REF	ERENCE ALGORIT	HM					
MBT-CA RESULT	high resolution species	high resolution low resolution species species / genus Negative		Total				
Organism ID ≥ 2.0	728	26	7	761				
(High Confidence)	720	20	,	/01				
Organism ID (≥1.7; <2.0)	0	0	4	4				
(Low Confidence)	U	U U	4	4				
- INCORRECT MBT-CA ID (≥1.7)	0	0	n/a	0				
- NO ID (<1.7)	U	Ŭ	ny a	v				
Total	728	26	11	765				

Posi	Positive				
high confidence ID	high & low confidence ID	Negative			
100.0%	100.0%	n/a			

Correct: Genus ID Correct: Species ID or Group / Complex ID			Correct: Genus ID Incorrect: Species ID or Group / Complex ID			Incorrect: Genus ID				no ID			
MBT-CA MBT-CA ≥2.0 ≥1.7 to <2.0		MBT-CA MBT ≥2.0 ≥1.7 t		Г-СА o <2.0	MBT-CA ≥2.0		MBT-CA ≥1.7 to <2.0		MBT-CA <1.7				
	RESOLUTION REFERENCE ALGORITHM												
high	low	high	low	high	high low high low high low high low			high	low				
728	26	0	0	3	0	2	1	0	4	0	1	0	0
95.2%	3.4%	0.0%	0.0%	0.4%	0.0%	0.3%	0.1%	0.0%	0.5%	0.0%	0.1%	0.0%	0.0%
75	54	()	3		3		2	1	1	L	()
98.	6%	0.0	0%	0.3	0.39% 0.39%		0.52%		0.13%		0.00%		
Correct: Genus ID Correct: Species ID or Group / Complex ID			Correct: Genus ID Incorrect: Species ID or Group / Complex ID			Incorrect: Genus ID				no	ID		
	7	54		6				5			0		
	98.	.6%			0.7	'8%			0.6	5%		0.00%	

Performance Gram positive bacteria								
	REF	HM						
MBT-CA RESULT	high resolution species	high resolution low resolution Nega		Total				
Organism ID ≥ 2.0	1101	105	11	1017				
(High Confidence)	1101	105	11	1217				
Organism ID (≥1.7; <2.0)	21		1	26				
(Low Confidence)	21	4	L	20				
- INCORRECT MBT-CA ID (≥1.7)	2	1	n/a	2				
- NO ID (<1.7)	2	T	11/d	5				
Total	1124	110	12	1246				

Table: Overall Performance – Gram Positive Bacteria:

Posi			
high	high & low	Negative	
confidence ID	confidence ID		
97.7%	99.8%	n/a	

Correct: Genus ID Correct: Species ID or Group / Complex ID		Correct: Genus ID Incorrect: Species ID or Group / Complex ID			Incorrect: Genus ID				no ID				
MBT-CA MBT-CA ≥2.0 ≥1.7 to <2.0		MBT-CA MBT-CA ≥2.0 ≥1.7 to <2.0		Г-СА o <2.0	MBT-CA ≥2.0		MBT-CA ≥1.7 to <2.0		MBT-CA <1.7				
	RESOLUTION REFERENCE ALGORITHM												
high	low	high	low	high	low	high	low	high	low	high	low	high	low
1101	105	21	4	2	9	0	1	0	0	0	0	2	1
88.4%	8.4%	1.7%	0.3%	0.2%	0.7%	0.0%	0.1%	0.0%	0.0%	0.0%	0.0%	0.2%	0.1%
12	06	2	5	11		1		()	()		3
96.	8%	2.0	1%	0.88%		0.08%		0.00%		0.00%		0.24%	
Correct: Genus ID Correct: Species ID or Group / Complex ID		Correct: Genus ID Incorrect: Species ID or Group / Complex ID			Incorrect: Genus ID			ID	no	ID			
1231		12			0			3					
	98.	.8%			0.9	6%			0.0	0%		0.24%	

Table: Overall Performance - Yeast:

Performance YEAST								
	REF	НМ						
MBT-CA RESULT	high resolution species	low resolution species / genus	Negative	Total				
Organism ID ≥ 2.0 (High Confidence)	77	0	0	77				
Organism ID (≥1.7; <2.0) (Low Confidence)	2	0	0	2				
- INCORRECT MBT-CA ID (≥1.7) - NO ID (<1.7)	1	0	n/a	1				
Total	80	0	0	80				

Posi		
high	high & low	Negative
confidence ID	confidence ID	
96.3%	98.8%	n/a

Correct: Genus ID Correct: Species ID or Group / Complex ID			Correct: Genus ID Incorrect: Species ID or Group / Complex ID				Incorrect: Genus ID				no ID		
MBT-CA MBT-CA ≥2.0 ≥1.7 to <2.0		MBT-CA ≥2.0		MBT-CA ≥1.7 to <2.0		MBT-CA ≥2.0		MBT-CA ≥1.7 to <2.0		MBT-CA <1.7			
	RESOLUTION REFERENCE ALGORITHM												
high	low	high	low	high	low	high	low	high	low	high	low	high	low
77	0	2	0	0	0	0	0	0	0	0	0	1	0
96.3%	0.0%	2.5%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	1.3%	0.0%
7	7	2	2	0		0		()	()	1	L
96.	3%	2.5	0%	0.00%		0.00%		0.00%		0.00%		1.25%	
Correct: Genus ID Correct: Species ID or Group / Complex ID			Correct: Genus ID Incorrect: Species ID or Group / Complex ID			In	Incorrect: Genus ID			no	ID		
79		0			0			1					
	98.	8%			0.0	0%			0.0	0%		1.25%	

Interpretation of Bruker Performance Report "Single Species":

The performance table of the single species *Arthrobacter cumminsii* was selected exemplarily to illustrate the meanings of the single sectors within the table:

Arth	Arthrobacter cumminsii									
	REFER	ENCE ALGORITHM								
MBT-CA RESULT	high resolution species	low resolution species / genus	Negative	Total						
Organism ID ≥ 2.0 (High Confidence)	4	8#(1)	0	12						
Organism ID (≥1.7; <2.0) (Low Confidence)	1	1	0	2						
- INCORRECT MBT-CA ID (≥1.7) - NO ID (<1.7)	0	0		0						
Total	5	9	0	14						
# One isolate was identified with high confidence by eDT procedure only. Matching hint is included in the labeling.										

	REFER	ENCE ALGORITHM							
MBT-CA RESULT	high resolution species	low resolution species / genus	Negative	Total					
Organism ID ≥ 2.0 (High Confidence)	I	II	E						
Organism ID (≥1.7; <2.0) (Low Confidence)	Ш	IV	F						
- INCORRECT MBT-CA ID (≥1.7) - NO ID (<1.7)	G	н							
Total									
comment field (J)									

The MBT-CA results as well as the results of the reference algorithm (gold standard) were subdivided in three parts. The MBT-CA results can be interpreted as "high confidence species ID", "low confidence species ID" and (combined) "incorrect ID" and "no ID". The reference algorithm ID was subdivided in "high resolution species" if the results was unambiguous and "low resolution species / genus" if the result was not unambiguous. Additionally the result of the reference algorithm could be different from the MBT-CA result ("negative").

Ι	The MBT-CA reported "high confidence species ID" and the result of the reference algorithm was unambiguous (both reported <i>A. cumminsii</i>).
II	The MBT-CA reported "high confidence species ID" and the result of the reference algorithm was not unambiguous (the MBT-CA reported <i>A. cumminsii</i> and the reference algorithm reported <i>A. cumminsii</i> and <i>A. albus</i> equivalently).
III	The MBT-CA reported "low confidence species ID" and the result of the reference algorithm was unambiguous (both reported <i>A. cumminsii</i>).
IV	The MBT-CA reported "low confidence species ID" and the result of the reference algorithm was not unambiguous (the MBT-CA reported <i>A. cumminsii</i> and the reference algorithm reported <i>A. cumminsii</i> and <i>A. albus</i> equivalently).
Е	The MBT-CA reported "high confidence species ID" and the result of the reference algorithm was different (the MBT-CA would report <i>A. cumminsii</i> but the reference algorithm would report a different species, false positive for <i>A. cumminsii</i>).
F	The MBT-CA reported "low confidence species ID" and the result of the reference algorithm was different (the MBT-CA would report <i>A. cumminsii</i> but the reference algorithm would report a different species, false positive for <i>A. cumminsii</i>).
G	The MBT-CA reported "incorrect ID" or "no ID" and the result of the reference algorithm was unambiguous (the MBT-CA would report a different species or no ID but the reference algorithm would report <i>A. cumminsii</i> , false negative for <i>A. cumminsii</i>).
Н	The MBT-CA reported "incorrect ID" or "no ID" and the result of the reference algorithm was not unambiguous (the MBT-CA would report a different species or no ID but the reference algorithm would report <i>A. cumminsii</i> , false negative for <i>A. cumminsii</i>).
J	KOOM IOF IURINET INFORMATION.

In addition to the performance table further tables with deeper differentiation of the incorrect results were created for demonstrating the MBT-CA performance. The incorrect MBT-CA results were divided into "correct genus, incorrect species", "incorrect genus" and "no ID".

The transition, a	accumulation and	d counting of	f data are	shown	below:
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Single S	pecies Perform	ance Table		Single Species Performance Table							
	REFER	ENCE ALGORITHM				REFER	ENCE ALGORITHM				
MBT-CA RESULT	high resolution species	low resolution species / genus	Negative Total		MBT-CA RESULT	high resolution species	low resolution species / genus	Negative	Total		
Organism ID ≥ 2.0 (High Confidence)	I	П	E		Organism ID ≥ 2.0 (High Confidence)	I.	Ш	A + B + E1 + E2			
Organism ID (≥1.7; <2.0) (Low Confidence)	Ш	IV	F		Organism ID (≥1.7; <2.0) (Low Confidence)	ш	IV	C + D + F1 + F2			
- INCORRECT MBT-CA ID (≥1.7) - NO ID (<1.7)	G	Н			- INCORRECT MBT-CA ID (≥1.7) - NO ID (<1.7)	G	н				
Total					Total						

	Co G	C orrect: rrect: Sp roup / Co	Genus I Decies ID Omplex I	D) or ID	(Ince G	Correct: orrect: S roup / C	Genus II pecies I omplex	D D or ID	In	correct	no ID				
species	MB1 ≥2	T-CA 2.0	MBT-CA ≥1.7 to <2.0		MB" ≥2	MBT-CA ≥2.0		MBT-CA ≥1.7 to <2.0		т-са 2.0	MBT-CA ≥1.7 to <2.0		MBT-CA <1.7		
	RESOLUTION REFERENCE ALGORITHM														
	high	low	high	low	high	low	high	low	high	low	high	low	high	low	
	1.1	Ш	Ш	IV	Α	В	С	D	E1	E2	F1	F2	G	н	
	4	8	1	1	0	0	0	0	0	0	0	0	0	0	
	28.6%	57.1%	7.1%	7.1%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	
Arthrobacter	1	2		2	(D	(0	(D	(D	0		
cumminsii	85.	.7%	14	.3%	0.	0%	0.	0%	0.	0%	0.	0%	0.	0%	
		1	4												
		100	.0%												

	(Co G	Correct: rrect: Sp roup / Co	Genus II Decies ID omplex I	D) or ID	(Ince G	Correct: orrect: S roup / C	Genus II pecies I omplex	D D or ID	In	correct	no ID				
species	MB ≥2	T-CA 2.0	MB1 ≥1.7 t	MBT-CA ≥1.7 to <2.0		MBT-CA ≥2.0		MBT-CA ≥1.7 to <2.0		T-CA 2.0	MBT-CA ≥1.7 to <2.0		MBT-CA <1.7		
		RESOLUTION REFERENCE ALGORITHM													
	high	low	high	low	high	low	high	low	high	low	high	low	high	low	
	4	8	1	1	0	0	0	0	0	0	0	0	0	0	
	28.6%	57.1%	7.1%	7.1%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	
Arthrobacter	1	2	:	2		0		0		0		D	(D	
cumminsii	85.	.7%	14.	.3%	0.	0.0%		0%	0.	0%	0.	0%	0.	0%	
		1	4												
		100	.0%												

All incorrect or "no ID" results of the performance tables from the single species (sections E and F in the above left table) were separately considered in the fields A, B, C, D, E1, E2, F1, F2, G and H in the tables above right and below. E1 and F1 results were not observed in the entire method comparison study.

	Co Corr Gro	rrect: C ect: Spe up / Co	Genus ID ecies ID mplex II) or D	C Inco Gi	correct: (orrect: Sproup / Co	Genus II pecies II mplex II)) or D	In	correc Il	t: Ge D	enus	no ID	
Bacteria	MBT ≥2	C-CA 2.0	MBT- ≥ 1.7 to	-CA <2.0		T-CA 2.0	$MBT \ge 1.7 \text{ to}$	-CA 0 <2.0		T-CA 2.0	MB ≥1. <	Γ-CA 7 to 2.0	MB7 <1	Г-СА 7
	1.1.1.	1.	1.1.1	KESU	LUTION REFER		RENCE ALGO		bigh low		1 1.	1.	1.1.1.	1.
	nign	low	nign	low	nign	low	nign	low	nign	low	nign	low	nign	low
	1/	0	0	0	0	0	0	0	0	0	0	0	0	0
	100%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Abiotrophia defectiva	100	/	00/			0	0	1	(0	0	0/	0) 0/
	100	J%0 17	0%)	, t	J %0	0%	0		J%0	0	70	0	70
		100	0/2											
	51	0	0	0	0	0	0	0	0	0	0	0	0	0
	100%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Acinotobactor baumannii /	100 /0	1	070	070	070	0	0,0	070	070	0	070	0 /0	070)
nosocomialis group	100	1)%	0%		()%	09	6	()%	0	%	0	%
nosocomians group	100	570	070)		770	07	0) /0	0	/0	0	/0
		100	%											
	5	0	0	0	0	0	0	0	0	0	0	0	0	0
	100%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Acinetobacter	10070	5	0	0		0		0		0		0)
calcoaceticus	10	0%	0%)	()%	0%		0%		0%		0%	
		5			~		0,	-				,.		
		100	%											
	32	0	0	0	0	0	0	0	0	0	0	0	0	0
	100%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
A * , T , *,	3	2	0			0	0			0		0	()
Acinetobacter pittii	100	0%	0%		0)%	0%	6	()%	0%		0	%
	32						•							
		100	%											
	16	2	0	0	0	0	0	0	0	0	0	0	0	0
	88.9%	11.1%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Actinotignum schaalii	1	8	0			0	0			0		0	()
group	100	0%	0%)	0)%	0%	6	()%	0	%	0	%
		18												
		100	%	•				0		1				
	34	1	0	0	0	0	0	0	0	0	0	0	0	0
	97.1%	2.9%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Actinomyces europaeus	3	5	0			0	0			0		0	()
Termoniyees emopuens	100)%	0%)	0)%	0%	6	()%	0	%	0	%
		35												
		100	%	-		-		_	-	-			_	
	3	0	0	0	0	0	0	0	0	0	0	0	0	0
	100%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Actinomyces funkei		3	0			0	0	,	0			0	0	
J	100	<u>J%</u>	0%)	()%	0%	Ó	0%		0%		0%	
		3	0/											
		100	%											

	Correct: Genus ID Correct: Species ID or Group / Complex ID				C Inco G1	Correct: Genus ID Incorrect: Species ID or Group / Complex ID					t: Ge D	enus	no ID		
Bacteria	MBT ≥2	с-СА .0	MBT- ≥1.7 to	-CA <2.0	MB ≥	T-CA 2.0	$\frac{\text{MBT}}{\geq 1.7 \text{ to}}$	-CA <2.0	MB ≥	T-CA 2.0	MB ≥1.	Γ -CA 7 to	MB7 <1	Г-СА I.7	
]	RESO	LUTIO	N REFE	RENCE	ALG	ORIT	ΉM		2.0	<u> </u>		
	high	low	high	low	high	low	high	low	high	low	high	low	high	low	
	3	0	0	0	0	0	0	0	0	0	0	0	0	0	
	100%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	
Actinomyces graevenitzii	3	3	0	0		0	0			0		0	()	
	100)%	0%)	()%	0%	0	()%	0	%	0%		
		<u> </u>	0/6												
	2 0		0	0	0	0	0	0	0	0	0	0	0	0	
	100%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	
A stin survey huser acinglia	2	2	0			0	0			0		0	()	
Actinomyces nyovaginatis	100)%	0%)	()%	0%	ó	()%	0	%	0	%	
		2													
	20	1009	%	0	0	0	0	0	0	0	0	0	0	0	
	100%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	
	2	0	0/0	070	070	0	070	070	070	0	070	0	()	
Actinomyces radingae	100)%	0%		0%		0%		0%		0%		0%		
		20							0,0						
		1009	%												
	39	1	0	0	0	0	0	0	0	0	0	0	0	0	
	97.5%	2.5%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	
Actinomyces turicensis	100	0	0		(0	0	4	-	0	0	0	0%		
	40					70	07	0	, t	70	0	70	070		
		100	%												
	8	0	0	0	0	0	0	0	0	0	0	0	0	0	
	100%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	
Actinomyces urogenitalis	8	8	0		0		0		0		0		0		
	100)%	0%)	0%		0%		0%		0%		0%		
		<u> </u>	%												
	31	0	0	0	0	0	0	0	0	0	0	0	0	0	
	100%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	
Aerococcus sanguinicola	3	1	0			0	0			0		0	()	
nerococcus sunguinicolu	100)%	0%	,)	()%	0%	0	()%	0	%	0	%	
		31	2/												
	22	100	%	0	0	0	0	0	0	0		0	0	0	
	32 100%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	
	3	2	070	070	070	0	070	070	070	0	070	0	070)	
Aerococcus viridans	100)%	0%	,)	()%	09	ó	()%	0	%	0	%	
		32					•		•		•				
		100	%			n	1								
	2	4	0	0	0	0	0	0	0	0	0	0	0	0	
A (*1	33.3%	66.7%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	
Aggregatibacter	100)%	00/		ſ	0	0	6	0		0			ر %	
acunomyceiencomuuns	100	6	0%	,	(///	0%	U		//		/0	0	/0	
		1009	%												

	Co Corr Gro	rrect: C ect: Spe up / Co	Genus ID ecies ID mplex II) or D	C Inco Gi	Correct: Genus ID Incorrect: Species ID or Group / Complex ID					: t: Ge D	enus	no	ID
Bacteria	MBT ≥2	MBT-CA ≥2.0		CA <2.0	MB ≥	T-CA 2.0	$\frac{\text{MBT-CA}}{\geq 1.7 \text{ to } < 2.0}$		MB ≥	T-CA 2.0	MB′ ≥1 <	Г-СА .7 to 2.0	MB7 <1	Г-СА 1.7
			I	RESO	LUTIC	N REFE	RENCE	ALG	ORIT	THM		1	1	
	high	low	high	low	high	low	high	low	high	low	high	low	high	low
	13	0	0	0	0	0	0	0	0	0	0	0	0	0
	100%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Aggregatibacter	1	3	0			0	0	,		0		0	0	
aphrophilus	100	<u>)%</u>	0%)	()%	0%	0	()%	0	%	0	%
		13	. /											
					0	0	0	0		0		0	0	
	3	4	0	0	0	0	0	0	0	0	0	0	0	0
	42.9%	57.1%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Aggregatibacter segnis	/ 0					0	0	/		0		0	()
	100% 0%				()%	09	0	(J%	0	%	0	%
		100	0/											
	2	100	%	0	0	0	0	0	0	0	0	0	0	
	3 1000/	0	0	0	0	0	0	0	0	0	0	0	0	0
	100%	0%	0%	0% 0%		0% 0%		0% 0%		0%	0% 0%		0% 0%	
Alloiococcus otitis	10) 20/	0/			0	0	/	0%		0	0	0%	
	100	J%	0%)	(J%	0%	0	(J%	0	%	0	%
		100	0/											
	5	100	^{%0}	0	0	0	0	0	0	0	0	0	0	0
	100%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
	100 /0	070	070	070	070	0	070	0 /0	070	0.0	070	070	0 /0	070
Alloscardovia omnicolens	100%		0%		0%		0	6	(0)%	0	0	0%	
	100	5	070	,		770	07	0		570	0	/0	0	/0
		100	%											
	7	0	0	0	0	0	0	0	0	0	0	0	0	0
	100%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
	10070	7	0	0,0	0,0	0	0	0,0	070	0	070	0	()
Anaerococcus murdochii	100)%	0%)	()%	09	6	()%	0	%	0	%
		7	I				1							
		100	%											
	4	8	1	1	0	0	0	0	0	0	0	0	0	0
	28.6%	57.1%	7.1%	7.1%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
	1	2	2			0	0			0		0	()
Arthrobacter cumminsu	85.	7%	14.3	%	()%	0%	6	()%	0	%	0	%
		14												
		100	%											
	27	0	0	0	0	0	0	0	0	0	0	0	0	0
	100%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Rastaroidas assas	2	7	0			0	0			0		0	()
Ducieroides cuccue	100	0%	0%)	()%	09	6	0%		0%		0%	
		27												
		100	%											
	Co Corr Gro	errect: C rect: Spe oup / Co	Genus IE ecies ID mplex II) or D	C Inco Gi	Correct: (Orrect: Sp roup / Co	Genus II pecies II mplex I)) or D	In	correc Il	e t: Ge	enus	no	ID
--------------------------	-------------------	------------------------------------	----------------------------------	--------------	-----------------	---------------------------------------	--	----------------	---------	--------------	-----------------	----------------------	-----------	-------------
Bacteria	MBT ≥2	Г-СА 2.0	MBT- ≥1.7 to	•CA <2.0	MB ≥	T-CA 2.0	$\frac{\text{MBT}}{\geq 1.7 \text{ to}}$	-CA o <2.0	MB ≥	T-CA 2.0	MB′ ≥1. <	Г-СА .7 to 2.0	MB7 <1	Г-СА 1.7
]	RESO	LUTIC	ON REFE	RENCE	ALG	ORIT	ΉM				
	high	low	high	low	high	low	high	low	high	low	high	low	high	low
	7	1	0	0	0	0	0	0	0	0	0	0	0	0
	87.5%	12.5%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Bacteroides nordii	3	3	0			0	0			0	_	0	()
	10	0%	0%)	()%	09	0	()%	0	%	0	%
		8	o./											
	10	100	%	0	0	0	0	0	0	0	0	0	0	0
	100%	0	0%	0	0%	0%	0	0	0	0%	0	0%	0	0%
	100%	0%	0%	0%	0%	0	0%	0%	0%	0%	0%	0%	0%	0%
Bacteroides pyogenes	10	0	0%		(0)%	0	6	()%	0	0 %	0	%
	10	10	07)	()/0	07	0) /0	0	/0	0	/0
		100	%											
	7	0	0	0	0	0	0	0	0	0	0	0	0	0
	100%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
	7	7 0% 0% 0 7 100% 7				0	0			0		0	()
Bacteroides salyersiae	10	0%	0%)	()%	0%	6	()%	0	%	0	%
		7												
		100	%										-	
	19	0	0	0	0	0	0	0	0	0	0	0	0	0
	100%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Bacteroides stercoris	1	9	0			0	0			0		0	()
group	10	0%	0%)	()%	09	0	()%	0	%	0	%
		100	%	1		1	I							1
	16	1	0	0	0	0	0	0	0	0	0	0	0	0
	94.1%	5.9%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Bifidobacterium breve	10	7	0			0	0	/		0		0	()
	100	<u>0%</u> 17	0%)	()%	09	0	()%	0	%	0	%
		1/	0/											
	0	3	0	0	0	0	0	0	0	0	0	0	0	0
	0%	100%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
	070	3	0,0	070	070	0	0,0	070	070	0	070	0	070)
Clostridium beijerinckii	10	0%	0%)	()%	09	6	()%	0	%	0	%
		3		·				-						
		100	%											
	9	0	2	0	0	0	0	0	0	0	0	0	0	0
	81.8%	0%	18.2%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Clostridium hifermontans	9)	2			0	0			0		0	()
Costruium oyermentuns	81.	8%	18.2	%	()%	09	6	()%	0	%	0	%
		11												
		100	%											

	Co Corr Gro	rrect: C ect: Spoup / Co	Genus IE ecies ID mplex II) or D	C Inco Gi	Correct: (orrect: Sproup / Co	Genus II pecies II mplex I)) or D	In	correc Il	e t: Ge D	enus	no	ID
Bacteria	MBT ≥2	C-CA 2.0	$\frac{\text{MBT-}}{\geq 1.7 \text{ to}}$	-CA <2.0	MB ≥	T-CA 2.0	MBT ≥1.7 to	-CA	MB ≥	T-CA 2.0	MB′ ≥1 <	Г-СА .7 to 2.0	MB] <]	Г-СА 1.7
	1.1.1	1.	1.1.1	RESO	LUTIO	N REFE	RENCE	ALG	$\frac{ORI1}{1 \cdot 1}$	'HM	h 1.	1.	1.1.1.	1.
	nign 16	low	nign	low	nign	low	nign	low	nign	low	nign	low	nign	low
	100%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
	10070	6	0,0	070	070	0	0,0	070	070	0	070	0	0 / 0)
Clostridium butyricum	10	0%	0%)	()%	09	6	()%	0	%	0	%
		16	5											
		100	%	1		T	1							1
	14	0	0	0	0	0	0	0	0	0	0	0	0	0
	100%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Clostridium	1	4	0			0	0			0		0	()
clostridioforme group	10)%	0%)	()%	09	6	()%	0	%	0	%
		14	•								1			
		100	%											
	34	0	0	0	0	0	0	0	0	0	0	0	0	0
	100%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Clostridium innocuum	3	4	0			0	0			0		0	()
Closin alam innocaam	10)%	0%)	C)%	09	6	()%	0	%	0	%
		34	ļ											
		100	%											
	17	0	0	0	0	0	0	0	0	0	0	0	0	0
	100%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Clostridium	1	7	0			0	0			0		0	()
paraputrificum	10	0%	0%)	C)%	0%	6	()%	0	%	0	%
		17	1											
		100	%											
	37	0	0	0	0	0	0	0	0	0	0	0	0	0
	100%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
	3	7	0	I		0	0			0		0	()
Clostridium ramosum	10	0%	0%)	()%	0%	6	()%	0	%	0	%
		37	 '				1		1		1		1	
		100	%											

	Co Cori Gro	orrect: C cect: Spe oup / Co	Genus ID ecies ID mplex II) or D	C Inco Gi	orrect: (orrect: Sp coup / Co	Genus II pecies II mplex I) D or D	In	correc Il	: t: Ge D	enus	no	ID
Bacteria		Г-СА 2.0	MBT- ≥1.7 to	-CA <2.0	MB ≥	T-CA 2.0	$MBT \ge 1.7 \text{ to}$	C-CA 0 <2.0	MB ≥	T-CA 2.0	MB′ ≥1 <	T-CA .7 to 2.0	MB] <1	Г-СА 1.7
]	RESO	LUTIC	N REFE	RENCE	ALG	ORIT	THM	I			
	high	low	high	low	high	low	high	low	high	low	high	low	high	low
	52 100%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
	3	2	0			0	0)		0		0	()
Clostridium septicum	10	0%	0%	,)	0)%	09	6	()%	0	9%	0	%
		32	,											
		100	%	1		1	T	T		[
	21	0	0	0	0	0	0	0	0	0	0	0	0	0
	100%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Clastui dium aandallii	2	1	0			0	0)		0		0	()
Closiriaium soraeitti	10	0%	0%	,)	()%	0%	6	()%	0)%	0	%
		21 100%												
		100	%				I	1				1	1	1
	0	0 12 0		0	0	0	0	0	0	0	0	0	0	1
	0%	92.3%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	7.7%
Clostridium sporogenes	1	2	0			0	0)		0		0		1
/C. botulinum (group I)	92.	3%	0%	,)	()%	09	6	()%	0)%	7.′	7%
		12					•							
		92.3	%	-				-				-	-	
	39	0	0	0	0	0	0	1	0	0	0	0	0	0
	97.5%	0%	0%	0%	0%	0%	0%	2.5%	0%	0%	0%	0%	0%	0%
Clostridium tertium	3	9	()		0	1			0		0	()
	97.	5%	0	%		0%	2.5	%	()%	0)%	0	%
		3	9											
		97.	5%											
	36	0	0	0	0	0	0	0	0	0	0	0	0	0
	100%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Corynebacterium	3	6	()		0	0)		0		0	()
accolens	10	0%	0	%		0%	0%	6	()%	0)%	0	%
		3	6 0%		_									
		10												

	Co Corr Gro	errect: C ect: Spe oup / Cor	Genus ID ecies ID mplex II	or O	C Inco Gr	orrect: (orrect: S ₁ oup / Co	Genus II pecies II mplex II)) or D	In	correc Il	: t: Ge D	enus	no	ID
Bacteria	MBT ≥2	Г-СА 2.0	MBT- ≥1.7 to	CA <2.0	MB′ ≥	T-CA 2.0	$\frac{\text{MBT}}{\geq 1.7 \text{ to}}$	-CA o <2.0	MB ≥	T-CA 2.0	MB′ ≥1 <	T-CA .7 to 2.0	MB] <]	Г-СА 1.7
			I	RESOI	LUTIO	N REFE	RENCE	ALG	ORIT	THM	.	-		
	high	low 1	high	low	high	low	high	low	high	low	high	low	high	low
	83.3%	16.7%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Corvnehacterium	(<u>.</u>	()		0	0			0		0	()
afermentans group	10	0%	09	%		0%	0%	6	()%	0)%	0	%
		e	5											
		100)%						1		1		1	1
	14	4	0	0	0	0	0	0	0	0	0	0	0	0
	77.8%	22.2%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Corvenabactarium covlaga	1	8	()		0	0			0		0	()
Corynebucierium coyieue	10	0%	09	%		0%	0%	6	()%	0)%	0	%
		1	8											
		100)%						1		1			
	8	0	0	0	0	0	0	0	0	0	0	0	0	0
	100%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Corvnehacterium frenevi	8	3	()		0	0			0		0	()
eoryneouelerium greneyi	10	0%	09	%		0%	0%	6	()%	0)%	0	%
		8	3											
		100)%	[1		[1	n	1	n	Γ	n
	3	0	0	0	0	0	0	0	0	0	0	0	0	0
	100%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Corynebacterium		3	()		0	0			0		0	()
glutamicum	10	0%	09	%		0%	0%	6	()%	0)%	0	%
		3	3											
		100	0%				-			-		-	-	-
	5	0	1	0	0	0	0	0	0	0	0	0	0	0
	83.3%	0%	16.7%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Corynebacterium mucifaciens /	4	5	1			0	0			0		0	()
ureicelerivorans group	83.	3%	16.	7%		0%	0%	6	()%	0)%	0	%
		100	5 0%		-									

	Co Corr Gro	rrect: C ect: Spe up / Co	Genus ID ecies ID mplex II	or O	C Inco Gr	orrect: (orrect: Sp coup / Co	Genus II pecies II mplex I)) or D	In	correc Il	: t: Ge D	enus	no	ID
Bacteria	MBT ≥2	C-CA 2.0	MBT- ≥1.7 to	•CA <2.0	MB′ ≥	T-CA 2.0	$\frac{\text{MBT}}{\geq 1.7 \text{ to}}$	-CA o <2.0	MB ≥	T-CA 2.0	MB′ ≥1. <	Г-СА .7 to 2.0	MB7 <1	Г-СА 7
			I	RESOI	LUTIO	N REFE	RENCE	ALG	ORIT	THM		-		
	high	low	high	low	high	low	high	low	high	low	high	low	high	low
	4	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Corvnehacterium	2	1	()		0	0			0		0	()
pseudotuberculosis	100)%	09	%		0%	0%	6	()%	0	%	0	%
		2	1											
		10	0%	-			-		_				-	
	2	0	0	0	0	0	0	0	0	0	0	0	0	0
	100%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Corvanhactorium resistens	2	2	()		0	0			0		0	()
Corynebucierium resisiens	100)%	09	%		0%	0%	6	()%	0	%	0	%
		4	2								I		1	
		10	0%											
	28	1	0	1	0	0	0	0	0	0	0	0	0	0
	93.3%	3.3%	0%	3.3%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Dermahacter hominis	2	9	1	l		0	0			0		0	()
Dermabacier nominis	96.	7%	3.3	3%		0%	0%	6	()%	0	%	0	%
		3	0											
		10	0%				•							
	6	0	0	0	0	0	0	0	0	0	0	0	0	0
	100%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Enterococcus durans	e	5	()		0	0			0		0	()
	100)%	09	%		0%	0%	6	()%	0	%	0	%
		(5											
		10	0%				_						-	
	4	0	0	0	0	0	0	0	0	0	0	0	0	0
	100%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Enterococcus mundtii	2	1	()		0	0			0		0	()
Emerococcus munum	100)%	09	%		0%	09	6	()%	0	%	0	%
		4	4											
		10	0%											

	Co Corr Gro	errect: C ect: Spe oup / Co	Genus ID ecies ID mplex II	or O	C Inco Gr	orrect: (rrect: S _l oup / Co	Genus II pecies II mplex I)) or D	In	correc Il	t: Ge	enus	no	ID
Bacteria	MBT ≥2	Г-СА 2.0	MBT- ≥1.7 to	CA <2.0	MB' ≥	Г-СА 2.0	$MBT \ge 1.7 \text{ to}$	-CA o <2.0	MB ≥	T-CA 2.0	MB′ ≥1 <	T-CA .7 to 2.0	MB7 <1	Г-СА 1.7
			I	RESOI	LUTIO	N REFE	RENCE	ALG	ORIT	THM	1			
	high	low	high	low	high	low	high	low	high	low	high	low	high	low
	100%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
	3	4	()		0	0			0		0	()
Enterococcus raffinosus	10	0%	09	%		0%	09	6	()%	0	%	0	%
		3	4				•				•			
		100	0%				1	[r	I			
	25	2	0	0	0	0	0	0	0	1	0	1	0	0
	86.2%	6.9%	0%	0%	0%	0%	0%	0%	0%	3.4%	0%	3.4%	0%	0%
Escherichia hermannii	2	7	()		0	0			1		1	()
Lisener tenta nermanni	93.	1%	09	%		0%	0%	6	3	.4%	3.	4%	0	%
		2	7				•							
		93.	1%											-
	10	2	0	0	0	0	0	0	0	0	0	0	0	0
	83.3%	16.7%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Escherichia vulneris	1	2	()		0	0			0		0	()
Escherichta vanieris	10	0%	09	%		0%	09	6	()%	0	%	0	%
		1	2											
		100	0%								1			
	4	0	0	0	0	0	0	0	0	3	0	0	0	0
	57.1%	0%	0%	0%	0%	0%	0%	0%	0%	42.9%	0%	0%	0%	0%
Ewingella americana	4	1	()		0	0			3		0	()
	57.	1%	09	%		0%	0%	6	42	2.9%	0	%	0	%
		2	1											
		57.	1%											
	17	0	0	0	0	0	0	0	0	0	0	0	0	0
	100%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Facklamia hominis	1	7	()		0	0			0		0	()
	100	0%	09	%		0%	0%	6	()%	0	%	0	%
		1	7											
		100	0%											

	Co Corr Gro	errect: C ect: Spe oup / Co	Genus ID ecies ID mplex II	or O	C Inco Gr	orrect: (rrect: S _j oup / Co	Genus II pecies II omplex II)) or D	In	correc Il	: t: Ge D	enus	no	ID
Bacteria	MBT ≥2	C-CA 2.0	$\frac{\text{MBT-}}{\geq 1.7 \text{ to}}$	•CA <2.0	MB′ ≥́	Г-СА 2.0	$\frac{\text{MBT}}{\geq 1.7 \text{ to}}$	-CA 0 <2.0	MB ≥	T-CA 2.0	MB′ ≥1. <	Г-СА .7 to 2.0	MB7 <1	Г-СА 1.7
	1 • 1	1	I I I I	RESOL	JUTIO	N REFE	RENCE	ALG	ORIT	THM	1 • 1	1	1 · 1	1
	nign	10W	nign	low	nign	low	nign	10W	nign	10W	nign	low	nign	low
	100%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
	2	2	()		0	0			0		0	()
Fluoribacter bozemanae	10	0%	09	%		0%	0%	6	()%	0	%	0	%
		10	2		-									
	25	1	0	0	0	0	0	0	0	0	0	0	0	0
	96.2%	3.8%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
	2	6	()		0	0			0		0	()
Gemella morbillorum	10	0%	09	%		0%	0%	6	()%	0	%	0	%
		2	6											
		10	0%			0						0		
	4	0	0	0	0	0	0	0	0	0	0	0	0	0
	100%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Helcococcus kunzii	4	1	()		0	0			0		0	()
neteococcus kunzu	10	0%	09	%		0%	0%	6	()%	0	%	0	%
		2	1											
		10	0%	-				-		-		-		
	5	0	0	0	0	0	0	0	0	0	0	0	0	0
	100%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Kingella denitrificans	-	5	()		0	0			0		0	()
	10	0%	09	%		0%	0%	6	()%	0	%	0	%
		-	5											
		10	0%										-	-
	86	4	0	0	1	0	0	0	0	0	0	0	0	0
	94.5%	4.4%	0%	0%	1.1%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Klabsialla praumoniaa	9	0	()		1	0			0		0	()
Riebstettu pheumoniue	98.	9%	09	%	1	.1%	0%	6	()%	0	%	0	%
		9	0											
		98.	9%											

	Co Corr Gro	orrect: C cect: Spe oup / Co	Genus ID ecies ID mplex II	or O	C Inco Gr	orrect: (orrect: Sp coup / Co	Genus II pecies II omplex II)) or D	In	correc Il	: t: Ge D	enus	no	ID
Bacteria	MBT ≥2	Г-СА 2.0	MBT- ≥1.7 to	CA <2.0	MB′ ≥	T-CA 2.0	$\frac{\text{MBT}}{\geq 1.7 \text{ to}}$	-CA 0 <2.0	MB ≥	T-CA 2.0	MB′ ≥1. <	Г-СА .7 to 2.0	MB7 <1	Г-СА 7
	1 • 1	1	I · 1	RESOI		N REFE	RENCE	ALG	ORIT	THM	1 • 1	1	1 · 1	1
	nign	10W	nign	low	nign	low	nign	10W	nign	10W	nign	low	nign	low
	100%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
	()	()		0	0			0		0	()
Klebsiella variicola	10	0%	09	%		0%	0%	6	()%	0	%	0	%
		ç)											
		100)%	-							_		-	
	4	26	0	0	1	0	0	0	0	0	0	0	0	0
	12.9%	83.9%	0%	0%	3.2%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Lastobasillus gassari	3	0	()		1	0			0		0	()
Laciobaciiius gasseri	96.	8%	09	%	3	3.2%	0%	6	()%	0	%	0	%
		3	0											
		96.	8%											
	0	24	0	0	0	0	0	0	0	0	0	0	0	0
	0%	100%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Lactobacillus jonsonii	2	4	()		0	0			0		0	()
Luciobucilius jensenii	10	0%	09	%		0%	0%	6	()%	0	%	0	%
		2	4											
		100)%										r	
	39	0	0	0	0	0	0	0	0	0	0	0	0	0
	100%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Lactobacillus rhamnosus	3	9	()		0	0			0		0	()
	10	0%	09	%		0%	0%	6	()%	0	%	0	%
		3	9											
		10)%											
	18	1	0	0	0	0	0	0	0	0	0	0	0	0
	94.7%	5.3%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Leclercia adecarborylata	1	9	()		0	0			0		0	()
	10	0%	09	%		0%	0%	6	()%	0	%	0	%
		1	9											
		10)%		1									

	Co Corr Gro	errect: C ect: Spe oup / Co	Genus ID ecies ID mplex II	or D	C Inco Gi	orrect: (rrect: S _j oup / Co	Genus II pecies II omplex II)) or D	In	correc I	: t: Ge D	enus	no	ID
Bacteria	MBT ≥2	C-CA 2.0	$\frac{\text{MBT-}}{\geq 1.7 \text{ to}}$	-CA <2.0	MB' ≥	Т-СА 2.0	$\frac{\text{MBT}}{\geq 1.7 \text{ to}}$	-CA 0 <2.0	MB ≥	T-CA 2.0	MB′ ≥1. <	Γ-CA 7 to 2.0	MB7 <1	Г-СА 1.7
	1 • 1	1	<u> </u>	RESO	LUTIO	N REFE	RENCE	ALG	ORIT	THM	4 · 1	1	1 · 1	1
	high 3	low	high	low	high	low	high	low	high	low	high	low	high	low
	100%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
	3	3	()		0	0			0		0	()
Legionella longbeachae	10	0%	09	%		0%	0%	6	()%	0	%	0	%
			3											
	22	10	0%			0		0		0	0	0	0	0
	33	0	0	0	0	0	0	0	0	0	0	0	0	0
	100%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Legionella preumophila	3	3	()		0	0			0		0	()
Legionena pheamophia	10	0%	0	%		0%	0%	6	()%	0	%	0	%
		3	3											
		10	0%											
	0	3	0	0	0	0	0	0	0	0	0	0	0	0
	0%	100%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
x	3	3	()		0	0			0		0	()
Leuconostoc citreum	10	0%	0	%		0%	0%	6	()%	0	%	0	%
			3											
		10	0%		_									
	4	0	0	0	0	0	0	0	0	0	0	0	0	0
	100%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Leuconostoc	2	1	()		0	0			0		0	()
pseudomesenteroides	10	0%	09	%		0%	0%	6	()%	0	%	0	%
		10	4 0%											
	25	0	0	0	0	0	0	0	0	0	0	0	0	0
	100%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
	2	5	()		0	0			0		0	()
Listeria monocytogenes	10	0%	0	%		0%	0%	6	()%	0	%	0	%
		2	5											
		10	0%											

	Co Corr Gro	rrect: C ect: Spe up / Co	Genus ID ecies ID mplex II	or D	C Inco Gr	orrect: (orrect: Sp coup / Co	Genus II pecies II omplex II) Dor D	In	correc I	: t: Ge D	enus	no	ID
Bacteria	MBT ≥2	C-CA 2.0	$\frac{\text{MBT-}}{\geq 1.7 \text{ to}}$	-CA <2.0	MB′ ≥	T-CA 2.0	$\frac{\text{MBT}}{\geq 1.7 \text{ to}}$	-CA o <2.0	MB ≥	T-CA 2.0	MB′ ≥1. <	T-CA .7 to 2.0	MB7 <1	Г-СА 1.7
	1 . 1	1]	RESO		N REFE	RENCE	ALG	ORIT	THM	h · 1	1	1 · 1	1
	nign 13	low	nign	10W	nign	low	nign	low	nign	10W	nign	low	nign	low
	100%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Mannheimia haemolytica	1	3	()		0	0			0		0	()
group	100)%	0	%		0%	0%	6	()%	0)%	0	%
		1	3											
		10	0%				1		1		1		1	1
	4	0	0	0	0	0	0	0	0	0	0	0	0	0
	100%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Micrococcus Ivlae	2	1	()		0	0			0		0	()
	100	0%	09	%		0%	0%	6	()%	0)%	0	%
		4	1						1		1		1	
		10	0%											
	3	0	0	0	0	0	0	0	0	0	0	0	0	0
	100%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
A 4 1 1	3	3	()		0	0			0		0	()
Mobiluncus curtisii	100)%	09	%		0%	0%	6	()%	0	%	0	%
			3						1		1			
		10	0%		_									
	3	0	0	0	0	0	0	0	0	0	0	0	0	0
	100%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Neisseria hacilliformis		3	()		0	0			0		0	()
iveisseria bacingorniis	100	0%	09	%		0%	0%	6	()%	0)%	0	%
			3											
		10	0%	T		1		[1		1	n	n	n
	1	1	0	0	0	0	0	0	0	0	0	0	0	0
	50%	50%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Noissonia oinonoa	2	2	()		0	0			0		0	()
weisseria cinerea	100)%	00	%		0%	0%	6	()%	0)%	0	%
			2											
		10	0%											

	Co Corr Gro	rrect: C ect: Sp up / Co	Genus ID ecies ID mplex II	or O	C Inco Gr	orrect: (rrect: Sj oup / Co	Genus II pecies II omplex I)) or D	In	correc I	: t: Ge D	enus	no	ID
Bacteria	MBT ≥2	C-CA 2.0	$\frac{\text{MBT-}}{\geq 1.7 \text{ to}}$	CA <2.0	MB′ ≥	Т-СА 2.0	$\frac{\text{MBT}}{\geq 1.7 \text{ to}}$	-CA o <2.0	MB ≥	T-CA 2.0	MB′ ≥1. <∕	Г-СА .7 to 2.0	MB] <1	Г-СА 1.7
	1 . 1	1] 1 · 1	RESOI		N REFE	RENCE	ALG	ORIT	THM	h · 1	1	1 • 1	1
	nign 8	10W	nign	10W	nign	low	nign	10W	nign	low	nign	low	nign	low
	100%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
	8	3	()		0	0			0		0	()
Neisseria elongata	100)%	0	%		0%	0%	6	()%	0	1%	0	%
		8	8											
		10	0%			0						0	0	
	56	0	0	0	0	0	0	0	0	0	0	0	0	0
	100%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Neisseria flavescens /	5	6	()		0	0			0		0	()
subflavagroup	100)%	0	%		0%	0%	6	()%	0	%	0	%
		5	6											
		10	00/		_									
		10	0%	-		-	г. <u>-</u>	-		-	-	_	-	_
	45	0	0	0	0	0	0	0	0	0	0	0	0	0
	100%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Naiggoria, conombosas	4	5	()		0	0			0		0	()
Neisseria gonormoede	100)%	09	%		0%	0%	6	()%	0	%	0	%
		4	5						•					
		10	0%											r
	11	0	0	0	0	0	1	0	0	0	0	0	0	0
	91.7%	0%	0%	0%	0%	0%	8.3%	0%	0%	0%	0%	0%	0%	0%
Neisseria lactamica	1	1	()		0	1			0		0	()
iversser a factamed	91.	7%	09	%		0%	8.3	%	()%	0	%	0	%
		1	1											
		91.	7%											
	35	0	0	0	2	0	0	1	0	0	0	0	0	0
	92.1%	0%	0%	0%	5.3%	0%	0%	2.6%	0%	0%	0%	0%	0%	0%
Noissonis moninoitidia	3	5	()		2	1			0		0	()
iveisseria meningiliais	92.	1%	09	%	5	5.3%	2.6	%	()%	0	%	0	%
		3	5											
		92.	1%											

	Co Corr Gro	rrect: C ect: Spe up / Co	Genus ID ecies ID mplex II	or O	C Inco Gi	orrect: (orrect: Sproup / Co	Genus II pecies II omplex I)) or D	In	correc I	: t: Ge D	enus	no	ID
Bacteria	MBT ≥2	C-CA	$\frac{\text{MBT-}}{\geq 1.7 \text{ to}}$	•CA <2.0	MB' ≥	T-CA 2.0	$\frac{\text{MBT}}{\geq 1.7 \text{ to}}$	-CA 0 <2.0	MB ≥	T-CA 2.0	MB′ ≥1. <	Г-СА .7 to 2.0	MB7 <1	Г-СА 1.7
	1 . 1	1	I I I I	RESOL	UTIO	N REFE	RENCE	ALG	ORIT	THM	4 • •	1	1 · 1	1
	nign 30	10W	nign	10W	nign	10W	nign	10W	nign	low	nign	low	nign	low
	100%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
	3	0	()		0	0			0		0	()
<i>Neisseria sicca</i> group	100)%	09	%		0%	09	6	()%	0	%	0	%
		3	0											
		10	0%	r	1		1		T		1	[n	r
	9	0	0	0	0	0	0	0	0	0	0	0	0	0
	100%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Neisseria weaveri	9)	()		0	0			0		0	()
	100)%	09	%		0%	0%	6	()%	0	%	0	%
		()		1									
		10	0%											
	6	8	0	2	0	0	0	0	0	0	0	0	0	0
	37.5%	50%	0%	12.5%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
X 1 1 1 1	1	4	2	2		0	0			0		0	()
Nocardia brasiliensis	87.	5%	12.	5%		0%	09	6	()%	0	%	0	%
		1	6								1		1	
		10	0%											
	20	0	3	0	0	0	0	0	0	0	0	0	0	0
	87.0%	0%	13.0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Nocardia cyriacigeorgica	2	0	3	3		0	0			0		0	()
nocurum cyrneigeorgicu	87.	0%	13.	0%		0%	0%	6	()%	0	%	0	%
		2	3											
		10	0%			T	T		1				1	
	26	0	3	0	0	0	0	0	0	0	0	0	0	0
	89.7%	0%	10.3%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
No oguđi a fanojujo a group	2	6	3	3		0	0			0		0	()
<i>Nocaraia jarcinica</i> group	89.	7%	10.	3%		0%	0%	6	()%	0	%	0	%
		2	9											_
		10	0%		1									

	Co Corr Gro	rrect: C ect: Spe up / Co	Genus ID ecies ID mplex II	or O	C Inco Gi	orrect: (orrect: Sp coup / Co	Genus II pecies II omplex I)) or D	In	correc Il	: t: Ge D	enus	no	ID
Bacteria	MBT ≥2	C-CA 2.0	$\frac{\text{MBT-}}{\geq 1.7 \text{ to}}$	CA <2.0	MB' ≥	T-CA 2.0	$\frac{\text{MBT}}{\geq 1.7 \text{ to}}$	-CA o <2.0	MB ≥	T-CA 2.0	MB′ ≥1. <	Г-СА .7 to 2.0	MB7 <1	Г-СА 7
	1 . 1	1	I I I I	RESO	LUTIO	N REFE	RENCE	ALG		THM	4 · 1	1	1 · 1	1
	high	low	high	low	high	low	high	low	high	low	high	low	high	low
	87.5%	0%	6.3%	0%	0%	0%	0%	0%	0%	0%	0%	0%	2 6.3%	0%
	2	8	2	2		0	0			0		0		2
Nocardia nova	87.	5%	6.3	3%		0%	0%	6	()%	0	%	6.3	3%
		3	0											
		93.	8%					-		-				
	4	0	0	0	0	0	0	0	0	0	0	0	0	0
	100%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Noogadig otitidigoguigmum	2	1	()		0	0			0		0	()
Nocarata ottitaiscaviarum	100	0%	09	%		0%	0%	6	()%	0	%	0	%
		4	1											
		10	0%											
	5	4	0	0	0	0	1	0	0	0	0	0	0	0
	50%	40%	0%	0%	0%	0%	10%	0%	0%	0%	0%	0%	0%	0%
Ochrobactrum anthropi	9)	()		0	1			0		0	()
Ochrobactrum anthropi	90	%	09	%		0%	10	%	()%	0	%	0	%
		())											
		90)%		_									
	2	0	0	0	0	0	0	0	0	0	0	0	0	0
	100%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Parabacteroides	2	2	()		0	0			0		0	()
goldsteinii	100)%	09	%		0%	0%	6	()%	0	%	0	%
		4	2											
		10	0%											
	13	0	0	0	0	0	0	0	0	0	0	0	0	0
Parabacteroides johnsonii	100%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
	1	3	()		0	0			0		0	()
/merdae group	100)%	09	%		0%	0%	6	()%	0	%	0	%
		1	3											
		10	0%											

	Co Corr Gro	rrect: C ect: Spe up / Co	Genus ID ecies ID mplex II	or O	C Inco Gr	orrect: (rrect: S _l oup / Co	Genus II pecies II mplex II)) or D	In	correc Il	: t: Ge D	enus	no	ID
Bacteria	MBT ≥2	C-CA 2.0	MBT- ≥1.7 to	•CA <2.0	MB′ ≥	Г-СА 2.0	$MBT \ge 1.7 \text{ to}$	-CA 0 <2.0	MB ≥	T-CA 2.0	MB′ ≥1. <	Г-СА .7 to 2.0	MB7 <1	Г-СА 1.7
			I	RESOI	LUTIO	N REFE	RENCE	ALG	ORIT	THM				
	high	low	high	low	high	low	high	low	high	low	high	low	high	low
	95.1%	4.9%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
	4	1	()		0	0			0		0	()
Parvimonas micra	10)%	09	%		0%	0%	6	()%	0	%	0	%
		4	1										1	
		10	0%				•							
	10	0	0	0	0	0	0	0	0	0	0	0	0	0
	100%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Pediococcus acidilactici	1	0	()		0	0			0		0	()
	10)%	09	%		0%	0%	6	()%	0	%	0	%
		1	0				•							
		10	0%				•							
	10	0	0	0	0	0	0	0	0	0	0	0	0	0
	100%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Pluralibacter gergoviae	1	0	()		0	0			0		0	()
	10	0%	09	%		0%	0%	6	()%	0	%	0	%
		1	0											
		100	0%	-			-	-	r -					
	1	0	0	0	0	0	0	0	0	0	0	0	0	0
	100%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Porphyromonas somerae	1		()		0	0			0		0	()
	100)%	09	%		0%	0%	0	()%	0	%	0	%
		10			_									
		100)%	1					r					
	9	0	0	0	0	0	0	0	0	0	0	0	0	0
-	100%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Ralstonia pickettii	9)	()		0	0			0		0	()
_	10)%	09	%		0%	0%	6	()%	0	%	0	%
		10)		-									
		10	0 /0											

	Co Corr Gro	rrect: C ect: Spe up / Co	Genus ID ecies ID mplex II	or O	C Inco Gr	orrect: (orrect: Sp coup / Co	Genus II pecies II omplex II)) or D	In	correc I	: t: Ge D	enus	no	ID
Bacteria	MBT ≥2	C-CA 2.0	$\frac{\text{MBT-}}{\geq 1.7 \text{ to}}$	CA <2.0	MB′ ≥	T-CA 2.0	$\frac{\text{MBT}}{\geq 1.7 \text{ to}}$	-CA 0 <2.0	MB ≥	T-CA 2.0	MB' ≥1. <2	Г-СА .7 to 2.0	MB7 <1	Г-СА 1.7
	1.1.1	1.	I L'.L	RESOI		N REFE	RENCE	ALG		THM	1. 1. 1.	1.	1. 1. 1.	1.
	nign 7	10W	nign	10W	nign	low	nign	10W	nign	low	nign	low	nign	low
	100%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
	7	1	()		0	0			0		0	()
Serratia fonticola	100)%	09	%		0%	0%	6	()%	0	%	0	%
		10	7 0%		-									
	8	0	0	0	0	0	0	0	0	0	0	0	0	0
	100%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
	8	3	()		0	0			0		0	()
Serratia odorifera	100)%	09	%		0%	0%	6	()%	0	%	0	%
		8	8											
		10	0%				1						1	1
	5	0	0	0	0	0	0	0	0	0	0	0	0	0
	100%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Sphingobacterium	5	5	()		0	0			0		0	()
multivorum	100)%	09	%		0%	0%	6	()%	0	%	0	%
		-	5											
		10	0%											
	7	0	0	0	0	0	0	0	0	0	0	0	0	0
	100%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Sphingobacterium	7	7	()		0	0			0		0	()
spiritivorum	100)%	09	%		0%	0%	6	()%	0	%	0	%
			7											
			0%											
	16	0	0	0	0	0	0	0	0	0	0	0	0	0
Sphingomonas paucimobilis group	100%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
	1	6	()		0	0			0		0	()
	100)%	09	%		0%	0%	6	()%	0	%	0	%
		1	6		-		_	_		_				
		10	070											

	Co Corr Gro	rrect: (ect: Sp up / Co	Genus ID ecies ID mplex II	or O	C Inco Gr	orrect: (rrect: S _j oup / Co	Genus II pecies II omplex II)) or D	In	correc I	: t: Ge D	enus	no	ID
Bacteria	MBT ≥2	C-CA 2.0	$\frac{\text{MBT-}}{\geq 1.7 \text{ to}}$	CA <2.0	MB' ≥	Т-СА 2.0	$\frac{\text{MBT}}{\geq 1.7 \text{ to}}$	-CA o <2.0	MB ≥	T-CA 2.0	MB′ ≥1 <	Г-СА .7 to 2.0	MB] <]	Г-СА 1.7
	1.1.1	1.	<u> </u>	RESO		N REFE	RENCE	ALG	ORIT	THM	h 1.	1.	1.1.1.	1.
	nign 5	10W	nign	low	nign	low	nign	low	nign	10W	nign	low	nign	low
	100%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
	5	5	()		0	0			0		0	()
Staphylococcus delphini	100)%	09	%		0%	0%	6	()%	0	%	0	%
		:	5											
		10	0%										-	-
	3	0	0	0	0	0	0	0	0	0	0	0	0	0
	100%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Staphylococcus	3	3	()		0	0			0		0	()
intermedius	100)%	09	%		0%	0%	6	()%	0	%	0	%
		-	3											
		10	0%		_									
	3	0	0	0	0	0	0	0	0	0	0	0	0	0
	100%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Staphylococcus lentus	3	3	()		0	0			0		0	()
Staphytococcus tentus	100)%	09	%		0%	0%	6	()%	0	%	0	%
		, -	3											
		10	0%											
	11	0	4	0	0	0	0	0	0	0	0	0	0	0
	73.3%	0%	26.7%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Staphylococcus sciuri	1	1	4	ļ		0	0			0		0	()
	73.	3%	26.	7%		0%	0%	6	()%	0	%	0	%
		1	.5											
		10	0%	1				1	1		1		n	n
	3	0	2	0	0	0	0	0	0	0	0	0	0	0
Staphylococcus xylosus	60%	0%	40%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
	3	3	2	2		0	0			0		0	()
	60	%	40	%		0%	0%	6	()%	0	%	0	%
			5											
		10	0%											

	Co Corr Gro	rrect: C ect: Spe up / Co	Genus ID ecies ID mplex II	or)	C Inco Gr	orrect: (rrect: S _I oup / Co	Genus II pecies II mplex II)) or D	In	correc Il	: t: Ge D	enus	no	ID
Bacteria	MBT ≥2	C-CA 2.0	MBT- ≥1.7 to	CA <2.0	MB' ≥	Т-СА 2.0	MBT ≥1.7 to	-CA 0 <2.0	MB ≥	T-CA 2.0	MB′ ≥1. <	Г-СА .7 to 2.0	MB7 <1	Г-СА 7
			F	RESOL	JUTIO	N REFE	RENCE	ALG	ORIT	ΉM	.	-		
	high	low	high	low	high	low	high	low	high	low	high	low	high	low
	100%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
	1	8	()		0	0			0		0	()
Streptococcus canis	100)%	09	%		0%	0%	6	()%	0	%	0	%
		1	8											
		10	0%										-	
	6	0	0	0	0	0	0	0	0	0	0	0	0	0
	100%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Strantococcus aqui	e	5	0)		0	0			0		0	()
Sirepiococcus equi	100)%	09	%		0%	0%	6	()%	0	%	0	%
		6	6											
		10	0%											
	33	3	0	0	0	9	0	0	0	0	0	0	0	0
	73.3%	6.7%	0%	0%	0%	20%	0%	0%	0%	0%	0%	0%	0%	0%
Streptococcus parasanguinis	3	6	0)		9	0			0		0	()
	80	%	09	%	4	20%	0%	6	()%	0	%	0	%
		3	6											
		80	1%										r	
	32	0	0	0	0	0	0	0	0	0	0	0	0	0
	100%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Streptococcus sanguinis	3	2	0)		0	0			0		0	()
1 0	100	0%	09	%		0%	0%	6	()%	0	%	0	%
		3	2											
		10	0%											
	5	0	0	0	0	0	0	0	0	0	0	0	0	0
Streptococcus sobrinus	100%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
	4	5	C)		0	0			0		0	()
	100)%	09	%		0%	0%	6	()%	0	%	0	%
		4	5											
		10	0%											

	Co Corr Gro	errect: C ect: Spe oup / Co	Genus ID ecies ID mplex II	or O	C Inco Gr	orrect: (rrect: S _j oup / Co	Genus II pecies II mplex I)) or D	In	correc Il	: t: Ge D	enus	no	ID
Bacteria	MBT ≥2	Г-СА 2.0	$\frac{\text{MBT-}}{\geq 1.7 \text{ to}}$	CA <2.0	MB' ≥	Т-СА 2.0	$\frac{\text{MBT}}{\geq 1.7 \text{ to}}$	-CA o <2.0	MB ≥	T-CA 2.0	MB′ ≥1. <	Г-СА .7 to 2.0	MB] <]	Г-СА 1.7
			F	RESOI	LUTIO	N REFE	RENCE	ALG	ORIT	THM	.	-		
	high 50	low	high	low	high	low	high	low	high	low	high	low	high	low
	92.2%	4 6.3%	0%	0%	1.6%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Streptococcus salivarius /	6	3	0)		1	0			0		0	()
vestibularis group	98.	4%	00	%	1	.6%	0%	6	()%	0	%	0	%
		6	3											
		98.	4%						1 -		—		-	-
	4	0	0	0	0	0	0	0	0	0	0	0	0	0
	100%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Streptococcus	4	1	C)		0	0			0		0	()
thermophilus	10	0%	09	%		0%	0%	6	()%	0	%	0	%
		4	4											
		10	0%		_									
	27	0	0	0	0	0	0	0	0	0	0	0	0	0
	100%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Trueperella bernardiae	2	7	0)		0	0			0		0	()
	10	0%	09	%		0%	0%	6	()%	0	%	0	%
		2	.7											
		10	0%											
	24	0	3	0	0	0	0	0	0	0	0	0	0	0
	88.9%	0%	11.1%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Turicella otitidis	2	4	3	3		0	0			0		0	()
Tuncena onnais	88.	9%	11.	1%		0%	09	6	()%	0	%	0	%
		2	.7											
		10	0%				•							
	5	0	0	0	0	0	0	0	0	0	0	0	0	0
	100%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Vagococcus fluvialis	4	5	0)		0	0			0		0	()
vagococcus jiuviaits	10	0%	09	%		0%	0%	6	()%	0	%	0	%
		10	5 0%		_									

	Co Corr Gro	rrect: C ect: Spe up / Cor	Genus ID ecies ID mplex II	or O	C Inco Gr	orrect: (rrect: S _l oup / Co	Genus II pecies II mplex II)) or D	In	correc Il	: t: Ge D	enus	no	ID
Bacteria	MBT ≥2	C-CA 2.0	MBT- ≥1.7 to	CA <2.0	MB' ≥	Г-СА 2.0	$MBT \ge 1.7 \text{ to}$	-CA o <2.0	MB ≥	T-CA 2.0	MB′ ≥1. <	Г-СА .7 to 2.0	MB7 <1	Г-СА 7
			ŀ	RESOI	LUTIO	N REFE	RENCE	ALG	ORIT	THM			1	1
	high	low	high	low	high	low	high	low	high	low	high	low	high	low
	33	0	0	0%	0%	0	0%	0	0	0	0	0	0	0%
*7 *11 11 1	3	3	0)		0	0			0		0	()
Veillonella parvula group	10)%	09	%		0%	0%	6	()%	0	%	0	%
		3	3											
		100)%				1						n	[
	10	0	0	0	0	0	0	0	0	0	0	0	0	0
	100%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Weeksella virosa	1	0	C)		0	0			0		0	()
	10	0%	09	%		0%	0%	6	()%	0	%	0	%
		1	0											
		100)%	1			I	1					1	
	5	0	0	0	0	0	0	0	0	0	0	0	0	0
	100%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Yersinia frederiksenii		5	()	_	0	0			0		0	()
	100	J%	09	%		0%	0%	6	()%	0	%	0	%
		100)%		_									
	8	0	0	0	0	0	0	0	0	0	0	0	0	0
	100%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Varsinia intermedia	8	3	C)		0	0			0		0	()
Tersinia intermedia	10	0%	00	%		0%	0%	6	()%	0	%	0	%
		8	3											
		100)%										-	
	4	3	0	0	0	0	0	0	0	0	0	0	0	0
Yersinia kristensenii	57.1%	42.9%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
		7	C)		0	0			0		0	()
	10	0%	09	%		0%	0%	6	()%	0	%	0	%
		100)%		-									

	Co Corr Gro	rrect: (ect: Sp up / Co	Genus ecies I mplex	ID D or ID	Co Inco or C	rrect: rrect: Group / Il	Genus Specie / Comj D	ID es ID plex	Inco	orrect:	Genu	ıs ID	no	ID
Yeast	MB7 ≥2	Г-СА 2.0	MB7 ≥1. <2	Г-СА 7 to 2.0	MB' ≥	Т-СА 2.0	MBT ≥1.7	7-CA 7 to .0	MB' ≥	T-CA 2.0	MB7 ≥1. <2	Г-СА 7 to 2.0	MBT <1	с-СА .7
			R	ESOL	UTIC	ON RE	FERE	NCE	ALG	ORITH	HM			
	high	low	high	low	high	low	high	low	high	low	high	low	high	low
	9	0	0	0	0	0	0	0	0	0	0	0	1	0
	90%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	10%	0%
Candida intermedia	Ģ	Ð	(0		0	0)		0	(0	1	
Canalaa intermedia	90	%	0	%	0	%	09	%	0)%	0	%	10	%
		9												
		909	%	0	0		0		0				0	
	6	0	0	0	0	0	0	0	0	0	0	0	0	0
	100%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Candida zeylanoides	10))%) %	0	0) Va	0	0	0	0 %)
	10	6	0	/0	0	//0	0,	0	U	//0	0	/0	07	/0
		100	%		_									
	5	0	0	0	0	0	0	0	0	0	0	0	0	0
	100%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
	4	5	()		0	C)		0	(0	0)
Cyberiinanera jaainii	10	0%	0	%	0	%	09	%	0)%	0	%	0%	%
		5												
		100	%			1	I				1	1	I	
	6	0	2	0	0	0	0	0	0	0	0	0	0	0
	75.0%	0%	25.0 %	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Malassezia furfur	6	6 75.0%		2		0	0)		0	(0	0)
	75.	0%	25.	0%	0	%	09	%	C)%	0	%	09	%
		100	0/											
	0	100	¹ %	0	0	0	0	0	0	0	0	0	0	0
	9	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Malassezia	100 /0)	070)	0 /0	0	070	070	070	0	070	070	0 /0)
pachydermatis	10)%	0	5 %	0	0	09	, /o	0	0)%	0	%	09	, %
r		9	Ű	/0		,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,	0,	0		.,.	Ű	/0	0,	•
		100	%											
	20	0	0	0	0	0	0	0	0	0	0	0	0	0
	100%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Rhodotorula	2	0	()		0	0)		0	(0	0)
mucilaginosa	10	0%	0	%	0	%	09	6	0)%	0	%	0%	%
	20			-										
	8	0	0	0	0	0	0	0	0	0	0	0	0	0
	100%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
<i></i>	100,0	3	()	- / 0	0	0)	270	0	(0	0)
Trichosporon inkin	10	0%	0	%	0	%	09	%	0)%	0	%	09	%
		8					•						•	
		100	%											

	Co Corr Gro	rrect: (ect: Sp up / Co	Genus ecies I mplex	ID D or ID	Con Incon or C	r rect: rrect: droup / Il	Genus Specie / Comj D	ID es ID plex	Inco	rrect:	Genu	ıs ID	no	ID
Yeast	MBT ≥2	Г-СА 2.0	MB7 ≥1. <2	Г-СА 7 to 2.0		Г-СА 2.0	MBT ≥1.7 <2	C-CA 7 to .0		Г-СА 2.0	MB7 ≥1.' <2	Г-СА 7 to 2.0	MBT <1	-CA .7
			R	ESOL	UTIO	N RE	FERE	NCE	ALG	ORITH	ΗM			
	high	low	high	low	high	low	high	low	high	low	high	low	high	low
	14	0	0	0	0	0	0	0	0	0	0	0	0	0
	100%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
Trichosporon mucoides	1	4	()		0	0)		0	()	0)
group	10	0%	0	%	0	%	09	%	0	%	0	%	0%	6
		14	1											
		100	%											

Co Corr Gro	ect: Sp oup / Co	Genus ecies I omplex	ID D or ID	Co Incor Gro	orrect: rect: S oup / C	Genus pecies omplex	ID ID or (ID	Inc	orrect:	Genus	ID	no	ID
REFER	RENCE /	ALGORI	THM				REFE	RENCE	ALGOR	THM			
high	low	high	low	high	low	high	low	high	low	high	low	high	low
res.	res.	res.	res.	res.	res.	res.	res.	res.	res.	res.	res.	res.	res.
				Α	В	С	D	E1	E2	F1	F2	G1	G2
1904	130	23	5	8 9 2 1 0 4 0 1							3	1	
91.1%	6.2%	1.1%	0.2%	0.4%	0.4%	0.1%	0.0%	0.0%	0.2%	0.0%	0.0%	0.1%	0.0%
203	34	2	8	17 3 4 1				2	1				
97.3	3%	1.	3%	0.8% 0.1% 0.2% 0.0%						0.2	2%		

Table: Overall Incidence of Incorrect Identifications

|--|

Reference Method

Code

	Correct genus ID - Incorrect spec	ies ID	
	Bacteroides stercoris	Bacteroides eggerthii	
	Klebsiella pneumoniae	Klebsiella variicola	
	Lactobacillus gasseri	Lactobacillus johnsonii	
	Neisseria meningitidis	Neisseria polysaccharea	Α
	Neisseria meningitidis	Neisseria polysaccharea	(8x)
	Sphingomonas paucimobilis	Sphingomonas zeae	
	Sphingomonas paucimobilis	Sphingomonas zeae	
	Streptococcus salivarius ssp salivarius	Streptococcus thermophilus	
1)	Streptococcus parasanguinis	Streptococcus sp. (Viridans)	
	Streptococcus parasanguinis	Streptococcus sp. (Viridans)	
	Streptococcus parasanguinis	Streptococcus sp. (Viridans)	
	Streptococcus parasanguinis	Streptococcus sp. (Viridans)	в
	Streptococcus parasanguinis	Streptococcus sp. (Viridans)	(Qv)
	Streptococcus parasanguinis	Streptococcus sp. (Viridans)	(37)
	Streptococcus parasanguinis	Streptococcus sp. (Viridans)	
	Streptococcus parasanguinis	Streptococcus sp. (Viridans)	
	Streptococcus parasanguinis	Streptococcus sp. (Viridans)	
	Neisseria lactamica	Neisseria cinerea	С
3)	Ochrobactrum anthropi	Ochrobactrum intermedium	(2x)
	Neisseria meningitidis	Neisseria sp.	D

	Incorrect genus ID		
	Ewingella americana	Rahnella sp.	
2	Ewingella americana	Rahnella sp.	E2
2)	Ewingella americana	Rahnella sp.	(4x)
	Escherichia hermannii	Enterobacter sp.	
4)	Escherichia hermannii	Enterobacter sp.	F2

	no ID		
	no ID	Nocardia nova	61
- \	no ID	Nocardia nova	(2)
5)	no ID	Candida intermedia	(5X)
	no ID	Clostridium sporogenes	G2

If an indication for the possibility of cross-matching patterns was found, the organism was included in the matching hint table found in the package labeling and as found below.

Matching Hint	Species / Group / Complex	Strains Included in Database	Different species are potentially associated with the displayed identification. Confirmatory tests are required to differentiate between listed organisms.	Matching Hint
a	Achromobacter xylosoxidans	A. xylosoxidans	A.xylosoxidans; A.denitrificans; A.insolitus; A.marplatensis; A.ruhlandii; A.spanius	Based on 16S rRNA gene sequencing a secure species differentiation between the displayed species is difficult. All species associated with the displayed identification have been reported as isolated from human specimens.
by	Acinetobacter calcoaceticus	A. calcoaceticus		The displayed species should be considered as member of the Acinetobacter baumannii complex. For organisms identified by the MBT-CA System as Acinetobacter calcoaceticus; Acinetobacter pittii or Acinetobacter baumannii/nosocomialis group the full Extraction procedure (Ext) is mandatory for secure species differentiation.
bz	Acinetobacter pittii	A. pittii		The displayed species should be considered as member of the Acinetobacter baumannii complex. For organisms identified by the MBT-CA System as Acinetobacter calcoaceticus; Acinetobacter pittii or Acinetobacter baumannii/nosocomialis group the full Extraction procedure (Ext) is mandatory for secure species differentiation.

Table: Matching Hint Table

Matching Hint	Species / Group / Complex	Strains Included in Database	Different species are potentially associated with the displayed identification. Confirmatory tests are required to differentiate between listed organisms.	Matching Hint
ca	Acinetobacter baumannii nosocomialis group	A. baumannii; A. nosocomialis	A. baumannii; A. nosocomialis	The displayed species should be considered as member of the Acinetobacter baumannii complex. For organisms identified by the MBT-CA System as Acinetobacter calcoaceticus; Acinetobacter pittii or Acinetobacter baumannii/ nosocomialis group the full Extraction procedure (Ext) is mandatory for secure species differentiation.
z	Actinomyces oris	A. oris	A. oris; A. naeslundii; A.viscosus	Based on 16S rRNA gene sequencing a secure species differentiation between the displayed species is difficult.
cb	Actinotignum schaalii group	A. schaalii; A. sanguinis	A. schaalii; A. sanguinis	Differentiation between members of the displayed group by the MBT-CA is not possible although the reference method (16S rRNA gene sequencing) is able to distinguish the mentioned species. Therefore they are grouped together.
aa	Aerococcus viridans	A. viridans	A. viridans; A. urinaeequi	Based on 16S rRNA gene sequencing a secure species differentiation between the displayed species is difficult.
ab	Aeromonas salmonicida	A. salmonicida	A. salmonicida; A. bestiarum; A. molluscorum; A. rivuli; A. encheleia	Based on 16S rRNA gene sequencing a secure species differentiation between the displayed species is difficult.

Matching Hint	Species / Group / Complex	Strains Included in Database	Different species are potentially associated with the displayed identification. Confirmatory tests are required to differentiate between listed organisms.	Matching Hint
с	Aeromonas sp[7]	A. allosaccharophilaA. caviaeA. culicicolaA. hydrophilaA. ichthiosmiaA. sobriaA. veronii	A.allosaccharophila; A.aquariorum; A.caviae; A.culicicola; A.enteropelogenes; A.fluvialis; A.hydrophila; A.hydrophila; A.ichthiosmia; A.jandaei; A.media; A.media; A.rivuli; A.sanarellii; A.sobria; A.taiwanensis; A.veronii	Differentiation between members of the displayed groups or complexes based on 16S rRNA gene sequencing is difficult; whereas they are grouped together. <i>A.hydrophila;</i> <i>A. caviae</i> and <i>A.sobria</i> (<i>A.veronii bv sobria</i>) are the most frequently reported species associated with human infection. <i>A.ichthiosmia</i> is considered a synonym of <i>A.veronii</i> and <i>A.punctata</i> is considered a synonym of <i>A.caviae</i> .
сс	Arthrobacter cumminsii	A. cumminsii	A. cumminsii; A. albus	Based on 16S rRNA gene sequencing a secure species differentiation between the displayed species is difficult.
	Bacteroides stercoris group	B. stercoris	B. stercoris; B. eggerthii	Differentiation between members of the displayed group by the MBT-CA is not possible although the reference method (16S rRNA gene sequencing) is able to distinguish the mentioned species. Therefore they are grouped together.
ac	Bacteroides ovatus group	B. ovatus; B. xylanisolvens	B. ovatus; B. xylanisolvens	Differentiation between members of the displayed group by the MBT-CA is not possible although the reference method (16S rRNA gene sequencing) is able to distinguish the mentioned species. Therefore they are grouped together.

Matching Hint	Species / Group / Complex	Strains Included in Database	Different species are potentially associated with the displayed identification. Confirmatory tests are required to differentiate between listed organisms.	Matching Hint
ad	Bacteroides thetaiotaomicron group	B. thetaiotaomicron; B. faecis	B. thetaiotaomicron; B. faecis	Differentiation between members of the displayed group by the MBT-CA is not possible although the reference method (16S rRNA gene sequencing) is able to distinguish the mentioned species. Therefore they are grouped together.
ae	Bacteroides vulgatus group	B. vulgatus; B. dorei	B. vulgatus; B. dorei	Differentiation between members of the displayed group by the MBT-CA is not possible although the reference method (16S rRNA gene sequencing) is able to distinguish the mentioned species. Therefore they are grouped together.
af	Bordetella group[3]	B. bronchiseptica; B. parapertussis; B. pertussis	B. bronchiseptica; B. parapertussis; B. pertussis	Differentiation between members of the displayed groups or complexes based on 16S rRNA gene sequencing is difficult; whereas they are grouped together.
ag	Brevundimonas diminuta group	B. diminuta; B. naejangsanensis	B. diminuta; B. naejangsanensis	Differentiation between members of the displayed group by the MBT-CA is not possible although the reference method (16S rRNA gene sequencing) is able to distinguish the mentioned species. Therefore they are grouped together.

Matching Hint	Species / Group / Complex	Strains Included in Database	Different species are potentially associated with the displayed identification. Confirmatory tests are required to differentiate between listed organisms.	Matching Hint
d	Burkholderia gladioli	B. gladioli	B.gladioli; B.glumae; B.caryopylii	Based on 16S rRNA gene sequencing a secure species differentiation between the displayed species is difficult. <i>B. glumae</i> and <i>B. caryopylii</i> have not been reported as isolated with human specimens.
cd	Burkholderia multivorans	B. multivorans		The displayed species should be considered as member of the Burkholderia cepacia complex.
e	Burkholderia cepacia complex [13]	B. ambifaria; B. anthina; B. cenocepacia; B. cepacia; B. diffusa; B. dolosa; B. lata; B. latens; B. latens; B. metallica; B. pyrrocinia; B. seminalis; B. stabilis; B. vietnamiensis	B.ambifaria; B.anthina; B.cenocepacia; B.cepacia; B.diffusa; B.dolosa; B.lata; B.lata; B.latens; B.metallica; B.pyrrocinia; B.seminalis; B.stabilis; B.vietnamiensis	Differentiation between members of the displayed groups or complexes based on 16S rRNA gene sequencing is difficult; whereas they are grouped together. All species associated with the displayed identification have been reported as isolated from human specimens.
ah	Chryseobacterium gleum	C. gleum	C. gleum; C. bernardetii	Based on 16S rRNA gene sequencing a secure species differentiation between the displayed species is difficult.
f	Citrobacter amalonaticus complex	C. amalonaticus; C. farmeri	C. amalonaticus; C. farmeri	Differentiation between members of the displayed groups or complexes based on 16S rRNA gene sequencing is difficult; whereas they are grouped together. All species associated with the displayed identification have been reported as isolated from human specimens.

Matching Hint	Species / Group / Complex	Strains Included in Database	Different species are potentially associated with the displayed identification. Confirmatory tests are required to differentiate between listed organisms.	Matching Hint
g	Citrobacter freundii complex	C. braakii; C. freundii; C. gillenii; C. murliniae; C. rodentium; C. sedlakii; C. werkmannii; C. youngae	C.braakii; C.freundii; C.gillenii; C.murliniae; C.rodentium; C.sedlakii; C.werkmannii; C.youngae	Differentiation between members of the displayed groups or complexes based on 16S rRNA gene sequencing is difficult; whereas they are grouped together. <i>C. rodentium</i> has not been reported as isolated from human specimens.
се	Clostridium beijerinckii	C. beijerinckii	C. beijerinckii; C. diolis; C. roseum; C. saccharoperbutylacet onicum	Based on 16S rRNA gene sequencing a secure species differentiation between the displayed species is difficult.
cf	Clostridium clostridioforme group	C. clostridioforme; C. bolteae	C. clostridioforme; C. bolteae	Differentiation between members of the displayed group by the MBT-CA is not possible although the reference method (16S rRNA gene sequencing) is able to distinguish the mentioned species. Therefore they are grouped together.
cg	Clostridium sporogenes	C. sporogenes	C. sporogenes; C. botulinum (group I)	Based on 16S rRNA gene sequencing a secure species differentiation between the displayed species is difficult. <i>C. botulinum</i> is not included in the MBT-CA database. <i>C.</i> <i>botulinum</i> is a select agent and should be ruled out; handle isolate with extreme caution and handle in accordance with local; state; and federal accrediting organizations' requirements as applicable.

Matching Hint	Species / Group / Complex	Strains Included in Database	Different species are potentially associated with the displayed identification. Confirmatory tests are required to differentiate between listed organisms.	Matching Hint
ch	Corynebacterium afermentans group	C. afermentans; C. pilbarense	C. afermentans; C. pilbarense	Differentiation between members of the displayed group by the MBT-CA is not possible although the reference method (16S rRNA gene sequencing) is able to distinguish the mentioned species. Therefore they are grouped together.
ak	Corynebacterium aurimucosum group	C. aurimucosum	C. aurimucosum; C. singulare	Differentiation between members of the displayed group by the MBT-CA is not possible although the reference method (16S rRNA gene sequencing) is able to distinguish the mentioned species. Therefore they are grouped together.
ci	Corynebacterium mucifaciens ureicelerivorans group	C. mucifaciens; C. ureicelerivorans	C. mucifaciens; C. ureicelerivorans	Differentiation between members of the displayed group by the MBT-CA is not possible although the reference method (16S rRNA gene sequencing) is able to distinguish the mentioned species. Therefore they are grouped together.
al	Corynebacterium striatum group	C. striatum; C. simulans	C. striatum; C. simulans	Differentiation between members of the displayed group by the MBT-CA is not possible although the reference method (16S rRNA gene sequencing) is able to distinguish the mentioned species. Therefore they are grouped together.
am	Cronobacter sakazakii group	C. sakazakii; C. dublinensis; C. muytjensii; C. turicensis	C. sakazakii; C. dublinensis; C. muytjensii; C. turicensis	Differentiation between members of the displayed group by the MBT-CA is not possible although the reference method (16S rRNA gene sequencing) is able to distinguish the mentioned species. Therefore they are grouped together.
an	Cupriavidus pauculus group	C. pauculus; C. metallidurans	C. pauculus; C. metallidurans	Differentiation between members of the displayed group by the MBT-CA is not

Matching Hint	Species / Group / Complex	Strains Included in Database	Different species are potentially associated with the displayed identification. Confirmatory tests are required to differentiate between listed organisms.	Matching Hint
				possible although the reference method (16S rRNA gene sequencing) is able to distinguish the mentioned species. Therefore they are grouped together.
ao	Delftia acidovorans group	D. acidovorans; D. lacustris; D. litopenaei; D. tsuruhatensis	D. acidovorans; D. lacustris; D. litopenaei; D. tsuruhatensis	Differentiation between members of the displayed group by the MBT-CA is not possible although the reference method (16S rRNA gene sequencing) is able to distinguish the mentioned species. Therefore they are grouped together.
ap	Edwardsiella tarda	E. tarda	E. tarda; E. hoshinae; E. ictaluri	Based on 16S rRNA gene sequencing a secure species differentiation between the displayed species is difficult.
aq	Elizabethkingia meningoseptica group	E. meningoseptica; E. anophelis; E. miricola	E. meningoseptica; E. anophelis; E. miricola	Differentiation between members of the displayed group by the MBT-CA is not possible although the reference method (16S rRNA gene sequencing) is able to distinguish the mentioned species. Therefore they are grouped together.
h	Enterobacter cloacae complex	E. asburiae; E. cancerogenus; E. cloacae; E. hormaechei; E. kobei; E. ludwigii	E. asburiae; E. cancerogenus; E. cloacae; E. cowanii; E. hormaechei; E. kobei; E. ludwigii; E. nudwigii; E. mori; E. nimipressuralis; E. soli	Differentiation between members of the displayed groups or complexes based on 16S rRNA gene sequencing is difficult; whereas they are grouped together. <i>E. mori</i> and <i>E. soli</i> have not been reported as isolated from human specimens.
i	Escherichia coli	E. coli	E. albertii; E. coli; E. fergusonii; Shigella spp.	Based on 16S rRNA gene sequencing a secure species differentiation between the displayed species is difficult. All species associated with the displayed identification have been reported as isolated from human specimens.
j	Haemophilus influenzae	H. influenzae	H. influenza;	Based on 16S rRNA gene

Matching Hint	Species / Group / Complex	Strains Included in Database	Different species are potentially associated with the displayed identification. Confirmatory tests are required to differentiate between listed organisms.	Matching Hint
			H. aegyptius	sequencing a secure species differentiation between the displayed species is difficult.
as	Haemophilus parahaemolyticus group	H. parahaemolyticus; H. paraphrohaemolyticu s	H. parahaemolyticus; H. paraphrohaemolyticus	Differentiation between members of the displayed group by the MBT-CA is not possible although the reference method (16S rRNA gene sequencing) is able to distinguish the mentioned species. Therefore they are grouped together.
k	Hafnia alvei	H. alvei	H. alvei; H. paralvei; Obesumbacterium proteus	Based on 16S rRNA gene sequencing a secure species differentiation between the displayed species is difficult. <i>Obesumbacterium proteus</i> is most commonly associated with brewery spoilage and has not reported as isolated from human specimens.
1	Klebsiella pneumoniae	K. pneumoniae	K. pneumoniae; K. granulomatis; K. singaporensis	Based on 16S rRNA gene sequencing a secure species differentiation between the displayed species is difficult. All species associated with the displayed identification have been isolated from human specimens; <i>K.</i> <i>pneumoniae</i> is the most common species reported as isolated from human specimens.
cj	Klebsiella variicola	K. variicola	K. variicola; K. granulomatis; K. singaporensis	Based on 16S rRNA gene sequencing a secure species differentiation between the displayed species is difficult.

Matching Hint	Species / Group / Complex	Strains Included in Database	Different species are potentially associated with the displayed identification. Confirmatory tests are required to differentiate between listed organisms.	Matching Hint
m	Klebsiella oxytocaRaoultella ornithinolytica	K. oxytocaR. ornithinolytica	K. oxytoca; R. ornithinolytica; R. planticola	Differentiation between members of the displayed groups or complexes based on 16S rRNA gene sequencing is difficult; whereas they are grouped together. All species associated with the displayed identification have been reported as isolated from human specimens.
ck	Lactobacillus gasseri	L. gasseri	L. gasseri; L. taiwanensis	Based on 16S rRNA gene sequencing a secure species differentiation between the displayed species is difficult.
cl	Lactobacillus jensenii	L. jensenii	L. jensenii; L. fornicalis; L. psittaci	Based on 16S rRNA gene sequencing a secure species differentiation between the displayed species is difficult.
cm	Leuconostoc citreum	L. citreum	L. citreum; L. holzapfelii	Based on 16S rRNA gene sequencing a secure species differentiation between the displayed species is difficult.
сп	Listeria monocytogenes	L. monocytogenes	L. monocytogenes; L. inocua; L. ivanovii; L. marthii; L. seeligeri; L. welshimeri	Secure species differentiation for <i>Listeria</i> species is difficult since several species within the genus Listeria are very closely related. Also alternative methods (e.g. DNA sequencing) show low discrimatory power. For <i>Listeria</i> identified by theMBT-CA it is recommended to proceed with full extraction procedure (Ext) for final identification. Differentiation between members of the displayed group by the MBT-CA is not possible although the
	Mannheimia haemolytica group	M. haemolytica	M. haemolytica; M. glucosida	possible although the reference method (16S rRNA gene sequencing) is able to distinguish the mentioned species. Therefore they are grouped together.
n	Moraxella sg Moraxella osloensis	M. osloensis	Moraxella osloensis; Enhydrobacter	Based on 16S rRNA gene sequencing a secure species

			Different species are	
Matching Hint	Species / Group / Complex	Strains Included in Database	potentially associated with the displayed identification. Confirmatory tests are required to differentiate between listed organisms.	Matching Hint
			aerosaccus	differentiation between the
				displayed species is difficult. The rare species <i>Enhydrobacter aerosaccus</i> is closely related to <i>Moraxella</i> <i>osloensis;</i> and has not reported as isolated from human specimens.
0	Morganella morganii	M. morganii	M. morganii; M. psychrotolerans	Based on 16S rRNA gene sequencing a secure species differentiation between the displayed species is difficult. Both species associated with the displayed identification have been reported as isolated from human specimens.
au	Myroides odoratimimus	M. odoratimimus	M. odoratimimus; M. profundi	Based on 16S rRNA gene sequencing a secure species differentiation between the displayed species is difficult.
ср	Neisseria flavescens subflava group	N. flavescens; N. perflava; N. subflava	N. flavescens; N. perflava; N. subflava	Differentiation between members of the displayed group by the MBT-CA is not possible although the reference method (16S rRNA gene sequencing) is able to distinguish the mentioned species. Therefore they are grouped together.
cq	Neisseria gonorrhoeae	N. gonorrhoeae		If the reference method reported N. gonorrhoeae NO false identification of the MBT-CA was observed during clinical tests.
cr	Neisseria meningitidis	N. meningitidis		In rare cases apathogenic <i>Neisseria</i> species could be identified as N.meningitidis with the MBT-CA. If the reference method reported <i>N.meningitidis</i> NO false identification of the MBT- CA was observed during clinical tests.
cs	Neisseria sicca group	N. macacae; N. mucosa; N. sicca	N. macacae; N. mucosa; N. sicca; Morococcus	Differentiation between members of the displayed group is not possible by the MBT-CA System or

Matching Hint	Species / Group / Complex	Strains Included in Database	Different species are potentially associated with the displayed identification. Confirmatory tests are required to differentiate between listed organisms.	Matching Hint
			cerebrosus	reference method (16S rRNA gene sequencing). Therefore they are grouped together.
ct	Nocardia brasiliensis	N. brasiliensis	N. brasiliensis; N. iowensis; N. vulneris	Based on 16S rRNA gene sequencing a secure species differentiation between the displayed species is difficult.
cu	Nocardia farcinica group	N. farcinica; N. kroppenstedtii	N. farcinica; N. kroppenstedtii	Differentiation between members of the displayed group is not possible by the MBT-CA System or reference method (16S rRNA gene sequencing). Therefore they are grouped together.
cv	Ochrobactrum anthropi	O. anthropi	O. anthropi; O. lupini	Based on protein gene sequencing a secure species differentiation between the displayed species is difficult.
p	Pantoea agglomerans	P. agglomerans	P. agglomerans; P. anthophila; P. brenneri; P. conspicua; P. eucalypti; P. vagans	Based on 16S rRNA gene sequencing a secure species differentiation between the displayed species is difficult. All species associated with the displayed identification have been reported as isolated from human specimens. <i>Pantoea</i> <i>agglomerans</i> ; is the most commonly reported <i>Pantoea</i> species isolated from human specimens.
cw	Parabacteroides johnsonii merdae group	P. johnsonii; P. merdae	P. johnsonii; P. merdae	Differentiation between members of the displayed group by the MBT-CA is not possible although the reference method (16S rRNA gene sequencing) is able to distinguish the mentioned species. Therefore they are grouped together.

Matching Hint	Species / Group / Complex	Strains Included in Database	Different species are potentially associated with the displayed identification. Confirmatory tests are required to differentiate between listed organisms.	Matching Hint
av	Peptoniphilus harei group	P. harei	P. harei; P. indolicus	Differentiation between members of the displayed group by the MBT-CA is not possible although the reference method (16S rRNA gene sequencing) is able to distinguish the mentioned species. Therefore they are grouped together.
q	Proteus vulgaris group	P. hauseri; P. penneri; P. vulgaris	P. hauseri; P. penneri; P. vulgaris	Differentiation between members of the displayed groups or complexes based on 16S rRNA gene sequencing is difficult; whereas they are grouped together. All species associated with the displayed identification have been reported as isolated from human specimens.
r	Providencia rettgeri	P. rettgeri	P. rettgeri; P. alcalifaciens; P. burhodogranariea; P. heimbachae; P. rustigianii; P. vermicola	Based on 16S rRNA gene sequencing a secure species differentiation between the displayed species is difficult. <i>P. burhodogranariea</i> and <i>P. vermicola</i> have not been reported as isolated from human specimens

Matching Hint	Species / Group / Complex	Strains Included in Database	Different species are potentially associated with the displayed identification. Confirmatory tests are required to differentiate between listed organisms.	Matching Hint
S	Pseudomonas fluorescens group	P. azotoformans; P. brenneri; P. cedrina; P. congelans; P. corrugata; P. extremorientalis; P. fluorescens; P. gessardii; P. libanensis; P. mandelii; P. marginalis; P. marginalis; P. migulae; P. mucidolens; P. orientalis; P. poae; P. rhodesiae; P. synxantha; P. tolaasii; P. trivialis; P. veronii	P. azotoformans; P. brenneri; P. cedrina; P. congelans; P. corrugata; P. extremorientalis; P. fluorescens; P. gessardii; P. libanensis; P. mandelii; P. marginalis; P. migulae; P. mucidolens; P. orientalis; P. poae; P. rhodesiae; P. synxantha; P. tolaasii; P. trivialis; P. veronii	Differentiation between members of the displayed groups or complexes based on 16S rRNA gene sequencing is difficult; whereas they are grouped together. Members of the <i>P</i> . <i>fluorescens</i> group are environmental organisms. <i>P</i> . <i>fluorescens</i> is the most commonly isolated species reported as isolated from human specimens.
aw	Pseudomonas oryzihabitans	P. oryzihabitans	P. oryzihabitans; P. oleovorans; P. psychrotolerans	Based on 16S rRNA gene sequencing a secure species differentiation between the displayed species is difficult.
t	Pseudomonas putida group	P. fulva; P. monteilii; P. mosselii; P. plecoglossicida; P. putida	P. fulva; P. monteilii; P. mosselii; P. plecoglossicida; P. putida	Differentiation between members of the displayed groups or complexes based on 16S rRNA gene sequencing is difficult; whereas they are grouped together. Members of the <i>P.</i> <i>putida</i> group are environmental organisms. <i>P.</i> <i>putida</i> is the most commonly isolated species reported as isolated from human specimens.
ax	Rhizobium radiobacter	R. radiobacter	R. radiobacter; R. massilense; R. leguminosarum	Based on 16S rRNA gene sequencing a secure species differentiation between the displayed species is difficult.
u	Salmonella sp	Salmonella sp		Identification is possible on genus level only.
	Serratia fonticola	S. fonticola		
Matching Hint	Species / Group / Complex	Strains Included in Database	Different species are potentially associated with the displayed identification. Confirmatory tests are required to differentiate between listed organisms.	Matching Hint
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v	Serratia liquefaciens	S. liquefaciens	S. liquefaciens; S. proteamaculans; S. grimesii; S. plymuthica; S. ficaria	Based on 16S rRNA gene sequencing a secure species differentiation between the displayed species is difficult. All species associated with the displayed identification have been reported as isolated from human specimens.
w	Serratia marcescens	S. marcescens	S. marcescens; S. nematodiphila; S. ureilytica	Based on 16S rRNA gene sequencing a secure species differentiation between the displayed species is difficult. <i>S. nematodiphila</i> and <i>S.</i> <i>ureilytica</i> have not been reported as isolated from human specimens.
ay	Serratia plymuthica	S. plymuthica	S. plymuthica; S. grimesii; S. proteamaculans; S. quinivorans	Based on 16S rRNA gene sequencing a secure species differentiation between the displayed species is difficult.
	Sphingomonas paucimobilis group	S. paucimobilis	S. paucimobilis; S. zeae	Differentiation between members of the displayed group by the MBT-CA is not possible although the reference method (16S rRNA gene sequencing) is able to distinguish the mentioned species. Therefore they are grouped together.
az	Staphylococcus carnosus	S. carnosus	S. carnosus; S. condimenti; S. piscifermentans	Based on 16S rRNA gene sequencing a secure species differentiation between the displayed species is difficult.
bc	Staphylococcus vitulinus	S. vitulinus	S. vitulinus; S. fleurettii	Based on 16S rRNA gene sequencing a secure species differentiation between the displayed species is difficult.
x	Stenotrophomonas maltophilia	S. maltophilia; Pseudomonas beteli; Pseudomonas hibiscola; Pseudomonas geniculata	S. maltophilia; Pseudomonas beteli; Pseudomonas hibiscola; Pseudomonas geniculata	S. maltophilia; P. beteli; P. hibiscola; P. geniculata are synonymously used taxonomical names.

Matching Hint	Species / Group / Complex	Strains Included in Database	Different species are potentially associated with the displayed identification. Confirmatory tests are required to differentiate between listed organisms.	Matching Hint
bd	Streptococcus lutetiensis	S. lutetiensis	S. lutetiensis; S. infantarius ssp. coli; S. infantarius ssp infantarius	The species designation Streptococcus lutetiensis and Streptococcus pasteurianus have been proposed for S. bovis biotype II.1 and biotype II.2 strains; respectively (Poyart et al.; 2002). However; S. lutetiensis exhibits both phenotypic and genetic similarity to S. infantarius subsp. coli (Schlegel et al.; 2000); and preliminary results show a close relationship between S. pasteurianus and S. gallolyticus.
сх	Streptococcus parasanguinis	S. parasanguinis	S. parasanguinis; S. australis; S. infantis; S. lactarius; S. rubneri	Based on 16S rRNA gene sequencing a secure species differentiation between the displayed species is difficult. Displayed species are members of the " <i>Streptococcus viridans</i> group".
be	Streptococcus pneumoniae	S. pneumoniae		For organisms identified by the MBT-CA as <i>Streptococcus pneumoniae</i> or <i>Streptococcus mitis/oralis</i> group; it is recommended to proceed with full extraction (EXT) for final identification.
су	Streptococcus salivarius vestibularis group	S. salivarius; S. vestibularis	S. salivarius; S. vestibularis	Differentiation between members of the displayed group by MBT-CA is not possible although the reference method (protein gene sequencing) is able to distinguish the mentioned species. Therefore they are grouped together. <i>S. salivarius</i> is also known as <i>S. salivarius ssp salivarius</i> .
CZ	Streptococcus thermophilus	S. thermophilus		known as S. salivarius ssp thermophilus.

Matching Hint	Species / Group / Complex	Strains Included in Database	Different species are potentially associated with the displayed identification. Confirmatory tests are required to differentiate between listed organisms.	Matching Hint
bg	Streptococcus mitis oralis group	S. mitis; S. oralis	S. mitis; S. oralis; S. australis; S. cristatus; S. dentisani; S. infantis; S. oligofermentans; S. pseudopneumoniae; S. rubneri; S. sanguinis; S. tigurinus	For organisms identified by the MBT-CA as Streptococcus pneumoniae or Streptococcus mitis/oralis group; it is recommended to proceed with full extraction (EXT) for final identification. During clinical trials several species were identified with the reference algorithm (low confidence) for isolates identified as "S. mitis oralis group" with the MBT-CA. The following species are mentioned: Streptococcus mitis; S. oralis; S. australis; S. cristatus; S. dentisani; S. infantis; S. nifantis; S. nigofermentans; S. pseudopneumoniae; S. rubneri; S. sanguinis and S. tigurinus. They may be identified by the Streptococcus mitis oralis reference spectrum in the MBT-CA.
da	Veillonella parvula group	V. parvula	V. parvula; V. denticariosi; V. dispar; V. rugosae	Differentiation between members of the displayed group by MBT-CA is not possible although the reference method (protein gene sequencing) is able to distinguish the mentioned species. Therefore they are grouped together.
bh	Vibrio parahaemolyticus	V. parahaemolyticus	V. parahaemolyticus; V. alginolyticus; V. harveyi; V. campbellii; V. natriegens; V. rotiferianus	Based on 16S rRNA gene sequencing a secure species differentiation between the displayed species is difficult.

Matching Hint	Species / Group / Complex	Strains Included in Database	Different species are potentially associated with the displayed identification. Confirmatory tests are required to differentiate between listed organisms.	Matching Hint
у	Yersinia pseudotuberculosis	Y. pseudotuberculosis	Y. pseudotuberculosis; Y. pestis; Y. similis	Based on 16S rRNA gene sequencing a secure species differentiation between the displayed species is difficult. <i>Y. pestis</i> is not included in the MBT-CA database. <i>Y. pestis</i> is a select agent and should be ruled out; handle isolate with extreme caution and handle in accordance with local; state; and federal accrediting organizations' requirements as applicable. All species associated with the displayed identification have been reported as isolated from human specimens.

Matching Hint	Species	strains used for database creation	Different species are potentially associated with the displayed identification. Based on ITS sequencing a secure species differentiation between the displayed species is difficult. Confirmatory tests are required to differentiate between listed organisms.	Matching Hint; Synonyms (anamorph or teleomorph)
bi	Candida famata	C. famata		Debaryomyces hansenii
bj	Candida guilliermondii	C. guilliermondii		C. guilliermondii; Meyerozyma caribbica; Meyerozyma guilliermondii; Pichia guilliermondii; Pichia caribbica
	Candida inconspicua	C. inconspicua	C. inconspicua; C. norvegensis; P. cactophilia	
bk	Candida kefyr	C. kefyr		C. kefyr; Kluyveromyces marxianus
bl	Candida krusei	C. krusei		C. krusei; Issatchenkia orientalis; Pichia kudriavzevii
bm	Candida lambica	C. lambica		C. lambica; Pichia fermentans
bn	Candida lipolytica	C. lipolytica		C. lipolytica; Yarrowia lipolytica
bo	Candida lusitaniae	C. lusitaniae		C. lusitaniae; Clavispora lusitaniae
bp	Candida norvegensis	C. norvegensis		C. norvegensis; Pichia norvegensis
bq	Candida pelliculosa	C. pelliculosa		C. pelliculosa; Pichia anomala; Wickerhamomyces anomalus
br	Candida valida	C. valida		C. valida; Pichia membranifaciens
bs	Cryptococcus gattii	C. gattii		Cryptococcus gattii; Cryptococus bacillisporus; Filobasidiella bacillispora
db	Cyberlindnera jadinii	C. jadinii		Candida utilis
bt	Geotrichum candidum	G. candidum		Dipodascus geotrichum; Galactomyces geotrichum; G. candidum
bu	Geotrichum capitatum	G. capitatum		Dipodascus capitatus; G. capitatum; Magnusiomyces capitatus

Table: Matching Hint Table / Synonym Table Yeasts

Matching Hint	Species	strains used for database creation	Different species are potentially associated with the displayed identification. Based on ITS sequencing a secure species differentiation between the displayed species is difficult. Confirmatory tests are required to differentiate between listed organisms.	Matching Hint; Synonyms (anamorph or teleomorph)
bv	Kloeckera apiculata	K. apiculata		Hanseniaspora uvarum; K. apiculata
bw	Pichia ohmeri	P. ohmeri		Candida guilliermondii var membranaefaciens; Kodamaea ohmeri; Pichia ohmeri
bx	Saccharomyces cerevisiae	S. cerevisiae		Candida robusta; S. cerevisiae
dc	Trichosporon mucoides group	T. dermatis, T. mucoides	T. mucoides; T. dermatis	Differentiation between members of the displayed group by MBT- CA is not possible. The reference method (ITS sequencing) is also not able to distinguish the mentioned species. Therefore they are grouped together.

- k. Clinical specificity: See clinical sensitivity results
- *l.* Other clinical supportive data (when a. and b. are not applicable):
- 2. <u>Clinical cut-off:</u>

See Assay cut-off

3. Expected values/Reference range:

See Assay cut-off

N. Instrument Name:

MALDI Biotyper CA (MBT-CA) System, MBT smart CA System

- O. System Descriptions: Also see K130831
 - 1. Modes of Operation:

Does the applicant's device contain the ability to transmit data to a computer, webserver, or mobile device?

Yes X or No

Does the applicant's device transmit data to a computer, webserver, or mobile device using wireless transmission?

Yes or No X

2. Software:

FDA has reviewed applicant's Hazard Analysis and software development processes for this line of product types:

Yes X or No

3. Specimen Identification:

The user manually enters the specimen identification information into the MALDI Biotyper CA System. The user first defines active sample positions (see section 4.2.1 of User Manual) and US IVD BTS control positions (see section 4.2.2 of User Manual). The defined sample positions are required to have a user entered and valid sample identifier in the Id column along with an optional description. All inoculated positions must contain a valid ID before the run can proceed. After all sample positions have been input, at least two US IVD BTS control positions must be defined. It is recommended that the cross-joint positions on the target are used as quality control positions.

Alternatively, the user can use an optional Honeywell (Hyperion 1300g) Barcode Reader USB cable is connected to the MALDI Biotyper CA System computer. The barcode reader scans the unique ten-digit target ID which appears in the Target ID box on the target plate. After the target ID has been entered, the a new Run page opens and the ten-digit target ID appears as the Plate Id and is appended to the Run name. Sample identifications are entered into the computer corresponding to the target plate position for that run.

4. Specimen Sampling and Handling:

After incubation of bacteria on recommended isolation media for 18-24 h at $(37^{\circ}C \pm 2^{\circ}C)$, colonies are stable for up to 12 h when held at room temperature.

Using a sterile colony transfer device, smear isolated colonies of bacteria as a thin film directly onto a sample position on a cleaned target. Inoculating an appropriate amount of the test organism onto the target is important. Excessive or insufficient amounts of inoculum may impact organism identification. The User Manual visually illustrates suitable and unsuitable inoculum amounts of organism on target.

Each of the sample positions and US IVD BTS control positions are overlayed with 1 μ L US IVD HCCA portioned solution. Use a new pipette tip to add matrix to each inoculated sample position. Dry the inoculated plate at room temperature. The inoculated MALDI target plate is now ready for use. For stability see Sections M.1.j and M.1.k above.

If the MALDI BIOTYPER CA System identification of the test organism does not result in a bacterial identification with a log(score) value of ≥ 2.0 , repeat testing using the extraction procedure in Section 3.6 of the User Manual. Briefly, the extraction of a sample from an isolated colony consists of multiple washing cycles of ethanol and water followed by addition of formic acid and acetonitrile followed by a centrifugation step. After centrifugation, a sample of the supernatant is applied to the target plate, dried and loaded into the MALDI Biotyper CA System for analysis.

- 5. <u>Calibration:</u> See K130831
- 6. <u>Quality Control</u>: See Section M.1.c above.

P. Other Supportive Instrument Performance Characteristics Data Not Covered In The "Performance Characteristics" Section above:

Q. Proposed Labeling:

The labeling is sufficient and satisfies the requirements of 21 CFR Part 809.10, 21 CFR 801.109, and special controls of regulation 21 CFR 866. 3361.

R. Conclusion:

The submitted information in this premarket notification is complete and supports a substantial equivalence decision.