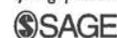


Safety Assessment of Microbial Polysaccharide Gums as Used in Cosmetics

International Journal of Toxicology
2016, Vol. 35(Supplement 1) 5S-49S
© The Author(s) 2016
Reprints and permission:
sagepub.com/journalsPermissions.nav
DOI: 10.1177/1091581816651606
ijt.sagepub.com



Monice M. Fiume¹, Bart Heldreth², Wilma F. Bergfeld³,
Donald V. Belsito³, Ronald A. Hill³, Curtis D. Klaassen³,
Daniel C. Liebler³, James G. Marks Jr³, Ronald C. Shank³,
Thomas J. Slaga³, Paul W. Snyder³, and F. Alan Andersen⁴

Abstract

The Cosmetic Ingredient Review Expert Panel assessed the safety of 34 microbial polysaccharide gums for use in cosmetics, finding that these ingredients are safe in cosmetic formulations in the present practices of use and concentration. The microbial polysaccharide gums named in this report have a variety of reported functions in cosmetics, including emulsion stabilizer, film former, binder, viscosity-increasing agent, and skin-conditioning agent. The Panel reviewed available animal and clinical data in making its determination of safety.

Keywords

safety, cosmetics, microbial polysaccharide gums

Introduction

As given in the *International Cosmetic Ingredient Dictionary and Handbook*, these 34 microbial polysaccharide gums function as emulsion stabilizers, film formers, binders, viscosity-increasing agents, and skin-conditioning agents¹:

Xanthan gum
Hydroxypropyl xanthan gum
Undecylenoyl xanthan gum
Dehydroxanthan gum
Xanthan gum crosspolymer
Xanthan hydroxypropyltrimonium chloride
Gellan gum
Welan gum
Biosaccharide gum-1
Biosaccharide gum-2
Biosaccharide gum-3
Biosaccharide gum-4
Biosaccharide gum-5
Pseudoalteromonas exopolysaccharides
Dextran
Carboxymethyl dextran
Dextran hydroxypropyltrimonium chloride
Sodium carboxymethyl dextran
Dextran sulfate
Sodium dextran sulfate
Sclerotium gum
Hydrolyzed sclerotium gum
Beta-glucan
Beta-glucan hydroxypropyltrimonium chloride

Beta-glucan palmitate
Hydrolyzed beta-glucan
Oxidized beta-glucan
Sodium carboxymethyl beta-glucan
Pullulan
Myristoyl pullulan
Levan
Rhizobian gum
Hydrolyzed rhizobian gum
Alcaligenes polysaccharides

The Cosmetic Ingredient Review (CIR) Expert Panel (Panel) reviewed other nonmicrobial gums and polysaccharides in 2012 and concluded the galactomannans, a group of 16 legume polysaccharides, are safe as used in cosmetics.² In 2009, the Panel reviewed the safety of hyaluronic acid, an amine-derived exopolysaccharide, finding it safe as used.³ In 1987, the Panel concluded that tragacanth gum (now named

¹ Cosmetic Ingredient Review Scientific Analyst/Writer, Cosmetic Ingredient Review, Washington, DC, USA

² Cosmetic Ingredient Review Chemist, Cosmetic Ingredient Review, Washington, DC, USA

³ Cosmetic Ingredient Review Expert Panel Member, Cosmetic Ingredient Review, Washington, DC, USA

⁴ Former Director, Cosmetic Ingredient Review, Washington, DC, USA

Corresponding Author:

Lillian J. Gill, Cosmetic Ingredient Review, 1620L Street, NW, Suite 1200,
Washington, DC 20036, USA.
Email: cirinfo@cir-safety.org

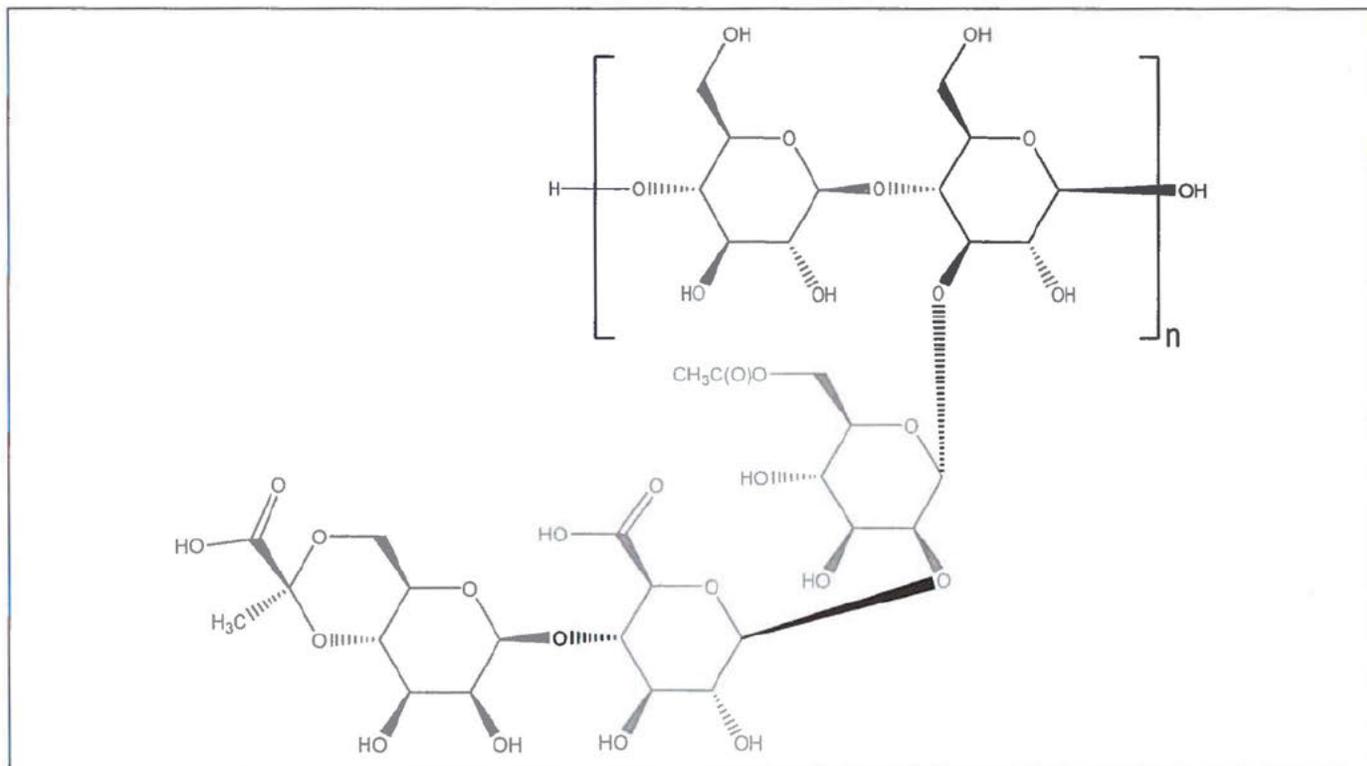


Figure 1. Xanthan gum—a polysaccharide composed of glucose, glucuronic acid, 6-acetyl mannose, and 4,6-pyruvylated mannose.

Astragalus gummifer gum) was safe as used; the Panel reaffirmed that conclusion in 2006.⁴ The Panel also concluded (2005) that Acacia senegal gum is safe as used but that the data are insufficient to support the safety of Acacia catechu gum and Acacia farnesiana gum as used in cosmetics.⁵

Three ingredients included in this safety assessment are hydroxypropyltrimonium chloride compounds. Although hydroxypropyltrimonium chloride itself is not a cosmetic ingredient, it is structurally analogous to some of the compounds included in the CIR safety assessment on trimoniums, which the Panel concluded are safe as used in cosmetics when formulated to be nonirritating.⁶

Some of these polysaccharide gums can be produced by more than one organism and sometimes by plants. For example, beta-glucans are produced by fungi, yeasts, and grains,⁷ and levan can be produced by bacteria, yeasts, or fungi.⁸

Many studies have been conducted with some of the microbial polysaccharide gums in regard to health claims, immunomodulatory activity, antioxidant activity, and so on. This safety assessment includes only studies and study types that relate directly to the safety of the cosmetic use of these ingredients.

Chemistry

Definition and Structure

Microbial polysaccharide gums are high-molecular-weight (MW) carbohydrate polymers that makeup a substantial component of the cellular polymers found in and surrounding most

microbial cells.⁹ These polysaccharide gums are produced by a wide variety of microorganisms and are water-soluble gums that have novel and unique physical properties. Microbial polysaccharide gums are generally divided into 3 groups: exocellular, cell wall, and intercellular.¹⁰ The exocellular polysaccharide gums are those that constantly diffuse into the cell culture medium and are easily isolated. The cell wall (ie, structural) and intercellular polysaccharide gums are integral parts of the cell wall or capsular products and are more difficult to separate from cell biomass.

Microbial polysaccharide gums may be ionic or nonionic and are primarily linear polysaccharides to which side chains of varying length and complexity are attached at regular intervals.⁹ Most microbial polysaccharide gums are linear heteropolysaccharides consisting of 3 to 7 different monosaccharides arranged in groups of 10 or less to form repeating units. The monosaccharides may be pentoses, hexoses, amino sugars, or uronic acids. For example, xanthan gum is a polysaccharide produced by a pure culture fermentation of a carbohydrate with *Xanthomonas campestris* and is composed of glucose, glucuronic acid, 6-acetyl mannose, and 4,6-pyruvylated mannose residues, as seen in Figures 1 and 2. The other ingredients in this report are related by having similar polymeric repeat units. The definitions and polymeric repeat units are provided in Table 1.

Physical and Chemical Properties

Available physical and chemical properties are provided in Table 2. The properties of the microbial polysaccharide gums

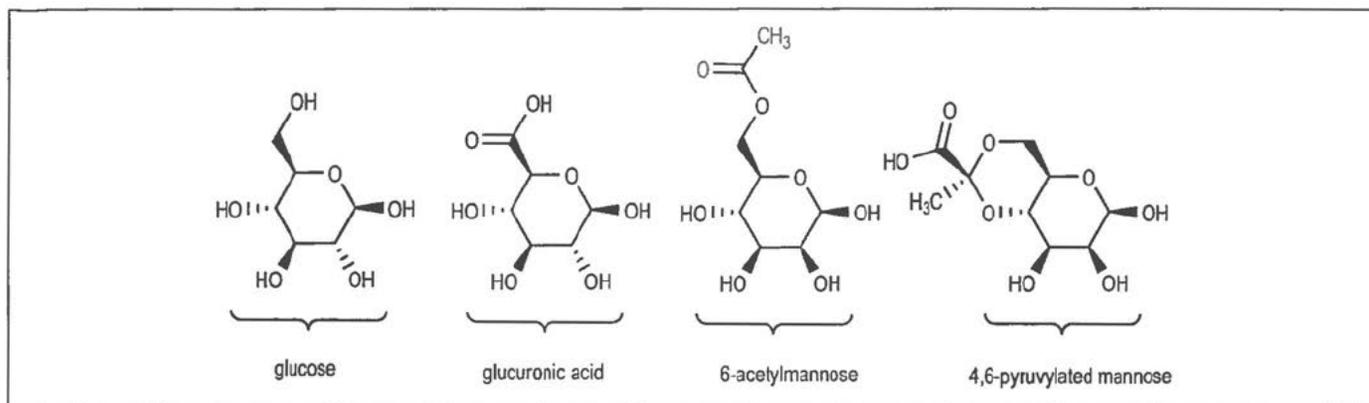


Figure 2. Glucose, glucuronic acid, 6-acetyl mannose, and 4,6-pyruvylated mannose, the monosaccharide components of xanthan gum.

can vary widely based on, among other parameters, the side groups, the ester substituents, or the bacterial strains.¹¹⁻¹⁴

Constituents/Impurities

The available constituent and impurity data are provided in Table 3.

Methods of Manufacture

Methods of manufacture for many of the microbial polysaccharide gums are provided in Table 4. Some of the polysaccharide gums discussed in this safety assessment can be produced by more than 1 organism and, in some cases, by plants. For example, beta-glucans are produced by fungi, yeasts, and grains,⁷ and levan can be produced by bacteria, yeasts, or fungi.⁸

Use

Cosmetic

The microbial polysaccharide gums named in this report have a variety of reported functions in cosmetics that include emulsion stabilizer, film former, binder, viscosity-increasing agent, and skin-conditioning agent.¹ The Food and Drug Administration (FDA) collects information from manufacturers on the use of individual ingredients in cosmetics as a function of cosmetic product category in its Voluntary Cosmetic Registration Program (VCRP). The VCRP data obtained from the FDA in 2012,¹⁵ and data received in response to a survey of the maximum reported use concentration by category conducted by the Personal Care Products Council (Council),^{16,17} indicate that 19 of the 34 microbial polysaccharide gums named in this safety assessment are currently used in cosmetic formulations. Xanthan gum is used in almost every category of cosmetic product, with 3,470 reported uses. Biosaccharide gum-1, sclerotium gum, and beta-glucan are reported to be used in 346, 193, and 137 cosmetic formulations, respectively. All other in-use ingredients have less than 70 uses. The ingredient with the highest concentration of use is pullulan; it is used at up to

12% in leave-on formulations (ie, tonics, dressings, and other hair-grooming aids) and 17% in "other" oral hygiene products (a breath freshener that dissolved in the mouth¹⁸). Both xanthan gum and biosaccharide gum-1 are used at up to 6% in leave-on formulations, and xanthan gum crosspolymer and biosaccharide gum-4 are used at 5% in leave-on formulations. All other in-use ingredients are used at concentrations of $\leq 3\%$.

In some cases, reports of uses were received in the VCRP but no concentration of use is available. For example, sodium carboxymethyl dextran is reported to be used in 10 formulations, but no use concentration data were available. In other cases, no reported uses were received in the VCRP, but a use concentration was provided in the industry survey. For example, hydrolyzed sclerotium gum was not reported in the VCRP to be in use, but the industry survey indicated that it is used in leave-on formulations at up to 1%. It should be presumed that hydrolyzed sclerotium gum is used in at least 1 cosmetic formulation.

Frequency and concentration of use data are provided in Table 5. The ingredients not listed in the VCRP or by the Council as being used are listed in Table 6.

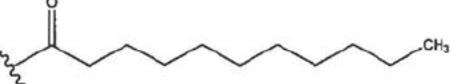
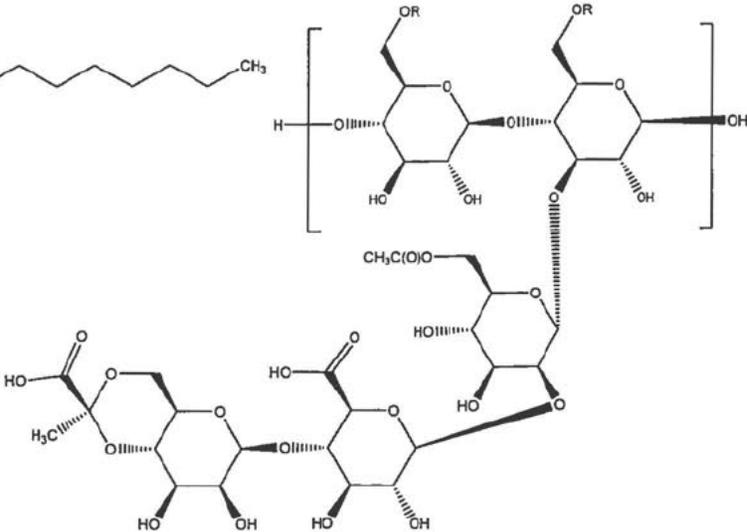
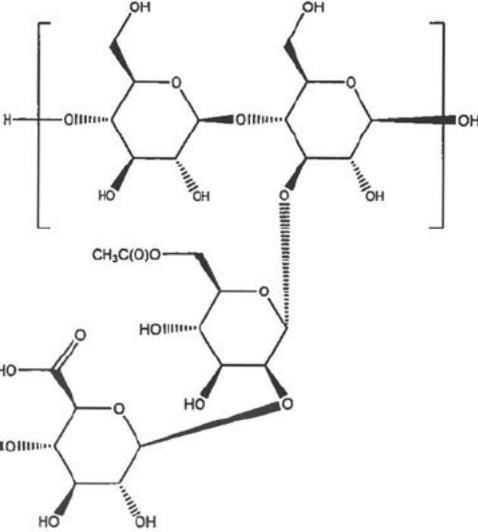
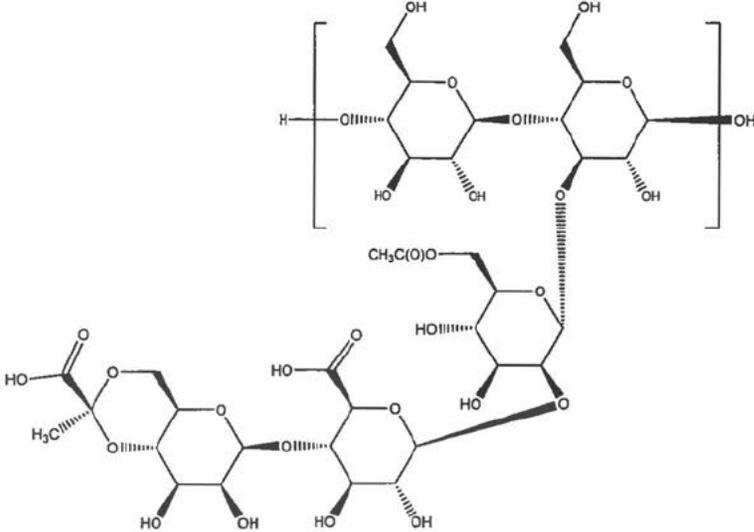
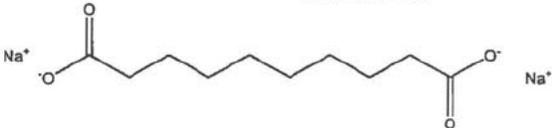
Products containing some of the microbial polysaccharide gums are reported to be used on baby skin, to be applied to the eye area or mucous membranes, or could possibly be ingested. Some of these ingredients are reported to be used in product types that may be inhaled; for example, dehydroxanthan gum is used in a face and neck spray at 0.2%. In practice, 95% to 99% of the particles released from cosmetic sprays have aerodynamic equivalent diameters in the 10 to 110 μm range.^{19,20} Therefore, most particles incidentally inhaled from these sprays are deposited in the nasopharyngeal region and are not respirable.^{21,22} Xanthan gum is reported to be used in deodorants at up to 0.6%, and it is not known whether these products are sprayed. There is some evidence indicating that deodorant spray products can release substantially larger fractions of particulates having aerodynamic diameters in the range considered to be respirable.²² However, the information is not sufficient to determine whether significantly greater lung exposures result from the use of deodorant sprays compared to other cosmetic sprays.

Table 1. Definition, Function, and Idealized Structure.

Ingredient; CAS No.	Definition	Reported function(s) ¹	Idealized structure
Xanthan gum; 11138-66-2	A high MW heteropolysaccharide gum produced by a pure-culture fermentation of a carbohydrate with <i>Xanthomonas campestris</i> , ¹ composed of glucose, glucuronic acid, 6-acetyl mannose, and 4,6-pyruvylated mannose residues ⁷²	Binder; emulsion stabilizer; skin-conditioning agent—miscellaneous; surfactant-emulsifying agent; viscosity-increasing agent—aq.	(Given below definition)
Hydroxypropyl xanthan gum; 106442-37-9	the hydroxypropyl ether of xanthan gum, ¹ wherein ether substitution occurs primarily at 6-position of the backbone sugar residues	Emulsion stabilizer; film former; viscosity-increasing agent—aq.	

(continued)

Table I. (continued)

Ingredient; CAS No.	Definition	Reported function(s) ¹	Idealized structure
Undecylenoyl xanthan gum	The condensation product of undecylenic acid chloride and xanthan gum, ¹ wherein esterification occurs primarily at the 6-position of backbone sugar residues	emulsion stabilizer; hair-conditioning agent	<p>wherein R = H or </p> 
Dehydroxanthan gum; 11138-66-2 (dehydro is colisted under this CAS no. with xanthan gum)	The product obtained by the dehydration of xanthan gum ¹	Emulsion stabilizer; film former; hair fixative; suspending agent—nonsurfactant; viscosity-increasing agent—aq.	
Xanthan Gum Crosspolymer	xanthan gum crosslinked with disodium sebacate ¹	Skin protectant; skin-conditioning agent—miscellaneous	 <p>Crosslinked with:</p> 

(continued)

Table I. (continued)

Ingredient; CAS No.	Definition	Reported function(s) ¹	Idealized structure
Xanthan hydroxypropyl trimonium chloride	The quaternary ammonium salt formed by the reaction of xanthan gum with 2,3-epoxypropyltrimethylammonium chloride	Hair-conditioning agent; skin-conditioning agent—humectant; viscosity-increasing agent—aq.	<p>wherein R = H or </p>
Other gums Gellan gum; 71010-52-1	A high MW heteropolysaccharide gum produced by pure-culture fermentation of a carbohydrate with <i>Pseudomonas elodea</i> , ¹ consists of the basic backbone (→3)-β-D-Glc-(1 → 4)-β-D-GlcA-(1 → 4)-β-D-Glc-(1 → 4)-α-L-Rha-(1 →); ¹² composed of glucose (some of which is glyceryl and/or acetyl esterified), glucuronic acid, and rhamnose residues ¹³	Emulsion stabilizer; viscosity-increasing agent—aq.	

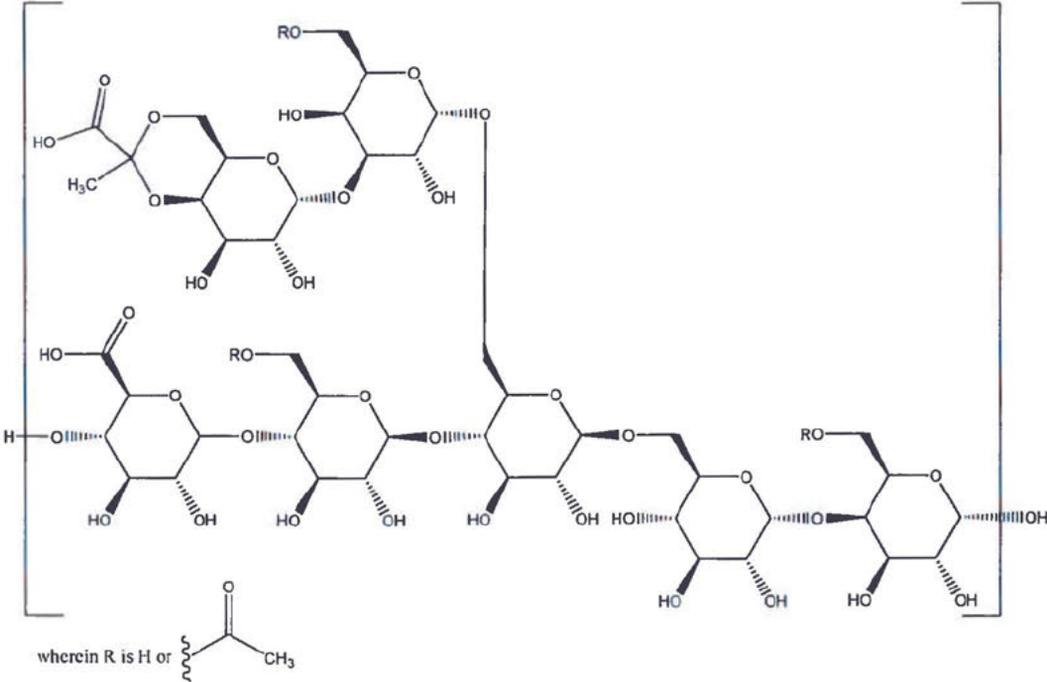
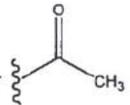
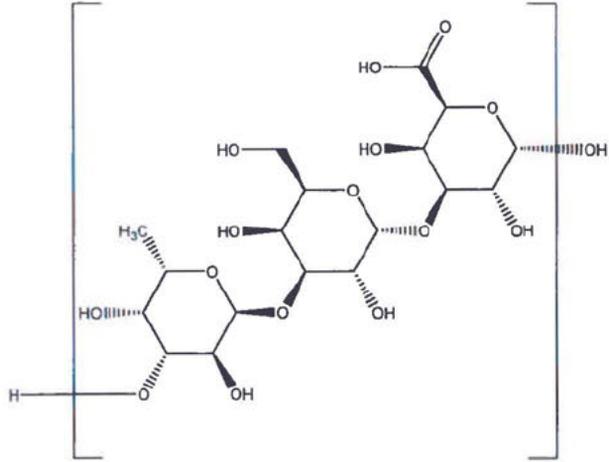
(continued)

Table I. (continued)

Ingredient; CAS No.	Definition	Reported function(s) ¹	Idealized structure
Welan gum; 96949-22-3	A gum produced by the fermentation of <i>Alcaligenes</i> ; ¹ consists of the basic backbone $\rightarrow 3$ - β -D-Glcp-(1 \rightarrow 4)- β -D-GlcpA-(1 \rightarrow 4)- β -D-Glcp-(1 \rightarrow 4)- α -L-Rhap-(1 \rightarrow 3)- α -L-Rhap or α -L-Manp; composed of glucose, glucuronic acid, rhamnose, and mannose residues, with rhamnopyranosyl and/or L-mannopyranosyl side chains ⁷³	Binder; emulsion stabilizer; film former; viscosity-increasing agent—aq.	
Sclerotium gum; 39464-87-4	The polysaccharide gum produced by the fungus, <i>Sclerotium rolfsii</i> , ¹ consists of the basic backbone of β -D-(1 \rightarrow 3)-glucopyranosyl units with a β D glucopyranosyl unit (1 \rightarrow 6) linked to every third unit ⁷⁴	Emulsion stabilizer; skin-conditioning agent—miscellaneous; viscosity-increasing agent—aq.	
Hydrolyzed sclerotium gum	The partial hydrolysate of sclerotium gum derived by acid, enzyme, or other method of hydrolysis ¹	Film former; skin protectant; skin-conditioning agent—humectant	

(continued)

Table I. (continued)

Ingredient; CAS No.	Definition	Reported function(s) ¹	Idealized structure
Rhizobian gum; 127712-52-1 (for rhizobium gum)	The polysaccharide gum produced by the fermentation by <i>Rhizobium bacterium</i> , ¹ composed of glucose, glucuronic acid, galactose, and pyruvylated galactose residues, with some acylation ⁷⁵	Film former; hair fixative; plasticizer; suspending agent—nonsurfactant; viscosity-increasing agent—aq	 <p>wherein R is H or </p>
Hydrolyzed rhizobian gum	The partial hydrolysate of rhizobian gum derived by acid, enzyme, or other method of hydrolysis ¹	Film former	
Biosaccharide gum-1; 223266-93-1	A fermentation gum derived from sorbitol; described as a polymer of α -L-fucose (1 \rightarrow 3) α -D-galactose(1 \rightarrow 3)- α -D-galacturonic acid trisaccharides; ¹ composed of fucose, galactose, and galacturonic acid residues	Skin-conditioning agent—miscellaneous	

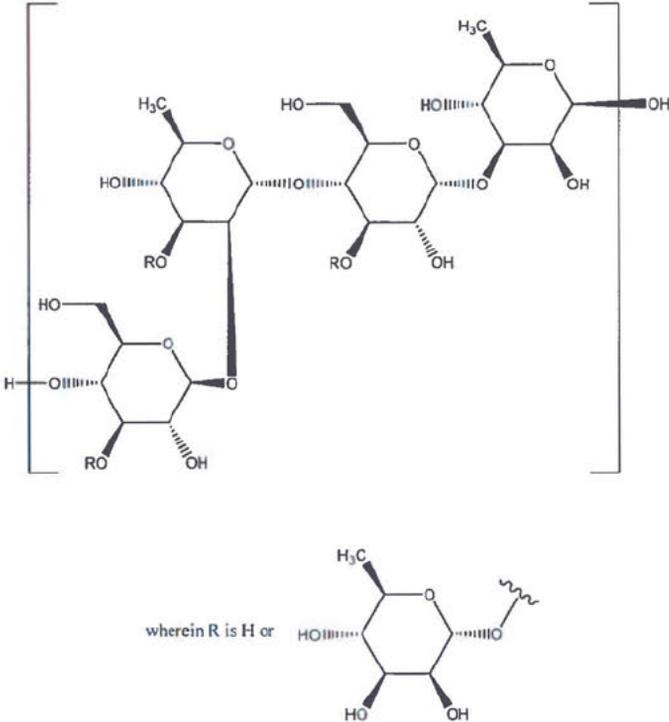
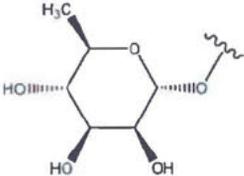
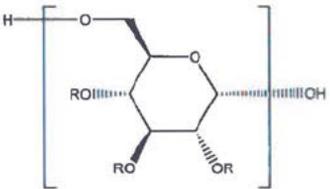
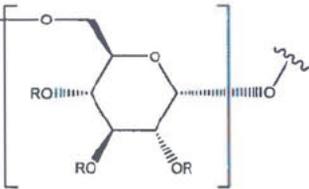
(continued)

Table 1. (continued)

Ingredient; CAS No.	Definition	Reported function(s) ¹	Idealized structure
Biosaccharide gum-2; 283602-75-5; 758716-52-8	A fermentation gum derived from sorbitol; described as a polymer of α -L-Rhap(1 \rightarrow 3)- β -D-Galp-(1 \rightarrow 2)- α -L-Rhap-(1 \rightarrow 4)- β -D-GlepA (1 \rightarrow 3)-[α -L-Rhap-(1 \rightarrow 2)]- α -D-Galp-(1 ¹); composed of rhamnose, galactose, and glucuronic acid residues)	Skin-conditioning agent—miscellaneous	
Biosaccharide gum-3; 896736-76-8	A fermentation gum derived from sorbitol; described as a polymer of α -L-fucose(1 ³)- α -D-galactose(1 ³)- α -D-galacturonic acid trisaccharides and is characterized by a smaller degree of polymerization and MW in comparison to biosaccharide gum-1 ¹	Skin-conditioning agent—miscellaneous	
Biosaccharide gum-4; 905593-86-4	A fermentation gum derived from sorbitol; it is a deacetylated branched polymer consisting of L-fucose, 2-D-glucose and glucuronic acid repetitive units ¹	Skin-conditioning agent—miscellaneous	

(continued)

Table I. (continued)

Ingredient; CAS No.	Definition	Reported function(s) ¹	Idealized structure
Biosaccharide gum-5	A fermentation gum derived from sorbitol; described as a polymer of -glucose- β -(1 \rightarrow 2)-rhamnose- α -(1 \rightarrow 4)-glucose- α -(1 \rightarrow 3)-rhamnose- β -(1 \rightarrow 4), with rhamnose - α -(1 \rightarrow 3) branching ¹	Skin-conditioning agent—miscellaneous	 <p style="text-align: center;">wherein R is H or </p>
Pseudoalteromonas exopolysaccharides Dextran; 9004-54-0	The exopolysaccharide excreted by the fermentation of <i>Pseudoalteromonas</i> A high MW glucose polymer produced by the action of the bacteria, <i>Leuconostoc mesenteroides</i> , on a sucrose substrate ¹ ; the degree of branching is dependent on the source of dextrans and can vary from 0.5% to 60% ⁷⁶	Skin-conditioning agent—miscellaneous Binder; bulking agent	 <p style="text-align: center;">wherein R = H or a branch of </p>

(continued)

Table I. (continued)

Ingredient; CAS No.	Definition	Reported function(s) ¹	Idealized structure
Carboxymethyl dextran; 9044-05-7	The carboxymethyl derivative of dextran ¹	Film former; viscosity increasing agent—aq.	
	wherein R = H, , or a branch of		
Dextran hydroxypropyltrimonium chloride; 83855-79-2	The quaternary ammonium compound formed by the reaction of dextran with glycidyltrimethylammonium chloride ^{1,77}	Hair-conditioning agent	
	wherein R = H, , or a branch of		
Sodium carboxymethyl dextran; 39422-83-8	The sodium salt of a carboxymethyl dextran ¹	Binder; emulsion stabilizer; viscosity-increasing agent—aq.	
	wherein R = H, Na , or a branch of		

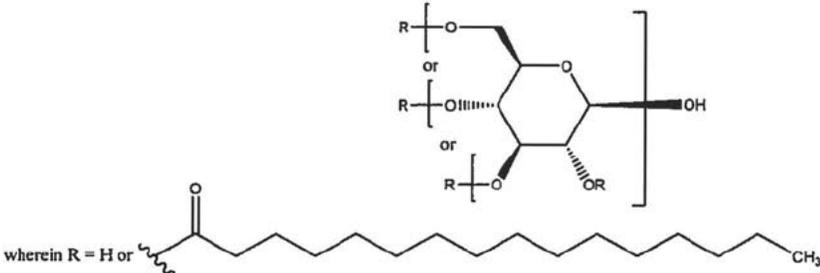
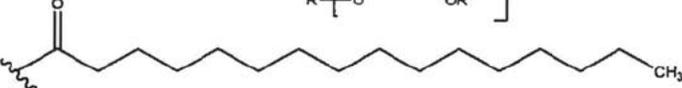
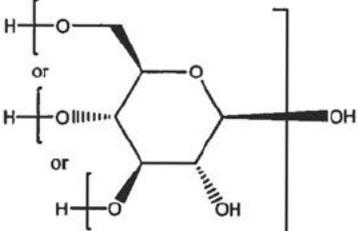
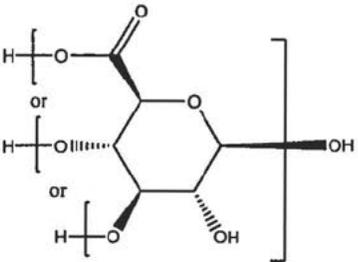
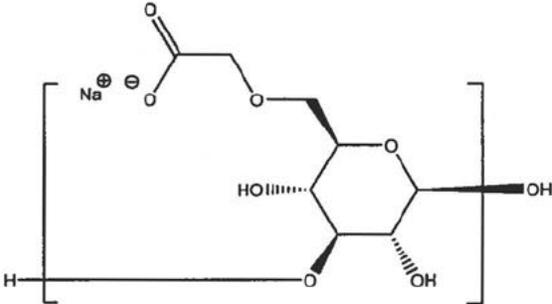
(continued)

Table I. (continued)

Ingredient; CAS No.	Definition	Reported function(s) ¹	Idealized structure
Dextran sulfate; 9042-14-2	The sulfuric acid ester of dextran ¹	binder; skin conditioning agent—miscellaneous	
Sodium dextran sulfate; 9011-18-1	The sodium salt of the sulfuric acid ester of dextran ¹	Suspending agent—nonsurfactant	
Beta-glucan; 55965-23-6 (CAS No. is specific to (1 → 3) (1 → 4)); 53238-80-5 (CAS No. is specific to (1 → 3) (1 → 6))	A polysaccharide consisting of β(1-3), β(1-4), and β(1-6) linked glucose units ¹	Bulking agent; skin-conditioning agent—miscellaneous	
Beta-glucan hydroxypropyltrimonium chloride	The quaternary ammonium compound formed by the reaction of beta-glucan with glycidyltrimethylammonium chloride	Antistatic agent; hair-conditioning agent; skin-conditioning agent—miscellaneous	

(continued)

Table I. (continued)

Ingredient; CAS No.	Definition	Reported function(s) ¹	Idealized structure
Beta-glucan palmitate	The product obtained by the condensation of beta-glucan with palmitoyl chloride ¹	Skin-conditioning agent—miscellaneous	 <p>wherein R = H or </p>
Hydrolyzed beta-glucan	The partial hydrolysate of beta-glucan, derived by acid, enzyme, or other method of hydrolysis	Skin-conditioning agent—miscellaneous	
Oxidized beta-glucan	The product obtained by the oxidation of beta-glucan, ¹ wherein the 6-position is oxidized to the acid ⁷⁸	Antiacne agent; antidandruff agent; antifungal agent; antimicrobial agent; deodorant agent; exfoliant; skin-conditioning agent—miscellaneous	
Sodium carboxymethyl beta-glucan; 9050-93-5 (CAS No. specific to (1 → 3) β D-Glucan)	The sodium salt of a carboxymethyl ether of beta-glucan, wherein the ether substitution occurs primarily at the 6-position ⁷⁹ ; 3 of 4 glucose units of the β-1,3-glucopyranose are carboxymethylated ⁸⁰	Binder; viscosity-increasing agent—miscellaneous	

(continued)

Table I. (continued)

Ingredient; CAS No.	Definition	Reported function(s) ¹	Idealized structure
Pullulan; 9057-02-7	An extracellular polysaccharide produced from starch by cultivating the yeast, <i>Aureobasidium pullulans</i> ; composed of 1 → 6 linked maltitriose residues (a maltitriose is a block of three 1 → 4 linked, glucose residues) ⁸¹	Binder; film former	
Myristoyl pullulan; 1033464-89-9	The reaction product of myristoyl chloride and pullulan ¹	Film former; hair fixative; hair-waving/straightening agent	
Levan; 9013-95-0	A polyfructose (fructofuranose), with some branching, produced by enzymatic action on sucrose ⁸¹ ; the degree of branching varies by organism but has been reported as high as 20% ⁸²	Film former; skin protectant; viscosity-increasing agent—aq.	

(continued)

Table I. (continued)

Ingredient; CAS No.	Definition	Reported function(s) ¹	Idealized structure
Alcaligenes polysaccharides; 188846-47-1 (for the pure disaccharide repeat unit)	The polysaccharides produced by a bacterial culture of <i>Alcaligenes latus</i> , composed of mannose and fucose ⁶³	Emulsion stabilizer; humectant; skin-conditioning agent—humectant; viscosity-increasing agent—aq.	

Abbreviations: Aq., aqueous; MW, molecular weight.

All of the microbial polysaccharide gums named in the report are listed in the European Union inventory of cosmetic ingredients.²³

Noncosmetic

Noncosmetic uses of microbial polysaccharide gums are summarized in Table 7. Some of the food and medical use information is described in the following paragraphs.

Xanthan gum²⁴ and gellan gum²⁵ are approved as direct food additives in gums, chewing gum bases, and related substances. Xanthan gum is also approved as an indirect food additive.²⁶ Beta-glucan (as curdlan, a specific beta-glucan that is a linear polymer consisting of β -(1 \rightarrow 3)-linked glucoside residues) is approved as a direct food additive for multipurpose addition.²⁷ Dextran is an indirect food additive that is generally recognized as safe.²⁸ The World Health Organization concluded that studies on the safety of xanthan gum,²⁹ gellan gum,³⁰ and pullulan³¹ provided sufficient information to be allocated an acceptable daily intake of "not specified." Pullulan appears in the Japanese List of Existing Food Additives.³²

Xanthan gum is used as a stabilizer, thickener, and emulsifying agent in water-based pharmaceutical preparations.³³ Dextran, as dextran 70, is an approved active ingredient for over-the-counter use as an ophthalmic demulcent at 0.1% when used with another approved polymeric demulcent.³⁴ Dextran is used as a plasma volume expander (as dextran 70) and as a blood flow adjuvant (as dextran 40).³⁵ Sodium dextran sulfate is used as a clinical reagent.

Toxicokinetics

Absorption, Distribution, Metabolism, and Excretion

Dermal

In Vitro

Beta-Glucan. A single application of 5 mg/cm² of a 0.5% (oat) beta-glucan solution was applied to human abdominal skin.³⁶ Beta-glucan penetrated the skin into the epidermis and dermis (no details were provided).

Human

Dextran. Fluorescent dextrans (FDs) in aqueous (aq.) solution were used to determine the dermal absorption of different MW dextrans (MW 3,000-70,000, identified as FD-3 and FD-70, respectively) through the skin that has been subjected to mini erosion.³⁷ Prior to testing, absorption of FD-20 from a test "Cellpatch" (cell) into systemic circulation via a mini-erosion site was verified in 2 male subjects and 1 female subject. The largest of the test molecules filtered from the blood into the urine was 20,000 MW, and with increasing erosion diameter, absorption increasingly occurs via the lymphatic system. Each subject received a single cell containing 0.5 mL solution (5 mmol). After 24 hours, a mean of 54.3% of the total FD-20 dose had been absorbed, and 12.6% of the absorbed dose was recovered in the urine.

The effect of molecular size on absorption of FDs was then determined. Four cells with 100 μ L of FD-3, FD-10, FD-20, or FD-70 in 0.5 mL isotonic saline were applied for 24 hours to 6 mm mini-eroded sites on 4 male and 3 female subjects. A fifth

Table 2. Chemical and Physical Properties.

Property	Description
Xanthan gum	
Appearance	Cream-colored, odorless, free-flowing powder ³⁵ ; white/beige powder with a characteristic odor ⁸⁴
MW	1,000,000-1,000,000 ³³ ; varies within a very wide range ⁸⁵
Solubility	Dissolves readily in water with stirring to give highly viscous solutions at low concentrations ³⁵ ; completely soluble in water, forming colloidal solution; insoluble in alcohol ⁸⁴ ; readily soluble in hot or cold water to form neutral, viscous, and nonthixotropic solutions that have relatively high viscosity ³³
Stability	Resistant to heat degradation ³⁵
Ionic nature	Anionic ^{10,81}
pH	5.5-8.5 (1% solution, 25°C) ⁸⁴
Gellan gum	
Appearance	Off-white powder ⁸⁵⁻⁸⁷
Types	Native, deacetylated, clarified (ie, filtered deacetylated gum) ⁸⁶
MW	Varies within a very wide range ⁸⁵ ; ~500,000, ⁸⁶ >70,000, with 95% above 500,000 ³⁹
Solubility	Soluble in hot or cold deionized water ⁸⁷ ; soluble in water; insoluble in ethanol ⁸⁶
Viscosity	Can exhibit high viscosity in solution ¹² ; high-acyl gellan gum is viscous; deacetylated gellan gum (treated with an alkali) has relatively low solution viscosity ¹⁴ ; cold dispersions of native gellan gum provide extremely high viscosities, and the solutions are highly thixotropic; the viscosity decreases with heating as the gum hydrates; hot native gum solutions are more viscous than deacetylated gellan gum solutions ¹⁴
Gelling property	Forms a weak gel in water in its native state but forms a rigid gel after treatment with an alkali ¹³
Ionic nature	Anionic ^{10,81}
Hydration	Native (acylated) gellan gum will swell in deionized water forming a very thick particulate system, and the hydration temperature is reached at ~70°C; the swelling behavior and hydration temperature are altered in the presence of ions ¹⁴ ; deacetylated gum will only partially hydrate in cold deionized water, with hydration occurring with a heated deionized water temperature of ~70°C; also hydration is poor in mildly acidic conditions and in the presence of divalent ions ¹⁴
Welan gum	
MW	865,000-932,000 ⁷³
Viscosity	Nongelling polysaccharide forming highly viscous aq. solutions ⁸⁸
Ionic nature	Anionic ¹⁰
Biosaccharide gum-1	
MW	1,000,000 (average) ⁸⁹
Biosaccharide gum-4	
MW	2,000,000 (average) ⁸⁹
Dextran	
Appearance	Dextran 1: white to off-white powder ³⁵
MW	Dextran 1: ~1,000 (average) ⁹⁰ ; dextran 40: ~25,000 (solution has an average MW of 40,000) ⁷¹ ; dextran 70: ~40,000 (solution has an average MW of 70,000) ⁷¹ ; MW distribution is dependent on the source of the dextran ⁷⁶
Solubility	Dextran 1: very soluble in water; sparingly soluble in alcohol ³⁵ ; degree of solubility decreases with an increase in the degree of branching; dextrans with >43% branching are insoluble ⁷⁶
Ionic nature	Nonionic ^{10,81}
Stability	Stable under mild acidic and basic conditions ⁷⁶
pH	Dextran 1: 4.5-7.0 (15% aq. solution) ⁹⁰
Specific rotation	Dextran 1: between +148° and +164° at 20°, for an aq. solution ⁹⁰ ; dextran 40: between +195° and +203° ⁹⁰ ; dextran 70: between +195° and +203° ⁹⁰
Dextran sulfate	
MW	5,000-500,000 ¹¹
Sodium dextran sulfate	
Appearance	White powder ³⁵
MW	4,000-50,000 ³⁵
Solubility	Freely soluble in water ³⁵
Sclerotium gum	
Solubility	Disperses easily in water at room temperature; refined grades dissolve readily in hot and cold water ⁹
Ionic nature	Nonionic ^{10,81}
Beta-glucan	
Appearance	White to pale yellow powder with a slight odor ⁹¹ ; white to nearly white powder (as curdlan) ⁸⁷
MW	~500,000 (native state) ⁹²

(continued)

Table 2. (continued)

Property	Description
Solubility	As curdlan: insoluble in water, alcohol, and most organic solvents; soluble in alkaline solutions ⁹
Ionic nature	Nonionic ¹⁰
Oxidized beta-glucan	
MW	Continuum of ~30,000 to >70,000 ²²
Sodium carboxymethyl beta-glucan	
Appearance	White/brown solid ⁹³ ; amber, white granulate with a characteristic isopropyl alcoholic odor ⁸⁰
MW	~100,000 ⁹³ ; 4.23×10^5 (average) ⁷⁹
Degree of substitution	$0.75 \pm 1^{80,93}$ (as a 2% aq. solution); 0.2-0.3 ⁷⁹
Solubility	Solubility up to 98% ⁷⁹
pH (2% aq. solution)	~7 ⁹³
Pullulan	
Appearance	White to slightly yellowish powder; tasteless; odorless (food grade) ⁹⁴ ; tasteless, odorless fine white powder ⁹⁵
MW	Can vary considerably; a commercially available product has a molecular mass of 200,000 Da ³¹ ; 8,000 to >2,000,000; approximately 200,000 (mean) ⁹⁴
Solubility	Highly soluble in cold or hot water; not soluble in organic solvents, except dimethylformamide or dimethyl sulfoxide (DMSO) ^{32,94}
Stability	Stable in aq. solution over a wide pH range; decomposes upon dry heating, carbonizing at 250°C-280°C ³¹
Viscosity	Dissolves in water producing a stable viscous solution; does not gel; viscosity is proportional to MW ⁹⁴ ; solutions are of relatively low viscosity; viscosity is stable to heating, changes in pH, and most metal ions ⁹⁶
Ionic nature	Nonionic ^{10,81}
pH	5.0-7.0 (food grade) ⁹⁴
Refractive index	Significant positive linear correlation of concentration and refractive index at 20°C and 45°C ⁹⁴
Levan	
MW	Up to several million Daltons; typically 2×10^6 to 10^8 ; usually >0.5 million and can be as high as 40,000,000 ^{8,82}
Particle size	Partially forms nanoparticles in water; 224.3 nm in water and 251.8 nm in ethanol ⁵⁷
Solubility	Highly soluble in water at room temperature ⁸ ; water soluble; does not swell in water ⁸²
Viscosity	"Exceptionally low" intrinsic viscosity ⁸²
Ionic nature	Nonionic ⁸¹
Rhizobian gum	
MW	1,500,000 (native molecule, in the fermentation broth) ⁷⁵
Melting point	~60°C; gets lower after sterilization ⁹⁷
Viscosity	With a 10 g/L solution, the viscosity decreases as the pH increases ⁷⁵
Alcaligenes polysaccharides	
MW	64,000 ⁸³

Abbreviations: Aq., aqueous; MW, molecular weight.

cell, without FD, was applied as a negative control. The FD concentration in each cell was measured using spectrofluorometry at various times. The dextrans were readily absorbed, but absorption decreased with increasing molecular size. The absorption of FD-3 was 37.9%, but for FD-70, it was 20.1%. Further testing using 3 male and 3 female subjects determined that the degree of transdermal absorption was directly related to the area of erosion; 20.5% of FD-3 absorbed through a 3-mm erosion area, whereas 60.7% of the same molecular size FD absorbed through a 10-mm erosion.

Oral.

Nonhuman

Xanthan gum. Rats were fed a diet containing 2% [¹⁴C]xanthan gum that was produced by fermentation of uniformly labeled glucose with *Xanthomonas campestris*.³⁸ No accumulation was found in the tissues. A maximum of 15% of the radioactivity was metabolized to carbon dioxide within

100 hours. Fecal analysis indicated that there was no accumulation of the polysaccharide material, except acetate (acetate and pyruvate accounted for only 9.8% of the label in the gum used). In the feces, 98% of the radioactivity was attributed to unchanged or slightly modified polysaccharide. In vitro testing indicated that nonenzymatic hydrolysis and fecal microorganisms were responsible for the in vivo breakdown of xanthan gum (no additional details were provided).

Gellan gum. In animal feeding studies using radiolabeled gellan gum, the majority of the gellan gum administered was recovered in fecal matter.¹⁴ This appears to indicate that no endogenous enzymes that are able to break down gellan gum are present in the small intestine (no additional details were provided).

The absorption, distribution, and excretion of gellan gum were determined in studies using a dually radiolabeled gum that was prepared in separate fermentations using [³H]glucose and [¹⁴C]glucose as carbon sources.³⁹ The ³H product was

Table 3. Constituents/Impurities.

Ingredient	Constituents/impurities
Xanthan gum	Nitrogenous constituents equal to ~1% nitrogen by wt; approximately half of the nitrogenous matter is proteinaceous and contains amino acid residues in the same proportions as other food-grade gums; the remainder is present as amino sugars, nucleic acids, and nucleotides ²⁹ Food grade contains D-glucose and D-mannose as the dominant hexose units, and D-glucuronic acid and pyruvic acid; NMT 2 mg/kg lead; NMT 0.075% ethanol and isopropyl alcohol, singly or combined ⁹⁸ 2.5%-4.8% pyruvic acid, present as side chains ³³
Gellan gum	Usually contains a small amount of nitrogen-containing compounds as a result of the fermentation procedure ⁸⁶ <2 mg/kg lead; <3% nitrogen; 750 mg/kg isopropyl alcohol ⁸⁶ Native: can contain 10% protein and 7% ash; deacetylated: can contain 17% protein and 8% ash ⁸⁶ Gellan gum is characterized by the polysaccharide content, percentage of O-acetylated substitution, and protein content (including nucleic acid residues and other organic nitrogen sources) ³⁹
Dextran sulfate	Can contain polymers of various MWs and degrees of sulfation ⁴¹
Beta-glucan	≥85% β-1,3-1,6-glucan, sucrose, yeast extract, minerals, <i>Aerobasidium pullulans</i> ⁹¹ As curdlan: ≥90% carbohydrate; ≤10% water ⁹ As food grade curdlan—NMT 0.5 mg/kg lead; microbial limits, aerobic plate count NMT 1,000 colony-forming unit/g; <i>Escherichia coli</i> , negative in 1 g ⁹⁸
Sodium carboxymethyl beta-glucan	~90% sodium carboxymethyl beta-glucan; <0.5% nitrogen (protein); <1.0% glycolic acid; <0.05% chloroacetic acid; ~10% volatile matter ⁸⁰
Pullulan	≥90% glucan on a dried basis; main impurities are mono-, di-, and oligosaccharides from the starting material ³¹ >90% pullulan; <10% mono-, di-, and oligosaccharides; <0.1 ppm lead; <2 ppm arsenic; <5 ppm heavy metals (food grade) ⁹⁴

subjected to multistage purifications for a relatively pure [³H]polysaccharide, which was then added to the ¹⁴C fermentation, giving a polysaccharide fraction that was dual labeled and a nonpolysaccharide fraction labeled only with ¹⁴CO₂. In the first study, 1 male and 1 female Sprague Dawley rats were dosed by gavage with a single dose of 960 mg/kg [³H/¹⁴C]gellan gum (4 μCi). Expired air was collected for 24 hours after dosing, and <0.55% of the dosed radioactivity was detected as ¹⁴C.

Four male and 3 female Sprague Dawley rats were then given a single dose by gavage of 870 mg/kg [³H/¹⁴C]gellan gum (2.9-4.1 μCi ¹⁴C; 0.7-0.9 μCi ³H). Urine and feces were collected for 7 days. Approximately 86% of the dosed ¹⁴C was excreted in the feces and 2% to 3% in the urine, and approximately 100% of the dosed ³H was excreted in the feces and 4% was in the urine. Tissue and carcass ¹⁴C radioactivity was approximately 3% to 4% of the dose. The ³H activities in the tissues were below the limits of accurate quantification.

In the last study, 1 male and 4 female Sprague Dawley rats were dosed with 1 g/kg [³H/¹⁴C]gellan gum by gavage, and blood samples were taken at various intervals over a 7-day period (it is not stated but it appears that 1 dose was administered). The peak level of radioactivity in blood, equivalent to 0.4% of the dose, occurred about 5 hours after dosing.

Dextran. Groups of 5 fasted male Sprague Dawley rats were given a single dose by gavage of 50 mg/kg fluorescein-labeled dextrans (FD-4: 4,400 average MW; FD-20: 19,000 average MW; or FD-40: 40,500 average MW).⁴⁰ The dextran solution was prepared as 25 mg/mL in isotonic phosphate buffer. Blood samples were taken at various intervals for up to 4 hours after dosing with FD-4, up to 8 hours after dosing with FD-20, and for up to 24 hours with FD-40. Urine samples were taken at intervals for up to 8 hours after dosing with FD-4 and FD-20 and for up to 24 hours after dosing with FD-40. None of the dextrans were detected in the serum after oral administration. Small amounts of the dose, ranging from 0.308% with FD-4 to 0.0138% FD-40, were detected in the urine. The oral bioavailability was 0.398%, 0.0728%, and 0.0431% for FD-4, FD-20, and FD-40, respectively.

Dextran sulfate. Two groups of 5 male Wistar rats were dosed by gavage with 5 mg/mL of 20 mg/kg dextran sulfate containing 12 μCi ³H-dextran sulfate/kg (8,000 average MW), and each group was killed 3 or 24 hours after dosing.⁴¹ Most of the radioactivity was detected in the feces; 22.5% of the dose was recovered after 24 hours. Metabolites, breakdown products, and ³H₂O were not found in the feces, and the researchers stated that it was most likely that the dose recovered in the feces was unabsorbed dextran sulfate. Approximately 10% of the dose was recovered in the urine after 24 hours, with 6% recovered after 3 hours. Urinary ³H elution profiles indicated that intermediate MW metabolites or breakdown products were formed and that either intact or partially intact dextran sulfate was absorbed through the epithelium of the gastrointestinal (GI) tract. The 6- to 24-hour samples indicated a marked shift toward smaller MW products.

Beta-glucan. Two male Sprague Dawley rats were dosed orally with 20 mg/kg body weight (BW) [U-¹⁴C]beta-glucan (as curdlan) in water prepared from [U-¹⁴C]glucose.⁴² Most of the radioactivity was recovered in expired CO₂; 77% of the dose was recovered in 24 hours and 89% in 72 hours. A total of 7.7% and 12% of the radioactivity as administered dose was recovered in the feces after 24 and 72 hours, respectively, and 2.6% and 3.3% was recovered in the urine after 24 and 72 hours, respectively.

Table 4. Methods of Manufacture/Organisms Used in Production.

Ingredient	Method
Xanthan gum	Pure-culture fermentation of glucose or corn syrup from the bacterium <i>Xanthomonas campestris</i> ; the polysaccharide is recovered by precipitation and purification with isopropyl alcohol, followed by drying and milling ^{33,60}
Gellan gum	Aerobic submerged fermentation using the bacterium <i>Pseudomonas elodea</i> ; small seed fermentation is followed by pasteurization; the gum is recovered by precipitation with isopropyl alcohol, followed by drying and milling ⁹⁹ Pure culture fermentation of carbohydrates by <i>Pseudomonas elodea</i> , purified by recovery with isopropyl alcohol, dried, and milled ⁸⁶ Gellan gum can be recovered using alcohol precipitation (high acyl gum) or with alkali (deacylated gum) ¹⁴ Produced by an organism that appears to belong to the <i>Auromonas</i> (ATCC 31461) genus; glycerate substitution predominates over acetate ¹² <i>Sphingomonas paucimobilis</i> produce gellan gum ⁸⁶ Production is affected by media components, carbon source, nitrogen source, precursors, agitation rate, pH, temperature, and oxygen ⁸⁶
Welan gum	Fermentative production by <i>Alcaligenes</i> CGMCC2428 ⁷³ Produced by an <i>Alcaligenes</i> species (ATCC 31555) ^{12,88}
Dextran	Dextran 40; dextran 70: obtained by the controlled hydrolysis and fractionation of polysaccharides elaborated by the fermentative action of certain appropriate strains of <i>Leuconostoc mesenteroides</i> on a sucrose substrate ⁹⁰ The fermentation process for reaching high MW dextran takes place at 25°C; at lower temperatures, the amount of low MW dextran increases at >25°C, higher branching occurs ¹¹ Different strains of the same bacterium produce dextrans with differing branched structures ⁷¹ The sucrose concentration also affects branching; increased sucrose content; the degree of branching and the yield of high MW dextran decrease ¹¹ Dextran can be synthesized by dextrinase of different <i>Gluconobacter</i> species ¹¹ Dextran can be produced enzymatically using cell-free culture supernatants that contain dextransucranase ¹¹ Dextran can be synthesized chemically via a cationic ring-opening polymerization of levoglucosan ¹¹
Carboxymethyl dextran	Carboxymethylation of dextran in water/organic solvent mixtures using monochloroacetic acid under strong alkaline conditions ¹¹
Sclerotium gum	Produced by <i>Sclerotium gluconicum</i> ⁸¹ or <i>Sclerotium rolfii</i> ⁹
Beta-glucan	Extraction of extracellular β -glucan produced by the black yeast <i>Aureobasidium pullulans</i> ⁹¹ Produced by fungi, yeasts, and grains (such as oat and barley); beta-glucans present in cereal bran are commonly produced as agricultural by-products ⁷ As curdlan: pure-culture fermentation of a carbohydrate by a nonpathogenic and nontoxic strain of <i>Agrobacterium biovar 1</i> (formerly <i>Alcaligenes faecalis</i> var. <i>myxogenes</i>) or <i>Agrobacterium radiobacter</i> ⁹⁸
Oxidized beta-glucan	Oxidation of beta-glucan is performed using phosphoric acid and sodium nitrite, with the actual oxidant being NO ₂ gas; extent of oxidation was typically 10%-20% ⁹²
Sodium carboxymethyl beta-glucan	Derived from the inner cell walls for baker's yeast (<i>Saccharomyces cerevisiae</i>); 3 of 4 glucose units of the β 1,3-glucopyranose are carboxymethylated ^{80,93} Particulate glucan and sodium hydroxide are mixed, the sodium salt of monochloroacetic acid in 95% ethanol is added and stirred, excess sodium hydroxide is neutralized, the product is washed with 80% ethanol and dried ⁷⁹
Pullulan	Fermentation of liquefied corn starch by <i>Aureobasidium pullulans</i> ; the fungal biomass is removed by microfiltration, the filtrate is heat sterilized, and the pigments and other impurities are removed by adsorption and ion-exchange chromatography ³¹ One company reports the following raw materials: ammonium sulfate; beer yeast extract; calcium hydroxide; caustic soda; corn syrup; diatomaceous earth; diammonium phosphate; dipotassium phosphate; hydrochloric acid; ion-exchange resin; magnesium sulfate; salts; silicone oil; sodium glutamate; zinc carbon chloride ⁹⁴ Produced by <i>Pullularia pullulans</i> IFO 6353, <i>Dermatium pullulans</i> IFO 4464, and so on, in a culture medium containing a carbon source (such as glucose, fructose, etc) under anaerobic conditions ¹⁰⁰
Levan	Produced extracellularly from sucrose- and raffinose-based substrates by levansucrase from a wide range of taxa, such as bacteria, yeasts, and fungi, but mainly by bacteria ⁸ Fermentation of <i>Zymomonas mobilis</i> in a medium that contains 10% sucrose and 1% yeast extract, centrifugation via ultrafiltration, precipitation by the addition of ethanol, resuspension with distilled water, and drying ⁵⁷ Sources include <i>Erwinia herbicola</i> , <i>Aerobacter lavanicum</i> , <i>Streptococcus salivarius</i> , <i>Pseudomonas prunicola</i> , <i>Arthrobacter acetigenum</i> , <i>Bacillus polymyxa</i> , <i>Bacillus subtilis</i> , <i>Actinomyces</i> sp ⁸¹
Rhizobian gum	Produced by fermentation of <i>Rhizobium</i> sp. strain ⁷⁵ Bacterial strain YAS 34 is isolated from the rhizosphere of the sunflower plant; selection of the isolates is carried out on high carbon:nitrogen ratio liquid media; the culture broth was inoculated and fermented; the exopolysaccharide is recovered with a multistep downstream processing that includes heating and centrifugation; diafiltration is used to eliminate fermentation residue, followed by further purification by alcoholic precipitation ¹⁰¹
Alcaligenes polysaccharides	Neutral polysaccharide: culture broth was precipitated with ethanol and redissolved in hot water, filtration was used to remove the water-insoluble cells and acidic polymers, and the polysaccharide was recovered by ethanol precipitation and further purified ⁸³

Table 5. Frequency and Concentration of Use According to Duration and Type of Exposure.

	No. of uses ¹⁵	Maximum concentrations of use (%) ¹⁶	No. of uses ¹⁵	Maximum concentrations of use (%) ¹⁶	No. of uses ¹⁵	Maximum concentrations of use (%) ¹⁶
		Xanthan gum	Dehydroxanthan gum		Xanthan gum crosspolymer	
Totals ^a	3,470	0.00001-6	15	0.1-1	2	0.03-5
Duration of use						
Leave-on	2,782	0.001-6	6	0.1-1	2	0.03-5
Rinse-off	678	0.000001-6	9	0.4-0.8	NR	NR
Diluted for (bath) use	10	0.5-3	NR	NR	NR	NR
Exposure type						
Eye area	292	0.001-2	1	NR	NR	NR
Incidental ingestion	35	0.03-2	NR	NR	NR	NR
Incidental inhalation—spray	121 ^b	0.2-1 ^b ; 0.05 ^c	1 ^b	0.1 ^b -0.2	NR	NR
Incidental inhalation—powder	19	0.3-6	NR	NR	NR	NR
Dermal contact	3179	0.001-6	12	0.1-0.8	2	0.03-5
Deodorant (underarm)	1 ^d	0.005-0.6 ^d ; not a spray: 0.4-1	NR	NR	NR	NR
Hair—noncoloring	129	0.000001-4	3	0.7-1	NR	NR
Hair—coloring	59	0.2-6	NR	NR	NR	NR
Nail	11	0.2-3	NR	NR	NR	NR
Mucous membrane	206	0.03-4	5	0.4	NR	NR
Baby products	29	0.2-0.6	NR	NR	NR	NR
		Gellan gum	Biosaccharide gum-1		Biosaccharide gum-2	
Totals ^a	37	0.0004-0.5	346	0.002-6	14	1
Duration of use						
Leave-on	35	0.0004-0.5	301	0.002-6	10	1
Rinse off	2	NR	43	0.006-5	4	NR
Diluted for (bath) use	NR	NR	2	NR	NR	NR
Exposure type						
Eye area	5	0.0004	28	0.01-1	2	NR
Incidental ingestion	1	0.0004	NR	0.08	NR	NR
Incidental inhalation—spray	NR	NR	3 ^b	0.002 ^b	NR	NR
Incidental Inhalation—powder	6	0.0004	1	NR	NR	NR
Dermal contact	34	0.0004-0.3	326	0.002-6	14	1
Deodorant (underarm)	NR	NR	NR	NR	NR	NR
Hair—noncoloring	1	0.5	19	NR	NR	NR
Hair—coloring	NR	NR	1	NR	NR	NR
Nail	NR	NR	NR	NR	NR	NR
Mucous membrane	2	0.0004	4	0.08	NR	NR
Baby products	NR	NR	NR	NR	NR	NR
		Biosaccharide gum-4	Dextran		Sodium carboxymethyl dextran	
Totals ^a	48	0.00001-5	51	0.000005-0.2	10	NR
Duration of use						
Leave-on	43	0.004-5	48	0.000005-0.1	10	NR
Rinse-off	5	0.00001-0.006	3	NR	NR	NR
Diluted for (bath) use	NR	NR	NR	0.2	NR	NR
Exposure type						
Eye area	7	0.00001-0.2	4	NR	NR	NR
Incidental ingestion	NR	NR	NR	NR	NR	NR
Incidental inhalation—spray	NR	1 ^b	1 ^b	0.01 ^b	NR	NR
Incidental inhalation—powder	NR	NR	NR	NR	NR	NR
Dermal contact	48	0.00001-5	48	0.000005-0.2	10	NR
Deodorant (underarm)	NR	NR	NR	NR	NR	NR
Hair—noncoloring	NR	NR	NR	NR	NR	NR
Hair—coloring	NR	NR	NR	NR	NR	NR
Nail	NR	NR	NR	NR	NR	NR
Mucous membrane	NR	NR	NR	0.2	NR	NR
Baby products	NR	NR	NR	NR	NR	NR

(continued)

Table 5. (continued)

	No. of uses ¹⁵	Maximum concentrations of use (%) ¹⁶	No. of uses ¹⁵	Maximum concentrations of use (%) ¹⁶	No. of uses ¹⁵	Maximum concentrations of use (%) ¹⁶
		Dextran sulfate		Sodium dextran sulfate		Sclerotium gum
Totals	9	0.01-0.1	9	0.005-0.5	193	0.003-2
Duration of use						
Leave-on	9	0.01-0.1	9	0.005-0.5	155	0.003-2
Rinse-off	NR	NR	NR	NR	37	0.003-1
Diluted for (bath) use	NR	NR	NR	NR	1	0.003
Exposure type						
Eye area	9	NR	2	0.01-0.2	25	0.2-0.8
Incidental ingestion	NR	NR	NR	NR	NR	NR
Incidental inhalation—spray	NR	NR	NR	NR	13 ^b	0.003
Incidental inhalation—powder	NR	NR	NR	NR	3	NR
Dermal contact	9	0.01-0.1	9	0.005-0.5	168	0.003-2
Deodorant (underarm)	NR	NR	NR	NR	NR	NR
Hair—noncoloring	NR	NR	NR	NR	20	0.003-1
Hair—coloring	NR	NR	NR	NR	NR	0.8
Nail	NR	NR	NR	NR	1	0.6
Mucous membrane	NR	NR	NR	NR	12	0.003-0.5
Baby products	NR	NR	NR	NR	3	NR
		Hydrolyzed sclerotium gum		Beta-glucan		Sodium carboxymethyl beta-glucan
Totals ^a	NR	1	137	0.000001-0.1	67	0.0001-0.1
Duration of use						
Leave-on	NR	1	103	0.0002-0.1	56	0.0002-0.1
Rinse-off	NR	NR	34	0.000001-0.03	11	0.0001-0.1
Diluted for (bath) use	NR	NR	NR	NR	NR	NR
Exposure type						
Eye area	NR	NR	9	NR	6	0.04
Incidental ingestion	NR	NR	2	NR	NR	NR
Incidental inhalation—spray	NR	NR	NR	NR	NR	NR
Incidental inhalation—powder	NR	NR	7	NR	NR	0.02
Dermal contact	NR	1	128	0.0002-0.1	66	0.0001-0.1
Deodorant (underarm)	NR	NR	NR	NR	NR	NR
Hair—noncoloring	NR	NR	4	0.000001	NR	0.04
Hair—coloring	NR	NR	NR	NR	NR	NR
Nail	NR	NR	NR	NR	NR	NR
Mucous membrane	NR	NR	8	0.03	NR	0.1
Baby products	NR	NR	12	NR	NR	0.02
		Pullulan		Rhizobian gum		Hydrolyzed rhizobian gum
Totals ^a	45 ^e	0.03-17 ^f	5	NR	4	0.4-3
Duration of use						
Leave-on	38	0.2-12	4	NR	3	0.4-3
Rinse-off	1	0.03	1	NR	1	NR
Diluted for (bath) use	NR	NR	NR	NR	NR	NR
Exposure type						
Eye area	10	3	2	NR	2	3
Incidental ingestion	6	17 ^f	NR	NR	NR	NR
Incidental inhalation—spray	1 ^b	NR	NR	NR	NR	NR
Incidental inhalation—powder	NR	NR	NR	NR	NR	NR
Dermal contact	36	0.2-3	5	NR	4	0.4-3
Deodorant (underarm)	NR	NR	NR	NR	NR	NR
Hair—noncoloring	NR	12	NR	NR	NR	NR
Hair—coloring	NR	0.03	NR	NR	NR	NR
Nail	NR	NR	NR	NR	NR	NR
Mucous membrane	6	17 ^e	NR	NR	NR	NR
Baby products	NR	NR	NR	NR	NR	NR

(continued)

Male Sprague Dawley rats were also given a single IV dose of 1, 25, or 100 mg/kg FD-4 (average MW 4,300) and FD-150 (average MW 145,000).⁴⁸ The dextran solution was prepared as isotonic phosphate buffer at a concentration that would result in a test volume of 2 mL/kg. Blood, urine, and tissue samples were taken at various intervals for up to 6 hours from groups of 4 rats dosed FD-4 and for up to 96 hours in rats dosed with FD-150. Renal excretion was a major excretion pathway for FD-4 but not FD-150. Urinary recovery ranged from 79% to 82% of the dose with FD-4 and only from 1.1% to 2.1% of the dose with FD-150; at each MW, the dose administered did not have a statistically significant effect on the amount excreted. Renal clearance ranged from 344 to 360 mL/h/kg with FD-4 and from 0.131 to 0.245 mL/h/kg for FD-150, and systemic clearance ranged from 420 to 457 mL/h/kg for FD-4 and from 8 to 20 mL/k/kg for FD-150. The highest concentrations of FD-4 were found in the kidneys at 1 minute after dosing (9.31%, 10.0%, and 10.4% of the dose with 1, 25, and 100 mg/kg, respectively) was linear with dose. The highest concentrations of FD-150 were found in the liver (68.5% at 5 hours, 51.6% at 24 hours, and 41.5% of the dose at 24 hours with 1, 25, and 100 mg/kg, respectively) and the spleen (11.5% at 12 hours, 2.09% at 48 hours, and 1.21% of the dose at 96 hours with 1, 25, and 100 mg/kg, respectively); recovery of dextran in the liver and spleen was nonlinear, with a greater difference seen in the spleen than in the liver. The researchers reported that the MW of recovered FD-4 remained relatively constant but that of recovered FD-150 changed significantly. The MW of FD-150 recovered in the urine was <40,000, and in the liver, the average MW at 96 hours was ~70,000. The estimated MW of the recovered FD-150 in the liver appeared to be dose dependent, with higher doses having a higher average MW. The researchers concluded that excretion of lower MW dextrans was independent of dose, whereas excretion of higher MW dextrans was dose dependent.

Another study also found that MW affected the distribution and excretion of dextran.⁴⁹ Female BALB/cCrSlc mice were dosed IV with 0.1 mL of 0.1 wt% ¹²⁵I-labeled dextran, MW ranging from 4,980 to 220,000, in phosphate-buffered saline (PBS); dextran was labeled through radioiodination of tyramine residues. Blood samples were taken at various intervals for some groups, and tissues were collected from others (the number of animals/group was not specified). High MW dextran remained in the blood longer than lower MW dextran. The $t_{1/2\beta}$ also increased with increasing MW, with a pronounced change occurring around 30,000 MW. After 3 hours, 83% of the dose of the lowest MW dextran was excreted, whereas only 41% of the highest MW dextran was found in excrement. Most of the high MW dextran was found in the liver; after 3 hours, 5.2% of the high MW dextran was found in the liver, whereas only 0.7% of the lowest MW was recovered in that tissue. At 3 hours, 3.5% versus 19% of the dose of the lowest and highest MW dextran, respectively, was recovered in the carcass. Two percent to 10% of the dose was recovered in the GI tract, but the amount recovered did not appear to be related to MW.

Dextran sulfate. A preliminary study was performed in which 2 male Wistar rats were dosed by IV injection with 20 mg/kg dextran sulfate 8,000 average MW containing 10 μ Ci [³H]dextran sulfate/kg via the penile vein (5 mg/100 μ L).⁴¹ The rats were killed 1 or 3 hours after dosing. The total ³H excreted in the urine accounted for approximately 50% of the administered dose. Within 1 hour after IV administration, rapid excretion of intact dextran sulfate occurred.

In the main study, groups of 5 male Wistar rats were dosed by IV injection via the penile vein with 5 mg/100 μ L of 20 mg/kg dextran sulfate containing 12 μ Ci [³H]dextran sulfate/kg, and each group was killed 3 or 24 hours after dosing. Urine was the major route of excretion following IV dosing, with approximately 46% of the dose excreted within 3 hours and 51% within 24 hours. Approximately 2% of the dose was recovered in the feces after 24 hours. Based on the ³H elution profiles, it was hypothesized that dextran sulfate or its metabolites were incorporated into higher MW compounds, such as glycogen, at 3 to 6 hours and smaller MW at 6 to 24 hours. In the plasma, the highest concentration of dextran sulfate was found at 3 hours; the amount decreased with time.

In the tissues, the highest amounts of radioactivity were distributed in the liver, kidney, and spleen. The amount of radioactivity recovered in the tissues following IV administration was compared to that found following oral administration (the oral study was described previously in this safety assessment). Concentrations of ³H in all tissues and fluids were statistically significantly higher in the animals dosed by IV injection compared to those in animals dosed orally.

Beta-glucan. Groups of 2 male Sprague Dawley rats were dosed by intraperitoneal (IP) injections with 20 mg/kg BW [¹⁴C]beta-glucan (as curdlan) in water, and the animals were killed 0.5, 3, 6, or 24 hours after dosing.⁴² After 24 and 48 hours, only 1.8% and 4.1% of the radioactivity was recovered in CO₂, respectively, 0.05% and 0.12% was recovered in the feces, respectively, and 3.5% and 4.1% of the radioactivity as percentage of dose was recovered in the urine, respectively. Whole-body radioautography showed that the radiolabel was distributed in the intestinal fluids.

Pullulan. The effect of MW on the distribution and excretion of pullulan was examined.⁴⁹ Female BALB/cCrSlc mice were dosed IV with 0.1 mL of 0.1 wt% [¹²⁵I]pullulan, MW ranging from 5,800 to 853,000, in PBS; pullulan was labeled through radioiodination of tyramine residues. Blood samples were taken at various intervals for some groups, and tissues were collected from others (the number of animals/group was not specified). High MW pullulan remained in the blood longer than lower MW pullulan. The $t_{1/2\beta}$ also increased with increasing MW. After 3 hours, 96% of the dose of the lowest MW pullulan was excreted, whereas only 15% of the highest MW pullulan was found in excrement. At 3 hours, most of the high MW pullulan, 50% of the dose, was recovered in the liver; only 1% of the dose of the lowest MW pullulan was recovered in the liver after 3 hours. The highest percentage of the dose recovered in the GI tract was 5.3% pullulan with a MW of

100,000 MW, and in the carcass, it was 10.1% pullulan with a MW of 48,000.

Fasted male Wistar rats (number not specified) were injected with a single IV dose 2 mL of 0, 6, 12, 18, or 24 mg/kg fluorescein-labeled pullulan (MW 58,200) in saline in the jugular vein.⁵⁰ Pullulan was rapidly eliminated from the blood; however, elimination decreased as dose increased. Hepatic uptake was also dose dependent; the hepatic uptake clearance of pullulan decreased with increased dose. In the liver, distribution of pullulan in the parenchymal cells was 2.5 times greater than in the nonparenchymal cells. The researchers did state that with a higher MW pullulan, average 70,000 MW, uptake was greater in the nonparenchymal cells.

Human

Dextran. Four male and 3 female subjects received an IV injection 100 mL, a partially degraded dextran (40,000 average MW; 10% wt/vol in 0.9% saline).⁵¹ Blood samples were taken at various intervals after dosing. There was an initial rapid decrease in the serum in the first hour after dosing. Different fractions of the dextran were eliminated from the plasma at different rates; higher MW fractions remained in the plasma longer.

Toxicological Studies

Single-Dose (Acute) Toxicity

Acute toxicity studies are summarized in Table 8. The acute toxicity of xanthan gum, gellan gum, beta-glucan, sodium carboxymethyl beta-glucan, and pullulan was assessed orally in mice, rats, and/or dogs, and dextran sulfate and beta-glucan were tested by IP and IV dosing in mice and rats. There was no notable toxicity observed in these studies. In acute inhalation studies, the LC₅₀ of xanthan gum was >21 mg/L in rabbits and of gellan gum was >5.06 mg/L in rats. A single 180-minute exposure of humans office dust containing 10 mg curdlan/g dust resulted in decreased nasal volume, swelling in the nasal turbinates, and an increase in nasal eosinophils when compared to "clean" dust.

Inflammatory response

Beta-glucan. The inflammatory response following a single exposure to beta-glucan (as curdlan) was evaluated in guinea pigs.⁵²⁻⁵⁴ Mostly, no effect or a slight decrease in inflammatory cells in lung lavage was observed. A 4-hour inhalation exposure to 1% beta-glucan in dust (mass mean aerodynamic diameter [MMAD] 5 µm) by guinea pigs produced a delayed subacute nasal congestion when compared to dust without beta-glucan (MMAD 6.5 µm) and resulted in decreased nasal volume; after 18 hours, there was a significant decrease in nasal volume.⁵² In humans, inhalation exposure to beta-glucan in dust for four 3-hour exposures resulted in increased nasal swelling and decreased nasal volume and an immediate increase in nasal eosinophil/mL count.⁵⁵

Pullulan. The effect of pullulan on inflammation was investigated in ICR mice using the xylene-induced acute inflammatory mouse ear model.⁵⁶ Groups of 9 mice were dosed orally with 0, 62.5, 125, or 250 mg/kg pullulan in distilled water 30 minutes prior to topical application of xylene to 1 ear. Two hours after xylene application, all animals were killed. Compared to xylene-treated controls, ear weights were statistically significantly decreased in a dose-dependent manner. Additional histological indicators of inflammation were not observed.

Levan. An interleukin (IL) 1α release assay was used to determine the anti-inflammation effect of a 5% aq. levan solution on artificial skin.⁵⁷ Primary skin irritation was first induced using sodium lauryl sulfate. A dose of 0.01 or 0.05 mg/mL of the solution was applied to the skin. Levan decreased IL-1α release, indicating an anti-inflammatory effect.

Cytotoxicity

Levan. The cytotoxic effect of 5% levan (wt/wt) was determined using human fibroblasts and keratinocytes.⁵⁷ The 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide assay was used to measure cell viability and proliferation after 24-hour incubation with levan. Levan, ≤100 µg/mL, was not cytotoxic to human fibroblasts. Levan had a proliferative effect in keratinocytes; proliferation was >30% at concentrations of >1 mg/ml.

Repeated Dose Toxicity

Repeated dose toxicity studies are summarized in Table 9. The oral toxicity of xanthan gum was evaluated in rats and dogs, of gellan gum in rats, dogs, and monkeys, of dextran in rats, of beta-glucan in mice, rats, and dogs, and of pullulan in rats. Most of the studies were dietary, and study durations lasted up to 2 years. Most observations were related to changes in feed consumption and intestinal effects. In guinea pigs, inhalation exposure to 100 µg/mL beta-glucan (as curdlan), 4 h/d, 5 d/wk for 4 weeks, did not have an effect on the cells of the lung lavage or cell wall, and there were no microscopic lesions of the lung. With IP administration, no toxicity was reported when mice were dose 10 times over 2 weeks with 5 mg xanthan gum in 0.5 mL water or mice or guinea pigs were dosed with 250 mg/kg BW beta-glucan for 7 days. Intravenous administration of 40 and 1,000 mg/kg BW beta-glucan for 30 days resulted in hepatosplenomegaly in mice.

Oral intake by humans

Xanthan gum. Five male subjects consumed 150 mg/kg BW/d xanthan gum as 3 measured portions daily for 23 days.⁵⁸ Ingestion of xanthan gum had no significant adverse effect on hematology, clinical chemistry, or urinalysis parameters.

Gellan gum. Five male and 5 female subjects consumed a daily dose of 175 mg/kg gellan gum for 7 days and 200 mg/kg/d for the next 16 days.⁵⁸ No adverse dietary or physiological effects and no allergenic effects were reported.

Table 8. Acute Toxicity Studies.

Ingredient	Animals	No./group	Vehicle	Concentration/dose/protocol	LD ₅₀ /results	Reference
Oral						
Xanthan gum	Mice	Not specified	Water	Not specified	>1 g/kg	104
Xanthan gum	Rats	Not specified	Not specified	5 g/kg, max	>5 g/kg	105
Xanthan gum	Dogs	Not specified	Not specified	20 g/kg, max	>20 g/kg	105
Gellan gum	Rats, M/F	Not specified	Not specified	Administered in diet and by gavage; nonacetylated; >95% polysaccharide	>5 g/kg	39
Beta-glucan (as curdlan)	Mice, M/F	Not specified	Water	Tested as a 10% suspension	>10 g/kg	42
Beta-glucan (as curdlan)	Rats, M	Not specified	Water	Tested as a 10% suspension	>10 g/kg	42
Beta-glucan (highly pure extract of <i>S cerevisiae</i>)	Rats	5 M/5 F	Water	100 mg/mL suspension administered at 2 g/kg BW (20 mL/kg BW) by gavage	>2 g/kg	106
Sodium carboxymethyl beta-glucan (>90% pure)	DDY mice	6 M/6 F	Purified water	2 g/kg of a 20 aq. solution	>2 g/kg; no signs of toxicity	107
Pullulan	Mice	Not specified	Olive oil	Not specified	>14 g/kg	108
Inhalation						
Xanthan gum	Albino rabbits	5	None	Calculated chamber concentration of 21 mg/L; 1-hour exposure	>21 mg/L; no toxicity; no gross lesions at necropsy	38
Gellan gum	Rats, M/F	Not specified	Not specified	Nonacetylated; >95% polysaccharide; duration of exposure not stated	>5.09 mg/L	39
Beta-glucan (as curdlan)	Guinea pigs	5	Not specified	100 µg/mL curdlan for 4 hours; continuous flow aerosol	No inflammatory cell response	53
Beta-glucan (as curdlan)	Guinea pigs	3	NaOH	1 µg/mL; continuous flow for 4 hours	Decrease in inflammatory cells with curdlan alone, increase in cells, especially neutrophils with NaOH	54
Beta-glucan (as curdlan)	Guinea pigs	5	Not specified	6 µg/mL for 40 minutes, and animals were examined 4 or 24 hours after exposure or 1 µg/mL for 4 hours, and animals were examined 5 or 9 days after exposure	In lung lavage: decrease in lymphocytes at 24 hours; in lung wall: statistically significant decrease in macrophages after 5 days; decrease in lymphocytes for 1 to 7 days	54
Beta-glucan (as curdlan)	Guinea pigs	5	Not specified	100 µg/mL; continuous flow for 40 minutes or 4 hours	In lung lavage: slight decrease in macrophages; (almost significant) decrease lymphocytes	54
Beta-glucan (as curdlan); MMAD 5 µm	Guinea pigs	Not specified	Dust	Office dust spiked with 1% beta-glucan (as curdlan; 280 g office dust was spiked with 2.8 g curdlan) for 4 hours in a whole-body exposure chamber; necropsied 5 or 18 hours after exposure	Glucan-spiked dust produced a delayed subacute nasal congestion in guinea pigs compared to dust without beta-glucan; at 18 hours after exposure, there was a significant decrease in nasal volume	52
Beta-glucan (as curdlan)	Humans	36	Dust	Addition of 10 mg beta-glucan (as curdlan) to office dust (10 mg curdlan/g dust); subjects were exposed to the dust in a climate chamber for 180 minutes	Compared to "clean" dust, nasal volume decreased, swelling in the nasal turbinate increased, and nasal eosinophil cell concentration increased	55
Pullulan	Guinea pigs	5	Not specified	100 µg/mL curdlan for 4 hours; continuous flow aerosol	No inflammatory cell response	53
Parenteral						
Dextran sulfate, average MW of 7,500, 47,000, and 458,000	Mice and rats	Not specified	Not specified	0.1-2 g/kg, IV and IP	"Some animals" died immediately; the largest MW dextran was more toxic	109
Beta-glucan (as curdlan)	Mice, M/F	Not specified	Saline	5% suspension, IP	Males: 2.75 g/kg; females: 2.5 g/kg	42
Beta-glucan (as curdlan)	Rats, M	Not specified	Saline	5% suspension, IP	2.75 g/kg; adhesions of beta-glucan to liver and spleen	42
Beta-glucan (soluble; extract of <i>S cerevisiae</i>)	ICR/HSD mice	Not specified	Not specified	≤1 g/kg, IV	>1 g/kg	110
Beta-glucan (soluble; extract of <i>S cerevisiae</i>)	Sprague Dawley rats	Not specified	Not specified	0.5 g/kg, IV	>0.5 g/kg	110

Abbreviations: Aq., aqueous; F, female; IP, intraperitoneal; IV, intravenous; M, male; Max, maximum; MMAD, mass mean aerodynamic diameter; MW, molecular weight.

Table 9. Repeated Dose Toxicity Studies.

Ingredient	Animals/group	Study duration	Vehicle	Dose/concentration	Results	Reference
Oral						
Xanthan gum	Rats (#/group not stated)	18 days	In diet	7.5%	Paired feeding study—wt gains were similar to controls	104
Xanthan gum	25 rats	30 days	In diet	1.5%	Average wt of test animals was greater than controls; no gross lesions	58
Xanthan gum	Albino rats, 5 M	91 days	In diet	3%, 6%, or 15%	Reduced weight gain in the 15% group; no effect on organ wts; no microscopic lesions at necropsy in the 15% group	104
Xanthan gum	Rats, 5 M/5 F	110 days	In diet	0%, 2.5%, 5.0%, or 10%	The gum product consisted of drum-dried whole fermentation medium (beer); no significant toxic effects	38
Xanthan gum	CD rats, 30 M/30 F	2 years	In diet	0, 0.25, 0.50, or 1.0 g/kg BW/d	No significant differences in growth rate, survival, hematology and clinical chemistry parameters, or organ weight were observed between treated and control animals; although not statistically significant (stat. sig.), there was an increase in uterine polyps in the high-dose groups compared to controls, 5 in the high-dose animals vs 2 in controls	61
Xanthan gum	Beagle dogs, 3 M/3 F	12 weeks	In diet	0, 0.25, or 0.5 g/kg BW/d	Growth of high-dose males was slightly less than controls; the no-observable adverse effect level (NOAEL) was 0.25 g/kg BW/d	38
Xanthan gum	Beagle dogs, 2 M/2 F	12 weeks	In diet	0, 1, or 2 g/kg BW/d; positive controls were given 20 g/kg BW/d powdered cellulose	Immediate and persistent diarrhea in the 2 g/kg group; body wt loss was observed in treated and control animals, with the wt loss greatest in the 2 g/kg group; red blood cell counts, hemoglobin, and serum cholesterol levels were decreased in high-dose animals; adrenal glands were slightly enlarged in the 2 g/kg group; no treatment-related microscopic lesion were observed at necropsy	111
Xanthan gum	Beagle dogs, 4 M/4 F	107 weeks	In diet	0, 0.25, 0.37, and 1.0 g/kg/d	No significant differences in survival, BW gain, feed consumption, organ weights, hematology parameters, or gross or microscopic lesions were observed between treated and control animals; a dose-related increased in urinary specific gravity was observed; ophthalmic findings were not considered treatment related	61
Gellan gum	20 rats	28 days	In diet	5%	No changes in urinalysis values; no gross lesions at necropsy	112
Gellan gum; nonacetylated; >95% polysac.	Sprague Dawley rats, 20 M/20 F	13 weeks	In diet	0%-6%	No mortality; no signs of toxicity	39
Gellan gum; varied degree of acetylation; 58.5% polysac.	Beagle dogs, 5 M/5 F	52 weeks	In diet	0%, 3%, 4.5%, or 6%	No mortality; feed consumption was greater in treated animals compared to controls; no adverse effects were observed	39
Gellan gum; varied degree of acetylation; 58.5% polysac.	Rhesus monkey, 2 M/2 F	28 days	Not provided	0, 1, 2, or 3 g/kg, by gavage	No signs of toxicity	39
Dextran Beta-glucan (as curdlan)	Albino rats, 6 M CD-1 mice, 10 M/10 F	62 days 8 weeks	In diet In diet	15% 0%, 1%, 5%, 10%, 20%, or 30%; equivalent to 0, 1.4, 7.1, 14, 29, and 43 g/kg B, respectively	Wt gain was normal One female of the 30% group died; body wt gains of male mice of the 30% group were decreased compared to controls; no gross abnormalities; differences in stools and cecal wts were reported; the NOEL was 5% based on an increase in full cecal wts and large stools at higher doses	104 42

(continued)

Table 9. (continued)

Ingredient	Animals/group	Study duration	Vehicle	Dose/concentration	Results	Reference
Beta-glucan (as curdlan)	CD-1 mice, 100 M/100 F	99-114 weeks (until survival was 20%)	In diet	0%, 1%, 5%, or 15%	No treatment-related differences in survival or body wts; feed consumption was decreased by ~ 13% in 15% females; NOEL of 5% due to decreased feed consumption	42
Beta-glucan (as curdlan)	Sprague Dawley Ta rats; 5 M	4 weeks	In diet	0%, 3%, 10%, or 30%; equivalent to 0, 2.5, 8.5, or 30 g/kg BWV; 30% powder agar group	No difference in body wts; dose-related increase in neutrophils, with a stat. sig. increase in the 10% group; occult blood was present in the urine four 10% rats, 1 30% rat, and 1 fed agar; relative (rel.) kidney wts were statistically significantly decreased in all groups; rel. liver weights were statistically significantly decreased and pituitary wts were stat. sig. increased in the 30% group	42
Beta-glucan (as curdlan)	20 Sprague Dawley rats, males	8 weeks	In diet	0%, 1%, 5%, or 15%	No mortality or signs of toxicity; body wt of animals of the 15% dose group were stat. sig. decreased; feed consumption in this group was slightly decreased	42
Beta-glucan (as curdlan)	Sprague Dawley rats, 10 M/10 F	3 months	In diet	0%, 5%, 10%, or 20%; equivalent to 0, 4.4, 9, and 19 g/kg BWV (males) and 0, 5.5, 12, and 24 g/kg BWV (females)	Body wt gains decreased with increasing dose, being stat. sig. in the 20% group; a stat. sig. decrease in platelet count in males and in total protein and globulin concentrations in males and females of the 10% and 20% groups; stat. sig. increases in body wt of males of the 10% and 20% group; stat. sig. decreases in absolute liver wts in 20% males, absolute (abs.) kidney wts of 10 and 20% males, abs. and rel. ovary wts in 20% females, abs. and rel. pituitary wts of in all females of all doses; and rel. adrenal wts in 10% and 20% males; differences in stools and cecal wts were reported; the NOEL was 5% based on fecal changes, diarrhea, and cecal wts and large stools at higher doses	42
Beta-glucan (as mushroom beta-glucan from <i>Ganoderma lucidum</i>)	CD (SD) IGS rats, 12 M, 12 F	3 months	Sterile water; by gavage	0, 500, 1,000, or 2,000 mg/kg BW (10 mL/kg dosing volume)	No test article-related adverse effects on mortality, toxicity; ophthalmoscopy, body wts or body wt gains, clinical chemistry, hematology, or urinalysis; statistically significant decreases in testes wt in males of the low-dose group and heart wts in females of the high-dose group were not considered test-article related; there were no treatment-related gross or microscopic lesions, the NOEL was 2,000 mg/kg BW	42
Beta-glucan (as curdlan)	CD rats; 60 M/60 F	2 years	In diet	0%, 1%, 5%, or 15%	No changes in mortality, behavior, appearance, or ophthalmic parameters; wt gain was decreased by ~ 10%, which was not stat. sig., in the 15% group; feed consumption was also reduced; no microscopic lesions were found, the NOEL was 5% based on decreased feed consumption, body wt gain, and increased cecal wts	42
Beta-glucan (as curdlan)	CD rats; 60 M/60 F, of the F ₁₃ generation of a reproductive toxicology study	124-127 weeks (20% survival)	In diet	0%, 1%, 5%, or 15%	No changes in mortality, behavior, appearance, or ophthalmic parameters; body wts of the 15% group were stat. sig. decreased, and body wts were stat. sig. decreased in 5% males until wk 65; feed consumption was decreased in the 15% group; no changes in hematology or urinary parameters, but some stat. sig. changes were reported in blood chemistry; stat. sig. increase in gross and microscopic incidences of uterine polyps in the 15% groups—the incidences were 0/450, 3/50, 4/51, and 7/50 for the 0%, 1%, 5%, and 15% groups, indicating that the polyps were possible treatment related; the NOEL was 1% based on increased cecal wts, decreased body wts and feed consumption, and the incidence of polyps	42

(continued)

Table 9. (continued)

Ingredient	Animals/group	Study duration	Vehicle	Dose/concentration	Results	Reference
Beta-glucan (as curdlan)	Beagle dogs, 4 M/4 F	52 weeks	In diet	0%, 1%, 5%, or 15%	One 15% male died at 37 weeks (not dose related); no stat. sig. treatment-related changes, except for fecal changes and cecal wts; NOEL was 5% based on fecal changes and cecal wts	42
Beta-glucan (highly pure extract of <i>S cerevisiae</i>)	SPF Fischer rats, 10 M/10 F	91 days	Water	0, 2, 33.3, or 100 mg/kg BW / d, volume 0.5 mL/100 g BW, by gavage	No mortality; no stat. sig. differences in wt gains, feed consumption, or gross or microscopic lesions; a dose-dependent and stat. sig. increase in clotting time in males, isolated stat. sig. changes in some clinical chemistry parameters, and slight but stat. sig. incr. in absolute liver, kidney, heart, spleen, adrenal, and testicle wts in males and absolute kidney and thymus wts in females were not considered toxicologically significant; NOAEL was 100 mg/kg BW/d	106
Beta-glucan (high-purity extract barley beta-glucan)	SPF Wistar rats, 5 M/5 F	28 days	In diet	0%, 1%, 5%, or 10%, supplemented with \leq 10% potato starch	No mortality; no sig. effects on body wts, feed consumption, or functional observational battery results; no treatment-related change in hematology or urinalysis values; empty caecum wt was increased	113
Beta-glucan (extracted from <i>Candida albicans</i>)	Sprague-Dawley rats, 20 M/20 F	52 weeks	Sterile saline (using a rubber catheter)	0, 50, 100, or 200 mg/kg/d	No sig. effects on mortality, body wts, feed consumption, or hematology, clinical chemistry, or urinalysis parameters; with the exception of cecal enlargement with variable hyperplasia, not gross or microscopic lesions were noted; the NOEL was 100 mg/kg/d	114
Pullulan	8 Wistar rats, males (test and cellulose control groups)	4 or 9 weeks	In diet	0%, 5%, 10%, 20%, or 40%; equivalent to 0, 2,500, 5,000, 10,000, or 20,000 mg/kg, respectively; cellulose controls: 20% or 40% cellulose	Body wt gains were decreased by day 10 in rats of the 20% and 40% groups when compared to untreated controls; wt gains of animals of the 5% and 10% pullulan group were lower than untreated controls after 7 weeks, but this difference was not stat. sig.; similar decreases were observed in the animals fed cellulose; diarrhea was observed with 40% pullulan; rel. wts of the stomach, small intestine, and large intestine were increased in treated animals	43,44
Pullulan (200,000 MVV)	SPF Wistar rats, 10 M/10 F	13 weeks	In diet	0%, 2.5%, 5%, or 10%; equivalent to 0, 1,960, 4,100, or 7,900 mg/kg BW/d; diet of the groups fed 0%, 2.5%, or 5% pullulan was supplemented with 10%, 7.5%, or 5% potato starch, respectively	No mortality; no dose-related clinical sign; stat. sig. reduced motor activity in females of the 5% and 10% groups was observed and appeared treatment related, but as a physiological phenomenon rather than a toxic effect; no difference in hematology parameters between treated and control groups; differences that were observed in clinical chem. parameters were not biologically significant or dose related; dose dependent, stat. sig. increases in abs. and rel. empty cecal wts were observed in males of the 5% and males and females of the 10% group; no microscopic changes were observed	44
Pullulan	Sprague-Dawley rats, 15 M/15 F	62 weeks (was to be 24 months; study was terminated because of poor survival due to pneumonia)	In diet	0%, 1%, 5%, and 10%	No treatment-related effects on body wts, feed consumption, or organ wts (except for cecal wts); changes in hematology or clinical chemistry parameters and microscopic lesions that were observed were not considered treatment related; the NOAEL was 10% in the diet (equivalent to 4,450 mg/kg BW /d)	115

(continued)

Table 9. (continued)

Ingredient	Animals/group	Study duration	Vehicle	Dose/concentration	Results	Reference
Inhalation						
Beta-glucan (as curdlan)	Guinea pigs, 16 F	5 weeks	Distilled water	100 µg/mL; 4 h/d, 5 d/wk	No significant change in lung lavage cells, but there was a decrease in the number of lymphocytes; slight but not statistically significant increase in macrophages and eosinophils in the lung wall cells; no lesions observed at microscopic examination of the lungs	116
Beta-glucan (as curdlan)	Guinea pigs, 6	5 weeks	Saline	100 µg/mL; 4 h/d, 5 d/wk; continuous flow exposure with a dose of 8 pg; animals were examined 24 hours after last dose	The only effect on lung lavage cells was a slight decrease in neutrophils; there was no effect on the number of lung wall cells	117
Parenteral						
Xanthan gum	Mice	2 weeks	Water	5 mg in 0.5 mL water; 10 IP injections over 2 weeks	No toxicity; there was no xanthan gum in the abdominal cavity at necropsy	104
Dextran, partly hydrolyzed, bacterial, 75,000 average MW	16	103-113 weeks	Physiological saline	30 mL of 6% solution, IV	4 animals died and 1 was killed in moribund condition; wt gain was similar to that of controls; increases in absolute heart and adrenal wts were probably statistically significant; statistically significant increase in liver and spleen wts; no microscopic lesions	47
Beta-glucan (soluble; extract of <i>S cerevisiae</i>)	Mice	7 days	Not specified	250 mg/kg BW, IP	No effect on weight gain	110
Beta-glucan (soluble; extract of <i>S cerevisiae</i>)	Guinea pigs	7 days	Not specified	250 mg/kg BW, IP	Sig. 10% decrease in weight gain	110
Beta-glucan (soluble; extract of <i>S cerevisiae</i>)	ICH/HSD mice, M	30 days	Not specified	≤ 1,000 mg/kg BW, IV	Sig. toxicity leading to hepatosplenomegaly in the 40 and 1,000 mg/kg BW groups	110
Beta-glucan (soluble; extract of <i>S cerevisiae</i>)	ICH/HSD mice, M	60 days	Not specified	≤ 1,000 mg/kg BW, IV	Dose-dependent increase in hepatosplenomegaly; was stat. sig. at 1,000 mg/kg	110

Abbreviations: F, female; IP, intraperitoneal; IV, intravenous; M, male; polysac, polysaccharide.

Dextran and pullulan. No adverse effects were seen in a study in which 10 g pullulan, dextran, and soluble starch were ingested by 8 male volunteers.^{42,59} Each ingredient was administered for 14 days, and there was a 14-day washout period between treatments.

Beta-glucan. Six male subjects ingested milkshakes with or without beta-glucan (as curdlan) for 28 days.⁴² The test subjects were given 6 g/d for 5 days, 35 g/d for 2 weeks, and 50 g/d from days 21 to 28. No evidence of toxicity was observed.

Pullulan. In a tolerance study, 13 subjects ingested 10 g/d of pullulan (50,000 MW) for 14 days.^{44,46} There were no effects on clinical chemistry parameters, and no adverse effects were reported.

Industrial exposure

Xanthan gum. The relationship between the handling of xanthan gum powder and adverse symptoms was examined using exposure groups based on average percentage of time spent in a plant that uses a fermentation process to manufacture xanthan gum, not on the basis of expected intensity of exposure to xanthan gum.⁶⁰ The exposure of interest was to the employees exposed to milling, blending, and packaging of the product; a total of 39 employees were surveyed. Analysis of the results found no significant acute or chronic effects in pulmonary function in any of the exposure groups.

Reproductive and Developmental Toxicity

Oral

Xanthan gum. A 3-generation reproductive toxicity study was performed in which albino rats were fed dietary levels of 0, 0.25, and 0.5 g/kg BW/d xanthan gum.⁶¹ Ten males and 20 females were used for the first generation, and 20 males and 20 females were used in the next 2 generations. Rats were mated to produce 2 litters per generation, and the successive generations were selected from weanlings of the second litter. Survival and reproductive parameters were similar for treated and control parental rats. Body weights of treated parental rats were slightly decreased compared to controls in each generation. There were no significant differences in developmental parameters between test and control litters, and no malformations were observed in any of the offspring.

Gellan gum. Groups of 25 gravid female Sprague Dawley rats were fed a diet containing 0%, 2.5%, 3.8%, or 5.0% gellan gum (varied degree of acetylation; 58.5% polysaccharide) on days 6 to 15 of gestation.³⁹ No fetotoxic or teratogenic effects were reported (no other details were provided).

Beta-glucan. A developmental study was performed with 0%, 5%, or 15% beta-glucan (as curdlan) with a control group of 40 male and 80 female CD rats and test groups of 20 male and 40 female rats.⁴² The animals, which were mated twice, were fed the test diet throughout the study. Twenty of the treated dams nursed their own litters; the other 20 treated dams switched

litters with the control dams so that treated animals would nurse control pups and control animals would nurse test pups. The F_{1a} offspring were killed prior to the second mating.

No changes in mortality, behavior, or appearance were observed. Male sires of the 15% beta-glucan group had decreased growth rates compared to controls, and males and females of the 15% group had decreased feed consumption. At birth, there were no differences in fertility or lactation among the groups, and no abnormalities were reported. However, survival of the F_{1a}, but not the F_{1b}, pups of the 5% group was statistically significantly decreased compared to controls. Weight gain of all F_{1a} litters of treated dams that nursed their own pups was statistically significantly decreased compared to controls; for the F_{1b} litters, the difference was statistically significant only in the 15% group. Statistically significant decreases in weight gain were also observed for pups of treated dams that were nursed by control dams, but the effect was reduced. Statistically significant decreased weight gain at some intervals was also observed for control pups nursed by treated dams. A no observable effect level (NOEL) was not established.

The researchers also examined whether there would be decreased weight gain by the pups if dosing was discontinued during lactation. The protocol was similar to that just described, except that all groups consisted of 20 male and 40 female CD rats, and there was no crossover at lactation. Weight gain by all pups during lactations was similar, although the researchers did state that the pups could have consumed parental diet from day 10+. The NOEL for parental toxicity and embryotoxicity was 15% beta-glucan.

A 3-generation reproductive study was performed in which groups of 20 male and 40 female CD rats were fed a diet containing 0%, 1%, 5%, or 15% beta-glucan (as curdlan) for 100 days.⁴² The F₀ parents were mated twice, and the number of parents was halved after weaning of the first litter. The F₁ parents were mated 3 times and the F₂ parents were mated twice. The F_{1b} and F_{2b} litters were used to produce the next generation. After the third mating of the F₁ parents, half of the F₁ dams were killed on day 13 of gestation, and the remaining dams were killed on day 20 of gestation.

Mean growth and feed consumption were slightly decreased in male parental rats of the F₀ and F₁ generations of the 15% group. No gross or microscopic changes were observed in F₂ parents. No treatment-related effects on reproductive and developmental parameters were observed, but BWs of pups in almost all litters in all generations were statistically significantly decreased during lactation in the 15% group. Biologically significant differences in BWs were not seen in litters of the other dose groups. No gross or microscopic lesions were observed in the F_{3b} pups of the 15% group. In the F₁ parents killed after the third mating, no reproductive or developmental effects were observed. Mean fetal weights in all groups were statistically significantly decreased compared to controls; however, there was no dose response. The NOEL for parental animals was 5%, based on decreased growth and increased cecal weights at 15% beta-glucan, and the NOEL for embryotoxicity

was also 5%, based on decreased weight gain during lactation in the 15% beta-glucan group.

The teratogenic potential of beta-glucan (as curdlan) was determined using groups of 15 to 20 gravid Dutch-belted rabbits.⁴² The rabbits were dosed orally with 0, 1, 2, or 5 g/kg BW/d beta-glucan in a gelatin capsule delivered using a syringe. The 5 g/kg dose was administered as 2 divided doses, and the controls received 2 empty capsules. The rabbits were killed on day 28 of gestation. None of the controls died, but 1, 1, and 3 dams of the 1, 2, and 5 g/kg BW/day groups, respectively, died during the study. Eleven resorptions were observed in the high-dose group, as compared to 4 in the control group, 6 in the 1 g/kg group, and 5 in the 2 g/kg group. The researchers stated, however, that the number of dams with resorptions was similar in all groups and that no teratogenic effects were observed. The NOEL for both maternal and embryotoxicity was 5 g/kg BW/d.

Genotoxicity

Genotoxicity studies are summarized in Table 10. The in vitro genotoxicity of gellan gum (≤ 20 mg/mL), sodium dextran sulfate (≤ 25 mg/plate), beta-glucan ($\leq 5,000$ μ g/plate or μ g/mL), sodium carboxymethyl beta-glucan ($\leq 50,000$ μ g/plate or μ g/mL), and pullulan (≤ 12 mg/mL) was evaluated using Ames test, chromosomal aberration assays, and/or DNA repair tests, with and without metabolic activation. The results were negative in these tests. The only nonnegative result was a weak positive outcome with 20 mg/plate pullulan in a rec assay using *Bacillus subtilis*. Negative results were also reported in in vivo mouse micronucleus tests with $\leq 5,000$ mg/kg beta-glucan and $\leq 1,800$ mg/kg pullulan.

Carcinogenicity

Oral

Gellan gum. Groups of 50 male and 50 female Swiss Crl mice were fed a diet containing 0%, 1%, 2%, or 3% gellan gum (varied degree of acetylation; 58.5% polysaccharide) for 96 weeks (males) or 98 weeks (females).³⁹ No treatment-related effects on BWs or feed consumption were observed. No treatment-related neoplastic or nonneoplastic lesions were reported.

In another study, groups of 50 male and 50 female F₁ generation Sprague Dawley rats were exposed in utero to gellan gum and were maintained on a diet containing 0%, 2.5%, 3.8%, or 5.0% gellan gum (varied degree of acetylation; 58.5% polysaccharide) for 104 weeks.³⁹ Survival of treated male rats was decreased when compared to controls, but survival of treated female rats was better than the concurrent controls. Male rats of the 3.8% and 5.0% test groups had decreased BWs compared to controls initially and after 76 weeks. However, the researchers stated that the growth pattern of these test animals was the same as that of the controls, and the lower BWs were not indicative of toxicity. There were no neoplastic or nonneoplastic lesions

associated with dosing, and gellan gum was not carcinogenic when fed to Sprague Dawley rats.

Dextran. A group of 15 male and 15 female ACI rats were fed a diet containing 2.5% dextran (21,500 MW) for 480 days, and the control group of 20 males and 20 females was fed a basal diet.⁶² Body weight gains of treated male rats were statistically significantly decreased compared to controls. An increased incidence in tumors was not reported, and no intestinal tumors were found.

Sodium dextran sulfate. A number of studies have demonstrated that oral exposure to sodium dextran sulfate produces colon cancer in rats; the mechanism is not genotoxic. Oral administration has been shown to induce colonic inflammation, and a 2-day study in which female Fischer 344 rats were given 3% or 6% sodium dextran sulfate in the drinking water indicated that oxidative DNA damage occurred in the colonic mucosa.⁶³ The MW of sodium dextran sulfate has been found to be a factor in carcinogenic activity. Although an extensive number of studies are available in the published literature, just a few representative studies are summarized below. An example of an inflammation-related mouse colon carcinogenesis model is described, with an indication of strain dependency.

In one study evaluating the carcinogenic potential of sodium dextran sulfate (54,000 average MW; sulfur content 18.9%) in ACI rats, 10 males were fed a diet containing 10% sodium dextran sulfate, 14 males and 12 females were fed 5% in the diet, and a control group of 9 males and 9 females were given a basal diet.⁶⁴ All animals were necropsied at natural death or when killed due to moribund condition. All animals in the 10% group died 6 to 14 days after initiation of dosing, and all had severe acute nephrosis. Two animals of the 5% group died on day 14, but most of the remainder of this group lived for more than 130 days. Blood was observed on the surface of the stools of these animals at 2.5 months. The weight gain of animals of this group was decreased compared to controls. Of the 23 rats that survived for more than 130 days, 15 rats developed intestinal tumors; tumors included 5 adenomas, 5 adenocarcinomas, and 3 papillomas in the colon and 6 adenomas and 2 adenocarcinomas in the cecum. Although most rats had a single tumor upon gross observation, microscopic examination found multicentric foci of atypical hyperplasia of the glandular epithelium. No intestinal tumors were reported in the control group.

In a second study testing the same sodium dextran sulfate, 15 male and 15 female ACI rats were fed 1% in the diet for 660 days; the average daily intake was 0.15 g/d/animal.⁶⁵ A control group of 10 males and 10 females were fed a basal diet. All but 2 treated male rats survived for 350+ days. Body weight gains of the test group were similar to that of the controls. Intestinal tumors were observed in 22 treated rats; 16 papillomas, 4 squamous cell carcinomas, 2 adenomas, and 5 adenocarcinomas were reported in the colon and rectum and 1 adenocarcinoma was found in the cecum. Thirteen tumors were reported at

Table 10. Genotoxicity Studies.

Ingredient	Concentration	Vehicle	Procedure	Test system	Results	Reference
In vitro						
Gellan gum (nonacetylated; >95% polysac.)	10, 30, 100, 300, and 1,000 µg/plate	Not provided	Ames test, with and without metabolic activation	<i>Salmonella typhimurium</i> TA98, TA100, TA1537, TA1538, TA1535	Negative	39
Gellan gum (nonacetylated; >95% polysac.)	3, 5, 10, and 20 mg/mL	Not provided	DNA repair test (details not provided)	Rat hepatocytes	Negative	39
Gellan gum (nonacetylated; >95% polysac.)	3, 5, 10, and 20 mg/mL	Not provided	V79/HGRPT (details not provided)	Chinese hamster lung fibroblasts	Negative	39
Sodium dextran sulfate (54,000 MW)	1.0, 7.5, 25 mg/plate	not provided	Ames test; appropriate positive controls were used	<i>S typhimurium</i> TA100, TA98	Negative	118
Sodium dextran sulfate (54,000 MW)	0, 10, 100 µg	Distilled water	Hepatocyte primary culture (HPC)/DNA repair test; appropriate positive controls were used	Rat hepatocytes	Negative	118
Sodium dextran sulfate (54,000 MW)	0, 10, 100 µg	Distilled water	Intestinal mucosal cell (IMC)/DNA repair test; appropriate positive controls were used	Intestinal mucosal cells from rat ileum or colon and rectum	Negative	118
Beta-Glucan (as curdlan)	15-5,000 µg/plate	Sterile water	Ames test, with and without metabolic activation	<i>S typhimurium</i> TA98, TA100, TA1537, TA1538, TA1535	negative	42
Beta-Glucan (as mushroom beta-glucan from <i>Ganoderma lucidum</i>)	313-5,000 µg/plate	not provided	Ames test, with and without metabolic activation	<i>S typhimurium</i> TA98, TA100, TA102, TA1535, TA1537		119
Beta-Glucan (as curdlan)	625-5,000 µg/mL	Tissue culture medium	Chromosomal aberration assay, with and without metabolic activation	Chinese hamster ovary (CHO) cells	Negative	42
Beta-Glucan (as mushroom beta-glucan from <i>Ganoderma lucidum</i>)	313-5,000 µg/mL	Culture medium	Chromosomal aberration assay, with and without metabolic activation	CHO cells	Negative	119
Beta-Glucan (as curdlan)	12.5-5,000 µg/mL	Tissue culture medium	Tk locus test, with and without metabolic activation	Mouse lymphoma L518Y cells	Negative	42
Sodium carboxymethyl beta-glucan	0, 3-5,000 µg/plate	Milli-Q water	Ames test, with and without metabolic activation, appropriate positive controls were used;	<i>S typhimurium</i> TA98, TA100, TA1537, TA1535	Negative	120
Sodium carboxymethyl beta-glucan	0, 100-5000 µg/plate	Milli-Q water	Ames test, with and without metabolic activation; appropriate positive controls were used	<i>S typhimurium</i> TA98, TA100, TA1537, TA1535	Negative	120
Sodium carboxymethyl beta-glucan	0, 195-50,000 µg/mL	Purified water	Ames test, with and without metabolic activation; appropriate positive controls were used;	<i>S typhimurium</i> TA98, TA100, TA1537, TA1535; <i>Escherichia coli</i> WP2 <i>uvrA</i>	Negative	121
Sodium carboxymethyl beta-glucan	0, 3125-50,000 µg/mL	Purified water	Ames test, with and without metabolic activation; appropriate positive controls were used	<i>S typhimurium</i> TA98, TA100, TA1537, TA1535; <i>Escherichia coli</i> WP2 <i>uvrA</i>	Negative	121
Carboxymethyl-glucan	12.5, 25, 50, 100, and 200 µg/mL		Electrophoresis test in single-cell gel (comet assay)	CHO-k1 cells	Negative	122

(continued)

Table 10. (continued)

Ingredient	Concentration	Vehicle	Procedure	Test system	Results	Reference
Pullulan	≤ 10,000 µg/plate	Not provided	Ames test, with and without metabolic activation; appropriate positive controls were used	<i>S typhimurium</i> TA98, TA100, TA1537, TA1535; <i>Escherichia coli</i> WP2 uvrA	Negative	115
Pullulan	12 mg/mL	Not provided	Chromosomal aberration assay	Chinese hamster lung fibroblasts	Negative	123
Pullulan	20 mg/plate	Distilled water	Rec assay; with and without metabolic activation	<i>Bacillus subtilis</i>	Weak positive	124
In vivo						
Beta-glucan (as curdlan)	0, 500, 1,000, or 2,000 mg/kg	Water	Micronucleus test; dosed by gavage at 24-hour intervals; number of doses not specified; followed OECD Guideline No. 474	Male and female CD-1 mice	Negative	42
Beta-glucan (as mushroom beta-glucan from <i>Ganoderma lucidum</i>)	0, 1,250, 2,500, and 5,000 mg/kg BW	Sterile water	Micronucleus test; single dose by gavage; blood samples were taken 24, 48, and 72 hours after dosing	CD-1 mice, 5/sex/group	Negative	119
Pullulan	1,800 mg/kg BW.	Not provided	Micronucleus test; 1 intraperitoneal (IP) injection	DDY mice	Negative	125
Pullulan	4 × 1,000 mg/kg BW	Not provided	Micronucleus test; 4 IP injections in 24 hours	DDY mice	Negative	125

miscellaneous sites. No intestinal tumors were reported in the control group.

To examine the effect of MW, 3 groups of 15 male and 15 to 16 female ACI rats were fed for 480 days a diet containing 2.5% of a sodium dextran sulfate, MW 520,000, 54,000, or 9,500; each sodium dextran sulfate had a sulfur content of 18% to 19%⁶² (the 54,000 MW substance was synthesized using the dextran described previously). A control group of 20 male and 20 female rats was fed a basal diet. There was no significant difference in survival time between any of the groups. Body weight gains of male rats of the 54,000 MW diet were statistically significantly decreased compared to control males; BW gains in the other 2 groups were similar to controls. The 54,000 MW sodium dextran sulfate had the strongest carcinogenic activity, with tumors being reported similar to the studies described previously. The other 2 MW substances did not show significant carcinogenic activity; only 2 colorectal adenocarcinomas were observed in the 520,000 MW group. Colorectal squamous metaplasia was observed in most rats in all test groups. Other miscellaneous tumors were reported, but there was no statistically significant difference between treated and control groups.

A colitis-related mouse colon carcinogenesis model was developed.⁶⁶ When 8 male Crj: CD-1 mice were given a single IP injection of 10 mg/kg azoxymethane (AOM), followed by 7 days of 2% sodium dextran sulfate in the drinking water 1 week later, there was a 100% incidence of colonic adenocarcinomas and 38% of adenomas at week 20. No adenocarcinomas were observed in any of the mice dosed with AOM and sodium dextran sulfate simultaneously, 1 week of sodium dextran sulfate, and then AOM, AOM only, or sodium dextran sulfate only, and only 2 adenomas (in 10 mice) were observed when

AOM was given during sodium dextran sulfate administration. All of the adenocarcinomas were positive for β -catenin, cyclooxygenase 2, and inducible nitric oxide synthase and negative for immunoreactivity of p53.

Strain sensitivity to the above model was then examined.⁶⁷ In testing with male Balb/c, C3H/HeN, C57BL/6N, and DBA/2N mice, Balb/c mice were very sensitive to the model. C57BL/6N mice also developed colonic adenocarcinomas, but to a lesser extent. C3H/HeN and DBA/2N mice developed only a few colonic adenomas. However, the greatest inflammation response was observed in C3H/HeN mice, followed by Balb/c mice. The researchers hypothesized that the strain differences in susceptibility of colon carcinogenesis induced by AOM and sodium dextran sulfate might be influenced by the response to nitrosation stress due to inflammation as determined by the genetic background.

Antitumor Effects

Xanthan gum. Groups of C57BL/6 mice were inoculated subcutaneously (SC) with 1×10^6 B16K^b melanoma cells.⁶⁸ A suspension of 10 mg/mL xanthan gum, 100 µL, or PBS was given by gavage once every 5 days, starting 1 day prior to inoculation. Rapid tumor growth occurred in PBS-treated mice, but tumor growth was statistically significantly reduced in xanthan gum-treated mice. Spleen cell composition was analyzed 22 days after inoculation. There was no change in overall cellular composition, but natural killer cells and tumor-specific cytotoxic T-lymphocyte activity were increased by xanthan gum. All PBS-treated mice died by day 46 after inoculation; 40% of the xanthan gum-treated mice were alive at day 100. The researchers demonstrated that the antitumor effect of

xanthan gum was highly dependent on Toll-like receptor 4-mediated signaling.

Pullulan. Male Balb/c mice were used to determine the anti-tumor and antimetastatic potential of pullulan (water-soluble low-MW β -(1 \rightarrow 3) with 50%-80% branched β -(1 \rightarrow 6); MW 100,000).⁶⁹ Colon-26 cells were implanted into the spleens of mice on day 0, and groups of mice were dosed orally with 25 or 50 mg/kg or IP with 5 or 15 mg/kg pullulan for 14 consecutive days, starting 12 hours after implantation. Control groups consisted of sham-operated mice (not implanted with colon-26 cells) and mice implanted with the cells but given physiological saline or distilled water instead of pullulan. Splenic tumor weights were reduced with the 50 mg/kg oral and 15 mg/kg IP doses of pullulan but not with the other doses. Liver metastasis was significantly inhibited with 50 (but not 25) mg/kg oral and 5 and 15 mg/kg IP pullulan.

Levan. The antitumor activity of levan produced from 4 different microorganisms, that is, *Gluconacetobacter xylinus*, *Rahnella aquatilis*, *Zymomonas mobilis*, and *Microbacterium laevaniformans* (G-levan, R-levan, Z-levan, and M-levan, respectively) was evaluated using female ICR mice.⁷⁰ The MWs of these levans were 40,000, 380,000, 570,000, and 710,000, respectively. On day 0, 0.2 mL (equivalent to 3×10^6 cells) of sarcoma-180 tumor cells was implanted SC into the groin of the mice. On days 1 through 7, 200 mg/kg of the levans in distilled water was applied to the mice, and the mice were killed on day 26. The tumor growth inhibition ratio (IR) ranged from 42.2% to 69.6% for the 4 levans. M-Levan had the highest IR value, but it was not statistically significant. The IRs for R- and Z-levan were comparable, and the G-levan was significantly less effective.

Irritation and Sensitization

Nonhuman and human dermal irritation and sensitization studies are summarized in Table 11. The dermal irritation and sensitization potentials of xanthan gum, beta-glucan, and sodium carboxymethyl beta-glucan were evaluated in animal studies. Xanthan gum, up to 1%, and beta-glucan, concentration not specified, were not irritating to rabbit skin. One study with 5% aq. xanthan gum on shaved rabbit skin produced localized irritation; study details were not provided. Neither xanthan gum, tested at 0.1%, nor beta-glucan (concentration not specified) were sensitizers in guinea pigs. Sodium carboxymethyl beta-glucan was at most a slight skin irritant in guinea pigs at a concentration of 50% aq.; a 10% aq. solution was not a sensitizer in guinea pigs.

In humans, neither beta-glucan nor sodium carboxymethyl beta-glucan were irritants or sensitizers. The test concentration of beta-glucan was not specified. Sodium carboxymethyl beta-glucan was applied neat in the irritation study and as a 2% aq. solution in the sensitization study.

Phototoxicity

A human photoallergenicity study is also summarized in Table 11. A 2% aq. solution of sodium carboxymethyl beta-glucan was not photosensitizing in clinical studies.

Adverse Reactions

Dextran. Over a 5-year period (1981-1986), 12,646 dextran 70 U were administered to 5,745 patients (mean of 2.2 U/patient) undergoing gynecological surgery or a cesarean section as a plasma volume expander.⁷¹ Fifteen immediate dextran-induced anaphylactoid reactions were reported, with an incidence of 1 reaction/383 patients treated. Life-threatening reactions occurred in 7 of these patients.

Ocular Irritation

Ocular irritation studies are summarized in Table 12. Xanthan gum, 1%, and up to 0.8% gellan gum were not ocular irritants in rabbit eyes, and up to 0.5% gellan gum was not irritating to human eyes. The ocular irritation potential of sodium carboxymethyl beta-glucan was described as weakly irritating in a HET-CAM assay but was practically nonirritating in rabbit eyes.

Summary

Microbial polysaccharide gums can be produced intercellularly, by the cell wall, or exocellularly. Exocellular polysaccharide gums constantly diffuse into the cell culture medium and are easily isolated, while cell wall and intercellular polysaccharide gums are more difficult to separate from cell biomass. Many of the 34 microbial polysaccharide gums discussed in this safety assessment are produced exocellularly. They are reported to have numerous functions in cosmetics, including emulsion stabilizer, film former, binder, viscosity-increasing agent, and skin-conditioning agent.

The same microbial polysaccharide gums can often be produced by more than 1 organism. For example, beta-glucan can be produced by fungi, yeasts, and grains and levan can be produced by bacteria, yeasts, or fungi. The properties of the microbial polysaccharide gums can vary widely based on, among other parameters, the side groups, the ester substituents, or bacterial strains. These polysaccharide gums are generally very large molecules, and the MW of each ingredient can vary considerably.

Xanthan gum is reported to be used in almost every category of cosmetic product, with 3,470 reported uses. Biosaccharide gum-1, sclerotium gum, and beta-glucan are reported to be used in 346, 193, and 137 cosmetic formulations, respectively. All other in-use ingredients have less than 70 uses. The ingredient with the highest concentration of use is pullulan; it is used at up to 12% in leave-on formulations and 17% in a breath freshener. Both xanthan gum and biosaccharide gum-1 are used at up to 6% in leave-on formulations and xanthan gum cross-polymer and biosaccharide gum-4 are used at 5% in leave-on

Table 11. Dermal Irritation and Sensitization.

Ingredient	Animals/subjects/ group	Dose/concentration/ vehicle	Procedure	Results	Reference
Irritation—nonhuman					
Xanthan gum	Rats	1%; vehicle not provided	Solution was applied daily for 15 days (details not provided)	Not an irritant	38
Xanthan gum	6 rabbits	0.5 mL of a 1% solution	Primary cutaneous irritation test; occlusive patches on intact and abraded skin	Nonirritant; primary irritation index (PII)—0.13	126
Xanthan gum	6 rabbits	0.5 mL of a 1% solution	Primary cutaneous irritation test; occlusive patches on intact and abraded skin	Nonirritant; PII—0.13	126
Xanthan gum	3 rabbits	2 mL/animal of a 1% solution	6-week cumulative cutaneous irritation test, 5 applications/wk	Very well tolerated; mean maximum irritation index—0	126
Xanthan gum	3 rabbits	1%	6-week cumulative cutaneous irritation test, 5 applications/wk	Very well tolerated; MMI—0	126
Xanthan gum	Rabbits	5% aq.	Daily applications to shaved skin (details not provided)	Localized irritation with bleeding and cracking; the effects may have been due to continuous moistening of the skin	104
Beta-glucan	Rabbits	Not provided	Primary skin irritation test (details not provided)	Not an irritant	91
Beta-glucan	Rabbits	Not provided	Repeated skin irritancy test (details not provided)	Not an irritant	91
Sodium carboxymethyl beta-glucan (>90% pure)	Japanese white rabbits, 6 M	0.5 g moistened with 0.2 mL distilled water	Occlusive patches were applied to intact and abraded shaved skin for 24 hours	Nonirritating to mildly irritating; primary irritation index (PII) of 0.33/8 at test site and 0.29/8 for adhesive control; no irritation was reported for intact skin	127
Sodium carboxymethyl beta-glucan (>90% pure)	3 guinea pigs	2%, 10%, or 50% (wt/vol) in distilled water	24-hour occlusive patch test; this test was used to determine test concentration for the maximization study described below	Slight skin irritation was observed at a concentration of 50%	128
Irritation—human					
Beta-glucan	Not provided	Not provided	Occlusive patch test	No an irritant	91
Sodium carboxymethyl beta-glucan (>90% pure)	40 subjects; 27 M/13 F	Applied neat; small amount of Vaseline was used for adhesion	24-hour occlusive patch test; 0.1 g of test material was applied	Not a primary skin irritant; no irritation was observed	129
Sensitization—nonhuman					
Xanthan gum	Guinea pigs, 18 M	0.1%; vehicle not provided	Intradermal challenge test; test solution was injected intracutaneously 3×/wk for 10 injections; the challenge was performed after a 10-day nontreatment period	Not a sensitizer	38
Beta-glucan	Guinea pigs	Not provided	Skin sensitization test (details not provided)	Not a sensitizer	91
Sodium carboxymethyl beta-glucan (>90% pure)	Hartley guinea pigs, 6 F	10% in distilled water	Maximization study using Freund complete adjuvant and SLS; 0.1% aq. 2,4-dinitro-1-chlorobenzene was used as the positive control; dose volumes were 0.1 mL for intradermal induction, 0.2 mL for dermal induction, and 0.1 mL for challenge	Not a sensitizer	128

(continued)

Table 11. (continued)

Ingredient	Animals/subjects/ group	Dose/concentration/ vehicle	Procedure	Results	Reference
Sensitization—human					
Beta-glucan (as curdlan)	213 subjects; M	Dose not provided; was an aq. paste	Modified Draize “multiple insult” patch test; occlusive patches were applied every other day for 10 applications; a 48-hour challenge was performed after a 10- to 14-day nontreatment period	Trace, insignificant, irritation observed during induction; not a sensitizer	42
Sodium carboxymethyl beta-glucan	32 subjects; 8 M/24 F	Induction and challenge: 2% in distilled water	Induction: 9 24-h occlusive patches were applied; challenge: occlusive patch applied after a 10-14 day non-treatment period to a previously unpatched site; reactions were graded on a scale of 0-4	Not a sensitizer; during induction, 2 subjects had doubtful reactions and 1 had a grade 1 reaction	130
Photoallergy—human					
Sodium carboxymethyl beta-glucan	8 male/24 female subjects	Induction and challenge: 2% in distilled water	Induction: six 24-hour patches were applied to nontanned skin; the test site was irradiated with 2× MED UVB within 10 minutes after patch removal; challenge: applied after a 10- to 14-day nontreatment period, a 24-hour patch was applied to a previously unpatched site; the site was irradiated with 18 J/cm ² UVA within 10 minutes after patch removal; MULTITESTER light source was used; 0-4 scale used for scoring	Not photoallergenic; 31 subjects had grade 1 skin reactions, and one had a grade 2 reaction, to UVB exposure during induction; 1 subject had a doubtful reaction after challenge	132

Abbreviations: aq., aqueous; F, female; M, male.

Table 12. Ocular Irritation.

Ingredient	Animals/ subjects/group	Concentration/vehicle	Procedure	Results	Reference
Alternative studies Sodium carboxymethyl beta-glucan		5% stock solution containing 2% sodium carboxymethyl beta- glucan	Ocular tolerance test using HET-CAM method	Weakly irritant	131
Nonhuman Xanthan gum	Rabbits	1%	Ocular irritation test	Nonirritating; acute ocular irritation index (AOII)— 2.50/110	126
Xanthan gum	Rabbits	1%	Ocular irritation test	Nonirritating; AOII—5.83/110	126
Xanthan gum	Rabbits	1%	Instilled in the conjunctival sac of rabbit eyes for 5 days (details were not provided)	Not irritating	38
Gellan gum	New Zealand White rabbits, 3 males	0.2% or 0.3%	Draize study; 50 μ L of the test solution was instilled into the conjunctival sac of the eye 3 times a day for 10 days	Not irritating	132
Gellan gum	Albino rabbits	0.8%	Details not provided	Not irritating	133
Sodium carboxymethyl beta-glucan (>90% pure)	Japanese white rabbits, 3 males	Undiluted	0.1 mg were instilled into the conjunctival sac of 1 eye, and the contralateral eye served as an untreated control; the eyes of 1 group were rinsed 1 minute after instillation	Practically nonirritating; slight redness of the conjunctiva in both washed and unwashed eyes at 1 hour, considered due to the powder	134
Human Gellan gum	3 subjects	0.1%-0.5%; vehicle not specified	25 μ L of a gel formulation was instilled in the conjunctival sac of the eye, remained in contact with the eye for 9 to 52 minutes	Not irritating	132

formulations. All other in-use ingredients are used at concentrations of $\leq 3\%$.

Xanthan gum, gellan gum, and beta-glucan are approved as direct food additives, and xanthan gum and dextran are approved indirect food additives. Xanthan gum and dextran also have pharmaceutical applications.

Xanthan gum, orally administered, is slowly broken down in the gut by enzymatic and nonenzymatic mechanisms and can be absorbed in some form to some extent. The absorbed fraction does not accumulate in the tissues and can be completely metabolized to CO_2 . Gellan gum, orally administered, does not breakdown to any substantial extent in the gut and is only very poorly absorbed.

Dextrans, 3,000 to 20,000 MW, are not absorbed through intact skin in humans. They are absorbed through the dermis (up to 38% for 3,000 MW) if the epidermis is removed (eg, via mini erosion); absorption in this case is inversely proportional to MW. Orally administered dextrans, 4,400 to 40,500 MW, are not absorbed to any appreciable extent in the gut (very low bioavailability). If dextrans were absorbed to a significant extent through the skin, animal studies in which dextrans were administered IV indicate that their half-lives in blood plasma would be directly related to the MW; they would be excreted in the urine to an extent that is inversely related to the MW; the lower MW dextrans would tend to be readily excreted unchanged in the urine; and the higher MW dextrans would have the potential to accumulate and to be broken down in the liver and other tissues and would be more likely to be excreted in the urine in a dose-dependent manner than the lower MW dextrans. Dextran sulfate, 7,000 to 8,000 MW, orally administered, is only very poorly absorbed in the human GI tract. After IV administration, it is rapidly excreted, mostly intact, in the urine, although at least some of it can accumulate in the tissues (to a substantially greater extent than observed after oral exposure), and some of it appears to be incorporated into glycogen and other substances in the body.

Beta-glucan in a topically applied solution can penetrate into the epidermis and dermis. There appears to be no information about its absorption through the skin into the bloodstream. Orally administered, it is readily metabolized to CO_2 , at least partially by microflora in the gut. Beta-glucan is not well absorbed or eliminated after IP injection. Pullulan, orally administered, is hydrolyzed to some small extent in the gut. It is partially hydrolyzed by amylases in the upper GI tract of human subjects and is subject to breakdown by intestinal microflora to form short-chain fatty acids; the rate of the latter depends on the degree of polymerization. However, pullulan can be essentially completely broken down to short-chain fatty acids in the human gut. It has not been determined to what extent, if any, the products of hydrolysis can be absorbed.

The acute toxicity of xanthan gum, gellan gum, beta-glucan, sodium carboxymethyl beta-glucan, and pullulan was assessed orally in mice, rats, and/or dogs, and dextran sulfate and beta-glucan were tested by IP and IV dosing in mice and rats. There was no notable toxicity observed in these studies. In acute inhalation studies, the LC_{50} of xanthan gum was >21 mg/L in

rabbits and of gellan gum was >5.06 mg/L in rats. A single 180-minute exposure of humans office dust containing 10 mg curdlan/g dust resulted in decreased nasal volume, swelling in the nasal turbinates, and an increase in nasal eosinophils when compared to "clean" dust.

The inflammatory response following a single exposure to beta-glucan (as curdlan) and to pullulan was also evaluated in guinea pigs. Mostly, no effect or a slight decrease in inflammatory cells in lung lavage was observed. A 4-hour inhalation exposure to beta-glucan in dust by guinea pigs produced a delayed subacute nasal congestion when compared to dust without beta-glucan and resulted in decreased nasal volume. Industrial exposure to xanthan gum powder did not appear to cause significant acute or chronic pulmonary effects. Using artificial skin, 5% levan had an anti-inflammatory effect in irritated skin.

Repeated dose oral toxicity of xanthan gum was evaluated in rats and dogs, of gellan gum in rats, dogs, and monkeys, of dextran in rats, of beta-glucan in mice, rats, and dogs, and of pullulan in rats. Most of the studies were dietary, and study durations lasted up to 2 years. Most observations were related to changes in feed consumption and intestinal effects. Inhalation exposure to 100 $\mu\text{g}/\text{mL}$ beta-glucan (as curdlan), 4 h/d, 5 d/wk for 4 weeks, did not have an effect on the cells of the lung lavage or cell wall, and there were no microscopic lesions of the lung. With IP administration, no toxicity was reported when mice were dosed 10 times over 2 weeks with 5 mg xanthan gum in 0.5 mL water or mice or guinea pigs were dosed with 250 mg/kg BW beta-glucan for 7 days. Intravenous administration of 40 and 1,000 mg/kg BW beta-glucan for 30 days resulted in hepatosplenomegaly in mice.

No toxic effects were observed in human subjects with oral ingestion of 150 mg/kg/d xanthan gum or 175 to 200 mg/kg/d gellan gum for 23 days, 10 g of dextran or pullulan for 14 days, or 6 to 50 g/d beta-glucan for up to 28 days. No significant or chronic effects in pulmonary function were reported in groups exposed occupationally to xanthan gum.

Dietary reproductive and developmental toxicity studies were conducted with xanthan gum, gellan gum, and beta-glucan. No reproductive or developmental effects were reported in a 3-generation reproductive study in which rats were fed diets containing up to 5 g/kg BW/day xanthan gum. Gellan gum, up to 5%, did not have a fetotoxic or teratogenic effect on rats. Dietary administration of beta-glucan in rats in reproductive and developmental studies did not have any reproductive effects, but there were statistically significant decreases in BWs and BW gains in offspring and parental animals. In a teratogenicity study in which rabbits were dosed with 5 g/kg BW/d beta-glucan, an increase in resorptions in the 5 g/kg group was considered similar to the other test groups and the controls, and 5 g/kg beta-glucan was not teratogenic in rabbits.

The *in vitro* genotoxicity of gellan gum (≤ 20 mg/mL), sodium dextran sulfate (≤ 25 mg/plate), beta-glucan ($\leq 5,000$ $\mu\text{g}/\text{plate}$ or /mL), sodium carboxymethyl beta-glucan ($\leq 50,000$ $\mu\text{g}/\text{plate}$ or /mL), and pullulan (≤ 12 mg/mL) was evaluated in Ames test, chromosomal aberration assays, and/or

DNA repair tests, with and without metabolic activation. Results were negative in all of these tests. The only nonnegative result was a weak positive result with 20 mg/plate pullulan in a rec assay using *Bacillus subtilis*. Negative results were also reported in in vivo mouse micronucleus tests with $\leq 5,000$ mg/kg beta-glucan and $\leq 1,800$ mg/kg pullulan.

Dietary studies examining the carcinogenic potential of $\leq 5\%$ gellan gum and 2.5% dextran reported that neither of these ingredients caused an increase in tumors. However, a number of studies have demonstrated that oral exposure to sodium dextran sulfate produces colon carcinogenesis in rats; the mechanism is nongenotoxic. In one study, the MW of sodium dextran sulfate was a factor in carcinogenic activity; a 54,000 MW sodium dextran sulfate produced colorectal tumors, but 9,500 and 520,000 MW sodium dextran sulfate did not have significant carcinogenic activity. Oral administration has been shown to induce colonic inflammation, and a 2-day study in which female Fischer 344 rats were given 3% or 6% sodium dextran sulfate in the drinking water indicated that oxidative DNA damage occurred in the colonic mucosa. An inflammation-related mouse colon carcinogenesis model indicated that the development of colonic tumors is strain dependent and that Balb/c mice were very sensitive to the model. C57BL/6N mice also developed tumors, but to a lesser extent, whereas C3H/HeN and DBA/2N mice only developed a few tumors.

Oral administration of 10 mg/mL xanthan gum had an anti-tumor effect in mice inoculated with melanoma cells, and 50 mg/kg pullulan, but not 15 mg/kg, significantly inhibited tumor growth in mice following implantation of colon-26 cells. Levan did not have an antitumor effect in mice.

Nonhuman and human dermal irritation and sensitization studies are summarized in Table 11. The dermal irritation and sensitization potentials of xanthan gum, beta-glucan, and sodium carboxymethyl beta-glucan were evaluated in animal studies. Xanthan gum, up to 1%, and beta-glucan, concentration not specified, were not irritating to rabbit skin. One study with 5% aq. xanthan gum on shaved rabbit skin produced localized irritation; however, study details were not provided. Neither xanthan gum, tested at 0.1%, nor beta-glucan (concentration not specified) was sensitizers in guinea pigs. Sodium carboxymethyl beta-glucan was at most a slight skin irritant in guinea pigs at a concentration of 50% aq.; a 10% aq. solution was not a sensitizer in guinea pigs.

In humans, neither beta-glucan nor sodium carboxymethyl beta-glucan was irritants or sensitizers. The test concentration of beta-glucan was not specified. Sodium carboxymethyl beta-glucan was applied neat in the irritation study and as a 2% aq. solution in the sensitization study. A 2% aq. solution of sodium carboxymethyl beta-glucan was not photosensitizing in clinical studies.

Xanthan gum, 1%, and gellan gum, up to 0.8%, were not ocular irritants in rabbit eyes, and up to 0.5% gellan gum was not irritating to human eyes. The ocular irritation potential of sodium carboxymethyl beta-glucan was described as weakly irritating in a HET-CAM assay, but the ingredient was practically nonirritating in rabbit eyes.

Discussion

Microbial polysaccharide gums are produced by a wide variety of microorganisms, and some can also be isolated from plants, for example, beta-glucan can be isolated from barley and oats. Although these ingredients are produced primarily by microbial sources, the cosmetic ingredients are purified during manufacture, thus, microbial contamination is not a concern.

The Panel determined that, even though there are some data gaps, the available data on polysaccharide gums included in this safety assessment may be extrapolated to support the safety of the entire group. And although there was no specific data on the hydroxypropyltrimonium chloride compounds, data included in a previous CIR safety assessment on trimonium ingredients are applicable in determining the safety of the 3 hydroxypropyltrimonium chloride compounds included in this report.

Parenterally administered polysaccharides appear to be bio-transformed to a limited but variable extent in animal and human studies. However, these very large compounds appear not to be significantly absorbed through the skin and would have negligible bioavailability. Coupled with a lack of significant toxicity associated with other routes of exposure, the CIR Expert Panel determined that systemic effects were unlikely to result from topical application of cosmetics containing these ingredients.

One study reported that 5% aq. xanthan gum caused irritation. However, this was the only finding of irritation among almost 20 studies on microbial polysaccharide gums. Given the absence of study details, including no mention of a control group, the Panel concluded that the irritation likely was the result of the study methodology (eg, shaved skin) and not by the xanthan gum. There was no evidence of sensitization in human or nonhuman testing.

The Panel also remarked on the induction of colon cancer in rodents with oral exposure to sodium dextran sulfate. Sodium dextran sulfate is a commonly used model for induction of colitis in a well-characterized mouse model to study colitis, and while the mode of action is unknown, it is not relevant to human cosmetic exposure.

Finally, the Panel discussed the issue of incidental inhalation exposure to microbial polysaccharide gums from powders and other products that may be aerosolized. These ingredients are reportedly used at concentrations up to 1% in cosmetic products that may be aerosolized and up to 6% in other products that may become airborne. The Panel noted that 95% to 99% of droplets/particles would not be respirable to any appreciable amount. Furthermore, droplets/particles deposited in the nasopharyngeal or bronchial regions of the respiratory tract present no toxicological concerns based on the chemical and biological properties of this ingredient. Coupled with the small actual exposure in the breathing zone and the concentrations at which the ingredients are used, the available information indicates that incidental inhalation would not be a significant route of exposure that might lead to local respiratory or systemic effects. Results of acute inhalation studies with xanthan gum

and gellan gum produced no significant toxicity nor did results from a 4-week inhalation study of beta-glucan in guinea pigs. Industrial exposure to xanthan gum powder caused no significant acute or chronic effects in pulmonary function. The Panel also considered the data available to characterize the potential for microbial polysaccharide gums to cause systemic toxicity, irritation, sensitization, or other effects. The Panel noted that testing with a number of microbial polysaccharide gums demonstrated they did not produce systemic toxicity in oral studies; they are not reproductive or developmental toxicants, are not genotoxic, and are not considered irritants or sensitizers. A detailed discussion and summary of the Panel's approach to evaluating incidental inhalation exposures to ingredients in cosmetic products is available at <http://www.cir-safety.org/cir-findings>.

Conclusion

The CIR Expert Panel concluded the microbial polysaccharide gums listed below are safe in the present practices of use and concentration in cosmetics.

Xanthan gum
 Hydroxypropyl xanthan gum*
 Undecylenoyl xanthan gum*
 Dehydroxanthan gum
 Xanthan gum crosspolymer
 Xanthan hydroxypropyltrimonium chloride*
 Gellan gum
 Welan gum*
 Biosaccharide gum-1
 Biosaccharide gum-2
 Biosaccharide gum-3*
 Biosaccharide gum-4
 Biosaccharide gum-5*
 Pseudoalteromonas exopolysaccharides*
 Dextran
 Carboxymethyl dextran*
 Dextran hydroxypropyltrimonium chloride*
 Sodium carboxymethyl dextran
 Dextran sulfate
 Sodium dextran sulfate
 Sclerotium gum
 Hydrolyzed sclerotium gum
 Beta-glucan
 Beta-glucan hydroxypropyltrimonium chloride*
 Beta-glucan palmitate*
 Hydrolyzed beta-glucan*
 Oxidized beta-glucan*
 Sodium carboxymethyl beta-glucan
 Pullulan
 Myristoyl pullulan*
 Levan*
 Rhizobian gum
 Hydrolyzed rhizobian gum
 Alcaligenes polysaccharides

*Ingredients in this group not in current use to be used in the future, the expectation is that they would be used in product categories and at concentrations comparable to others in the group.

Authors' Note

Unpublished sources cited in this report are available from the Director, Cosmetic Ingredient Review, Washington, DC, USA.

Author Contributions

M. Fiume contributed to conception and design, contributed to acquisition, analysis, and interpretation, and drafted the manuscript. B. Heldreth contributed to conception and design, contributed to acquisition, analysis, and interpretation, drafted the manuscript, and critically revised the manuscript. L. Gill contributed to conception and design, contributed to analysis and interpretation, and critically revised the manuscript. F. Alan Andersen, W. Bergfeld, D. Belsito, R. Hill, C. Klaassen, D. Liebler, J. Marks, R. Shank, T. Slaga, and P. Snyder contributed to conception and design, contributed to analysis and interpretation, and critically revised the manuscript. All authors gave final approval and agree to be accountable for all aspects of work ensuring integrity and accuracy.

Declaration of Conflicting Interests

The author(s) declared no potential conflicts of interest with respect to the research, authorship, and/or publication of this article.

Funding

The author(s) disclosed receipt of the following financial support for the research, authorship, and/or publication of this article: The articles in this supplement were sponsored by the Cosmetic Ingredient Review. The Cosmetic Ingredient Review is financially supported by the Personal Care Products Council.

References

1. Gottschalck TE, Breslawec HP. *International Cosmetic Ingredient Dictionary and Handbook*. 14 ed. Washington, DC: Personal Care Products Council; 2012.
2. Johnson W Jr, Heldreth B, Bergfeld WF, et al. Safety assessment of galactomannans as used in cosmetics. *Int J Toxicol*. 2015;34(I Suppl):35S-65S.
3. Becker LC, Bergfeld WF, Belsito DV, et al. Final report of the safety assessment of hyaluronic acid, potassium hyaluronate, and sodium hyaluronate. *Int J Toxicol*. 2009; 28(suppl 1):5-67.
4. Elder RL, ed. Final report on the safety assessment of tragacanth gum. *J Am Coll Toxicol*. 2011;6(1):1-22.
5. Andersen FA, ed. Final report on the safety assessment of acacia catechu gum, acacia concinna fruit extract, acacia dealbata leaf extract, acacia dealbata leaf wax, acacia decurrens extract, acacia farnesiana extract, acacia farnesiana flower wax, acacia farnesiana gum, acacia senegal extract, acacia senegal gum, and acacia senegal gum extract. *Int J Toxicol*. 2005;24(suppl 3): 75-118.

6. Becker LC, Bergfeld WF, Belsito DV, et al. Safety assessment of trimoniums as used in cosmetics. *Int J Toxicol*. 2012; 31(6 Suppl):296S-341S.
7. Driscoll M, Hansen R, Ding C, Cramer DE, Yan J. Therapeutic potential of various β -glucan sources in conjunction with anti-tumor monoclonal antibody in cancer therapy. *Cancer Biol Ther*. 2009;8(3):218-225.
8. Rhee S, Song K, Kim C, et al. Levan. Chapter: 14. In: Vandamme EJ, De Baets S, Steinbüchel A, eds. *Biopolymers*. Weinheim, Germany: Wiley VCH; 2002:351-377.
9. Manjanna KM, Shivakumar B, Pramodkumar TM. Natural exopolysaccharides as novel excipients in drug delivery: a review. *Arch Appl Sci Res*. 2009;1(2):230-253.
10. Mathur V, Mathur NK. Microbial polysaccharides based food hydrocolloid additives. *Sci Tech Entrepren*. 2006:1-10.
11. Heinze T, Liebert T, Heublein B, Hornig S. Functional polymers based on dextran. *Adv Polym Sci*. 2006;205:199-291.
12. Moorhouse R. Structure/property relationships of a family of microbial polysaccharides. Symposium on the Applications and Modifications of Industrial Polysaccharides. 1987.
13. Kuo M, Mort AJ, Dell A. Identification and location of L-glycerate, an unusual acyl substituent in gellan gum. *Carbohydr Res*. 1986;156:173-187.
14. Valli RC, Miskiel FJ. Gellan gum. *Food Sci Technol (New York, NY, United States)*. 2001;113(Handbook of Dietary Fiber): 695-720.
15. Food and Drug Administration. *Frequency of Use of Cosmetic Ingredients. FDA Database*. 2012. Washington, DC: Food and Drug Administration; 2012.
16. Personal Care Products Council. 10-26-2011. Concentration of Use by FDA Product Category: Microbial Polysaccharides. Unpublished data submitted by Personal Care Products Council. 10 pages.
17. Personal Care Products Council. 3-28-2012. Concentration of Use by FDA Product Category: Xanthan Hydroxypropyltrimonium Chloride. Unpublished data submitted by Personal Care Products Council. 1 pages.
18. Personal Care Products Council. 7-9-2012. Comments on the Tentative Report of Microbial Polysaccharide Gums. Unpublished data submitted by Personal Care Products Council. 1 pages.
19. Johnsen MA. The influence of particle size. *Spray Technology and Marketing*. 2004;14(11):24-27.
20. Rothe H. Special Aspects of Cosmetic Spray Evaluation. 9-26-2011. Unpublished data presented at the 26 September CIR Expert Panel meeting. Washington, DC.
21. Rothe H, Fautz R, Gerber E, et al. Special aspects of cosmetic spray safety evaluations: principles on inhalation risk assessment. *Toxicol Lett*. 2011;205(2):97-104.
22. Bremmer HJ, Prud'homme de Lodder LCH, Engelen JGM. Cosmetics Fact Sheet: To assess the risks for the consumer. Updated version for ConsExpo 4. 2006. Report No. RIVM 320104001/2006. 1-77.
23. European Commission. Health and Consumers Cosmetic Cosing Database. Web site. <http://ec.europa.eu/consumers/cosmetics/cosing/>. 2011. Accessed October 20, 2011.
24. Food and Drug Administration. 21CFR172.695. Web site. <http://www.accessdata.fda.gov/scripts/cdrh/cfdocs/cfCFR/CFRSearch.cfm?FR=172.695&CFRPart=&FRSearch=>. 4-1-2011. Accessed October 21, 2011.
25. Food and Drug Administration. 21CFR172.665. Web site. <http://www.accessdata.fda.gov/scripts/cdrh/cfdocs/cfCFR/CFRSearch.cfm?FR=172.665&CFRPart=&FRSearch=>. 4-1-2011. Accessed October 21, 2011.
26. Food and Drug Administration. 21CFR176.170. Web site. <http://www.accessdata.fda.gov/scripts/cdrh/cfdocs/cfCFR/CFRSearch.cfm?FR=176.170&CFRPart=&FRSearch=>. 4-1-2011. Accessed October 21, 2011.
27. Food and Drug Administration. 21CFR172.809. Web site. <http://www.accessdata.fda.gov/scripts/cdrh/cfdocs/cfCFR/CFRSearch.cfm?fr=172.809>. 4-1-2011. Accessed November 19, 2011.
28. Food and Drug Administration. 21CFR186.1275. Web site. <http://www.accessdata.fda.gov/scripts/cdrh/cfdocs/cfCFR/CFRSearch.cfm?FR=186.1275&CFRPart=&FRSearch=>. 4-1-2011. Accessed November 19, 2011.
29. Joint FAO/WHO Expert Committee on Food Additives. Evaluation of certain food additives and contaminants. Thirtieth report of the Joint FAO/WHO Expert Committee on Food Additives. WHO Technical Report Series 751. Geneva, Switzerland: World Health Organization; 1987. Web site. http://whqlibdoc.who.int/trs/WHO_TRS_751.pdf. Accessed March 22, 2012.
30. Joint FAO/WHO Expert Committee on Food Additives. Evaluation of certain food additives and contaminants. WHO Technical Report Series 806. Thirty-Seventh Report of the JOINT FAO/WHO Expert Committee on Food Additives. Geneva, Switzerland: World Health Organization; 1991. Web site. http://whqlibdoc.who.int/trs/WHO_TRS_806.pdf. Accessed March 22, 2012.
31. Joint FAO/WHO Expert Committee on Food Additives. Evaluation of certain food additives. WHO Technical Report Series 934. Sixty-fifth report of the joint FAO/WHO Committee on Food Additives. Geneva, Switzerland: World Health Organization (WHO); 2006. Web site. http://whqlibdoc.who.int/trs/WHO_TRS_934_eng.pdf. Accessed September 22, 2011.
32. Chaen H. Pullulan. Food Stab., Thickeners Gelling Agents. 2010. CAPLUS AN 2010:484781(Conference; General Review).
33. Leung AY, Foster S. *Encyclopedia of Common Natural Ingredients Used in Foods, Drugs, and Cosmetics*. 2nd ed. New York NY: John Wiley & Sons, Inc; 1996.
34. Food and Drug Administration. 21CFR349.12. Web site. <http://www.accessdata.fda.gov/scripts/cdrh/cfdocs/cfCFR/CFRSearch.cfm?FR=349.12&CFRPart=&FRSearch=>. 4-1-2011. Accessed October 21, 2011.
35. Merck & Co, Inc. The Merck Index. 2011. Web site. <http://themerckindex.cambridgesoft.com/themerckindex/Forms/Search/ContentArea/ChemBioVizSearch.aspx?FormGroupId=200000&AppName=THEMERCINDEX&AllowFullSearch=true&KeepRecordCountSynchronized=false&SearchCriteriaId=5&SearchCriteriaValue=77-93-0&CurrentIndex=0>. Accessed 2011.

36. Pillai R, Redmond M, Röding J. Anit-wrinkle therapy: significant new findings in the non-invasive cosmetic treatment of skin wrinkles with beta-glucan. *J Cosmet Sci.* 2005; 56(3):211.
37. Svedman P, Lundin S, Hoeglund P, Hammarlund C, Malmros C, Pantzar N. Passive drug diffusion via standardized skin mini-erosion; methodological aspects and clinical findings with new device. *Pharm Res.* 1996;13(9):1354-1359.
38. Inchem. Xanthan Gum. Web site. <http://www.inchem.org/documents/jecfa/jecmono/v21je13.htm>. 2010. Accessed November 15, 2011.
39. Lin FSD. Gellan Gum—First Draft. 1991. Web site. <http://www.inchem.org/documents/jecfa/jecmono/v28je17.htm>. Accessed November 15, 2011.
40. Mehvar R, Shepard TL. Molecular-weight-dependent pharmacokinetics of fluorescein-labeled dextrans in rats. *J Pharm Sci.* 1992;81(9):908-912.
41. Foster BC, Gallicano KD, Whitehouse LW, McGilveray IJ, Khan SR. Dextran sulfate disposition in the rat. *Biopharm Drug Dispos.* 1990;11(7):595-606.
42. Joint FAO/WHO Expert Committee on Food Additives. Safety Evaluation of Certain Food Additives and Contaminants. WHO Food Additive Series 44. Curdlan. 2000. Web site. <http://www.inchem.org/documents/jecfa/jecmono/v44jec04.htm>. Accessed November 15, 2011.
43. Oku T, Yamada K, Hosoya N. Effect of pullulan and cellulose on the gastrointestinal tract of rats. *Eiyo to Shokuryo (Nutr Diets).* 1982;32(4):235-241.
44. Dixon B, Abbott PJ, Verger P, DiNovi M. Pullulan—First draft. 2005. Web site. <http://www.inchem.org/documents/jecfa/jecmono/v56je05.pdf>. Accessed November 9, 2011.
45. Lorentsen KJ, Hendrix CW, Collins JM, et al. Dextran sulfate is poorly absorbed after oral administration. *Ann Intern Med.* 1989; 111(7):561-566.
46. Yoneyama M, Okada K, Maddai T, Aga H, Sakai S, Ichikawa T. Effects of pullulan intake in humans. *Starch Chem (Denpun Kagaku).* 1990;37(3):123-127.
47. Friberg U, Graf W, Åberg B. Effects of prolonged dextran administration to rabbits. *Acta Pharmacol Toxicol.* 1953;9(3): 220-234.
48. Mehvar R, Robinson MA, Reynolds JM. Dose dependency of the kinetics of dextrans in rats: effects of molecular weight. *J Pharm Sci.* 1995;84(7):815-818.
49. Yamaoka T, Tabata Y, Ikada Y. Body distribution profile of polysaccharides after intravenous administration. *Drug Delivery.* 1993;1(1):75-82.
50. Kaneo Y, Tanaka T, Nakano T, Yamaguchi Y. Evidence for receptor-mediated hepatic uptake of pullulan in rats. *J Control Release.* 2001;70(3):365-373.
51. Arturson E., Wallenius G. The intravascular persistence of dextran of different molecular sizes in normal humans. *Scand J Clin Lab Invest.* 1964;16(1):76-80.
52. Straszek SP, Adamcakova-Dodd A, Metwali N, Pedersen OF, Sigsgaard T, Thorne PS. Acute effect of glucan-spiked office dust on nasal and pulmonary inflammation in guinea pigs. *J Toxicol Environ Health A.* 2007;70(22):1923-1928.
53. Fogelmark B, Sjostrand M, Williams D, Rylander R. Inhalation toxicity of (1→3)-β-D-glucan: recent advances. *Mediators Inflamm.* 1997;6(4):263-265.
54. Fogelmark B, Goto H, Yuasa K, Marchat B, Rylander R. Acute pulmonary toxicity of inhaled β-1—3-glucan and endotoxin. *Agents Actions.* 1992;35(1-2):50-56.
55. Bonlokke JH, Stridh G, Sigsgaard T, et al. Upper-airway inflammation in relation to dust spiked with aldehydes or glucan. *Scand J Work Environm Health.* 2006;32(5):374-382.
56. Kim H, Cho H, Moon S, et al. Effects of β-glucan from *Aureobasidium pullulans* on acute inflammation in mice. *Arch Pharm Res.* 2007;30(3):323-328.
57. Kim KH, Chung CB, Kim YH, Kim KS, Han CS, Kim CH. Cosmeceutical properties of levan produced by *Zymomonas mobilis*. *J Cosmet Sci.* 2005;56(6):395-406.
58. Eastwood MA, Brydon WG, Anderson DMW. The dietary effects of xanthan gum in man. *Food Addit Contam.* 1987; 4(1):17-26.
59. Mitsushashi M, Yonetama M, Sakai S. Growth promoting agent for bacteria containing pullulan with or without dextran. 1990. (EP 0 382 355 B1): Secondary reference in INCHEM (2005).
60. Sargent EV, Adolph J, Clemmons MK, Kirk GD, Pena BM, Fedoruk MJ. Evaluation of flu-like symptoms in workers handling xanthan gum powder. *J Occup Med.* 1990;32(7): 625-630.
61. Woodard G, Woodard MW, McNeely WH, Kovacs P, Cronin MTI. Xanthan gum. Safety evaluation by two-year feeding studies in rats and dogs and a three-generation reproduction study in rats. *Toxicol Appl Pharmacol.* 1973;24(1):30-36.
62. Hirono I, Kuhara K, Yamaji T, Hosaka S, Golberg L. Carcinogenicity of dextran sulfate sodium in relation to its molecular weight. *Cancer Lett (Shannon, Ireland).* 1983;18(1):29-34.
63. Tardieu D, Jaeg JP, Cadet J, Embvani E, Corpet DE, Petit C. Dextran sulfate enhances the level of an oxidative DNA damage biomarker, 8-oxo-7,8-dihydro-2'-deoxyguanosine, in rat colonic mucosa. *Cancer Lett (Shannon, Ireland).* 1998;134(1):1-5.
64. Hirono I, Kuhara K, Hosaka S, Tomizawa S, Golberg L. Induction of intestinal tumors in rats by dextran sulfate sodium. *J Natl Cancer Inst.* 1981;66(3):579-583.
65. Hirono I, Kuhara K, Yamaji T, Hosaka S, Golberg L. Induction of colorectal squamous cell carcinomas in rats by dextran sulfate sodium. *Carcinogenesis (London).* 1982;3(3):353-355.
66. Tanaka T, Kohno H, Suzuki R, Yamada Y, Sugie S, Mori H. A novel inflammation-related mouse colon carcinogenesis model induced by azoxymethane and dextran sodium sulfate. *Cancer Sci.* 2003;94(11):965-973.
67. Suzuki R, Kohno H, Sugie S, Nakagama H, Tanaka T. Strain differences in the susceptibility to azoxymethane and dextran sodium sulfate-induced colon carcinogenesis in mice. *Carcinogenesis.* 2006;27(1):162-169.
68. Takeuchi A, Kamiryu Y, Yamada H, et al. Oral administration of xanthan gum enhances antitumor activity through Toll-like receptor 4. *Int Immunopharmacol.* 2009;9(13-14): 1562-1567.
69. Kimura Y, Sumiyoshi M, Suzuki T, Sakanaka M. Antitumor and antimetastatic activity of a novel water-soluble low molecular

- weight β -1, 3-D-glucan (branch β -1,6) isolated from *Aureobasidium pullulans* IA1 strain black yeast. *Anticancer Res.* 2006; 26(6B):4131-4142.
70. Yoo SH, Yoon EJ, Cha J, Lee HG. Antitumor activity of levan polysaccharides from selected microorganisms. *Int J Biol Macromol.* 2004;34(1-2):37-41.
71. Paull JD. Dextran. *Dev Biol Stand.* 1987;67:133-138.
72. Ott CM, Day DF. *Modification of Natural Gums.* Ipswich, MA: EBSCO Publishing; 2000:186-228.
73. Li H, Xu H, Li S, Ying HJ, Ouyang PK. Enhanced welan gum production using a two-stage agitation speed control strategy in *Alcaligenes* sp. CGMCC2428. *Bioprocess Biosyst Eng.* 2011; 34(1):95-102.
74. Manjanna KM, Kumar TMP, Shivakumar B. Natural polysaccharide hydrogels as novel excipients for modified drug delivery systems: a review. *Int J ChemTech Res.* 2010;2(1):509-525.
75. Bresin A, Sanhajl G, Reynaud R. Rhizobium gum: a novel cosmetic ingredient from soil to the skin. *Cosmet Toiletries Mag.* 2004;19(4):86-92.
76. Mehvar R. Dextran for targeted and sustained delivery of therapeutic and imaging agents. *J Contr Release.* 2000;69(1):1-25.
77. Miyake M, Kakizawa Y. Morphological study of cationic polymer-anionic surfactant complex precipitated in solution during the dilution process. *J Cosmet Sci.* 2010;61(4):289-301.
78. Park SY, Bae IY, Lee S, Lee HG. Physicochemical and hypocholesterolemic characterization of oxidized oat β -Glucan. *J Agric Food Chem.* 2009;57(2):439-443.
79. Ding XL, Wang M. Development of a water-soluble α -carboxymethyl- β -(1,3)-glucan derived from *Saccharomyces Cerevisiae*. 2008:412-419.
80. Mibelle Biochemistry. 2010 CM-Glucan granulate specifications. 2010. Unpublished data submitted by Personal Care Products Council.
81. Kaplan DL, Wiley BJ, Mayer JM, et al. Biosynthetic polysaccharides. In: Shalaby SW, ed. *Biomedical Polymers: Designed-to-Degrade Systems.* New York, NY: Hanser Publishers; 1994: 189-212.
82. Combie J. [Chapter 13:] Properties of levan and potential medical uses. In: Marchessault RH, Ravenelle F, Zhu X, eds. *ACS symposium series 934. Polysaccharides for Drug Delivery and Pharmaceutical Applications.* Washington, DC: American Chemical Society; 2006:263-269.
83. Nohata Y, Azuma J, Kurane R. Structural studies of a neutral polysaccharide produced by *Alcaligenes latus*. *Carbohydr Res.* 1996;293(2):213-222.
84. Jungbunzlauer. Safety Data Sheet: Xanthan Gum. 2009. Unpublished data submitted by Personal Care Products Council.
85. Green R. Letter from Biopolymer International to the European Commission concerning molecular weight of polymer additives. 2010. Unpublished data submitted by Personal Care Products Council.
86. Bajaj IB, Survase SA, Saudagar PS, Singhal RS. Gellan gum: fermentative production, downstream processing and applications. *Food Technol Biotechnol.* 2007;45(4):341-354.
87. Food Chemicals Codex. Rockville, MD: The United States Pharmacopeial Convention; 2008.
88. Chandrasekaran R, Radha A, Lee EJ. Structural roles of calcium ions and side chains in welan: an X-ray study. *Carbohydr Res.* 1994;252(93):183-207.
89. Solabia. Active Ingredients Catalog [pamphlet]. 2011.
90. The United States Pharmacopeia, 32nd Revision/The National Formulary, 27th edition. Rockville, MD: The United States Pharmacopeial Convention; 2009.
91. ADEKA Corporation. Technical Information. Fermented Beta-Glucan (LQ/Powder) [pamphlet]. 2009.
92. Cross GG, Jennings HJ, Whitfield DM, et al. Immunostimulant oxidized β -glucan conjugates. *Int Immunopharmacol.* 2001; 1(3):539-550.
93. Klecak J. Summary report CM-Glucan (Sodium Carboxymethyl Betaglucan) in skin care products. 1998. RCC Project 678082. Unpublished data submitted by Personal Care Products Council.
94. Hayashibara International Inc. Pullulan GRAS Notification. Web site. http://www.accessdata.fda.gov/scripts/fcn/gras_notices/215492E.PDF. 2002. Accessed November 6, 2011.
95. Nakamura S. Pullulan. *J Synthetic Organ Chem Jpn.* 1984;43(6): 584-588.
96. Leathers TD. Biotechnological production and applications of pullulan. *Appl Microbiol Biotechnol.* 2003;62(5-6):468-473.
97. Bozou JC, Gautry L, Pianelli G. A new biopolymer for refreshment. *SOFW J.* 2004;130(3):17-21.
98. Food Chemicals Codex. 8 ed. Rockville, MD: United States Pharmacopeia (USP); 2012.
99. Biopolymer International. Xanthan gum—Manufacturing Process. 2010. Web site. <http://www.biopolymer-international.com/html/manufacturing.php>. Accessed August 31, 2011.
100. Hijiya H, Shiosaka M, inventors. Shaped bodies of pullulan esters and their use. US patent 3871892. March 18, 1975.
101. Crompin JM, Garnier T, Payot T, DeBaynast R. A new polysaccharide derived from plant rhizosphere: production, purification and physico-chemical properties. *Eng Manuf Biotechnol.* 2002;4(8):423-428.
102. Biopolymer International. Xanthan gum. Product description. 2010. Web site. http://www.biopolymer-international.com/html/xanthan_gum.php. Accessed August 30, 2011.
103. Biopolymer International. Gellan Gum—Product description. 2010. Web site. http://www.biopolymer-international.com/html/gellan_gum.php. Accessed August 30, 2011.
104. Booth AN, Hendrickson AP, DeEds F. Physiologic effects of three microbial polysaccharides on rats. *Toxicol Appl Pharmacol.* 1963;5:478-484.
105. McNeely WH, Kovacs P. The physiological effects of alginate in xanthum gum. In: [American Chemical Society] *Symposium Series 15.* Washington, DC: The Society; 1975:263-269.
106. Babicek K, Cechova I, Simon RR, Harwood M, Cox DJ. Toxicological assessment of a particulate yeast (1,3/1,6)- β -D-glucan in rats. *Food Chem Toxicol.* 2007;45(9):1719-1730.
107. Life Science Laboratory. Single oral dose toxicity study of CM-Glucan J in mice (at a dose level of 2000 mg/kg) . 1999. Test Code No. 99-IA1-1004. Unpublished data submitted by Personal Care Products Council.
108. Juntendo University, Department of Public Hygiene, School of Medicine. Report on acute toxicity test on pullulan with